

## Evaluation of the Antioxidant Properties and Aromatic Profile During Maturation of The Blackberry (*Rubus glaucus* Benth) and The Bilberry (*Vaccinium meridionale* Swartz)

Evaluación de las Propiedades Antioxidantes y el Perfil Aromático Durante la Maduración de Mora (*Rubus glaucus* Benth) y Agraz (*Vaccinium meridionale* Swartz)

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**Abstract.** The blackberry (*Rubus glaucus* Benth) and the bilberry (*Vaccinium meridionale* Swartz) are natural sources of antioxidants; they are known for their preventive role against degenerative diseases. In this study, the aromatic profile was evaluated using an electronic nose, including the antioxidant properties and the vitamin C, phenolic and anthocyanin contents during three stages of blackberry and bilberry ripening. A completely random statistical design was followed and the results presented differences in the aromatic profile: a higher anthocyanin content (1.59 mg of cyn-3-glu g<sup>-1</sup> in the bilberry and 0.26 mg of cyn-3-glu g<sup>-1</sup> in the blackberry) and total phenols (5.57 mg of caffeic acid g<sup>-1</sup> bilberry and 2.68 mg caffeic acid g<sup>-1</sup> blackberry). The behavior of the evaluated properties was independent in each of the fruits.

**Keywords:** Antioxidant activity, electronic nose, Folin-Ciocalteu, FRAP, TEAC.

**Resumen.** Los frutos como la mora (*Rubus glaucus* Benth) y el agraz (*Vaccinium meridionale* Swartz) son fuentes naturales de sustancias antioxidantes reconocidas por su papel preventivo en el desarrollo de enfermedades degenerativas. En este estudio se evaluó el perfil aromático por medio de nariz electrónica, las propiedades antioxidantes y el contenido de vitamina C, fenoles y antocianinas totales, durante tres estados de maduración de mora y agraz. El diseño estadístico que se siguió fue completamente aleatorio y los resultados muestran que las frutas en el último estado de madurez evaluado se diferencian por su perfil aromático, un contenido mayor de antocianinas (1,59 y 0,26 mg cyn-3-glu g<sup>-1</sup> en agraz y mora, respectivamente) y fenoles totales (5,57 y 2,68 mg ácido caféico g<sup>-1</sup> en agraz y mora, respectivamente). El comportamiento de las propiedades evaluadas es independiente en cada una de las frutas.

**Palabras clave:** Actividad antioxidante, nariz electrónica, Folin-Ciocalteu, FRAP, TEAC.

The blackberry (*Rubus glaucus* Benth) is a fruit composed of drupes and is characterized by its red-blue color and its aroma. It is a source of vitamins, minerals and phytochemicals. In Colombia, the most cultivated species is known as "mora de Castilla" (*Rubus glaucus* Benth); harvest is constant during the whole year and is only influenced by the rainy season. For cultivation, altitudes between 1,800 and 2,000 masl are preferred. The climacteric character of this fruit is unclear and, additionally, it is a product that is difficult to handle due to its high perishability and sensitivity to mechanical damage (Perkins-Veazie *et al.*, 2000).

The bilberry (*Vaccinium meridionale* Swartz), also known as "mortiño", is a small shrub that has two harvests per year. The fruit is fleshy with numerous seeds, has a sour taste, and is characterized by a dark purple color when maturity is reached. It is found between 2,200 and 3,400 masl, with the Colombian regions of Boyacá and Antioquia being the areas

where domestic production is focused (Ligarreto, 2011). It belongs to the same genus as blueberries and the climacteric fruits are harvested in the state of sensory maturity, which ensures the aroma and flavor characteristics that are desirable by consumers (Mitcham *et al.*, 1998).

These red fruits are a natural source of antioxidants, with a predominant group of phenolic acids and flavonoids, the latter being responsible for the color (Pietta *et al.*, 2003; Castrejón *et al.*, 2008). Antioxidants retard oxidation reactions, key deterioration reactions that result from the relocation of the aromatic rings that compose them (Balasundram *et al.*, 2006). In addition, recent studies have shown the importance of preventing degenerative diseases such as Alzheimer's and Parkinson (Wang and Stoner, 2008). It is a fact that the content of phenolic compounds in berries is affected by genetic differences, pre- and post-harvest conditions, and, also, the degree of maturity.

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In fruits, ripening control is important at the time of establishing the point of harvest, storage conditions and the evolution of the sensory characteristics. During this process, the biochemical changes undergone by the fruits give way to the generation of volatiles and changes in color, texture and flavor. A nondestructive method that provides results in real time is the electronic nose, an instrument that emulates the human olfactory process and that is comprised of a system that is sensitive to volatiles and another that is in charge of converting responses into electrical signals. Applications in this area have been numerous: peaches (Benedetti *et al.*, 2008), pears (Brezmes *et al.*, 2000), and apples (Pathange *et al.*, 2006) have been analyzed and distinguished in different stages of ripening, postharvest operations (Torri *et al.*, 2010) and fermentations (Bhattacharyya *et al.*, 2007). In fruits such as berries, the available information is limited (Peris and Escuder-Gilabert, 2009); however, there is a report from Li *et al.*, (2010), who employed bilberries for disease detection.

In Colombia, the blackberry and bilberry have gained commercial positioning among consumers as a food and a source of functional substances. So far, no studies have been conducted to evaluate changes in antioxidant properties and the aromatic profile during the ripening stage. This study aimed to evaluate changes in the antioxidant capacity by using spectrophotometric techniques and in the aromatic profile by using an instrumental measure of aroma in three ripening stages (green, intermediate and harvesting).

## MATERIALS AND METHODS

**Samples.** The blackberries were collected from a crop located in the village of El Triunfo, Icononzo-Tolima and the bilberries were collected from wild bushes in the village of Arrayanes, Tinjacá - Boyacá. Three stages of ripening were selected that were associated with a subjective color indicator: green (G), where the fruits had developed their size and the surfaces had a green color with small yellow and pink areas; intermediate (I), characterized by a pink color with small yellow drupes on the surface of the blackberries and a red surface in the bilberries; and harvesting (H), the latter corresponded to the state of commercial maturity where the blackberries had developed a deep red color and the bilberries a purple tone. For every physicochemical analysis, a representative sample was taken for use in the corresponding procedure.

**Physical and chemical analyses.** For analyses of fruit characterization, official methods were employed:

**Moisture.** 10 g of sample were dried at 105 °C (ThermoHeraus™ Instruments Function Line Oven) for 4 hours. A.O.A.C. Official Method 934.06; A.O.A.C. Official Method 934.01. This measurement was performed in triplicate.

**Total soluble solids (TSS).** The content of °Brix was determined by using a refractometer according to Colombian Technical Standard NTC 4624. This measurement was performed in triplicate.

**pH and acidity.** A potentiometric titration was performed (SCHOTT Handy labpH11) according to the method described in the Colombian Technical Standard NTC 440 and A.O.A.C. Official Method 942.15.

**Aromatic profile determination.** An Airsense Analytics™ GmbH PEN3 Electronic Nose was employed with 10 semi-conductor metal oxide sensors - MOS (Table 1). 25 g of sample were weighed and kept in a plastic chamber with a hermetic seal that measured 11.0 x 10.6 x 10.6 cm and a cover that measured 3.8 x 11.2 x 11.2 cm at room temperature (15 °C approx.) for 5 min to stabilize the content of volatile compounds. The aromatic profile was evaluated in cycles of 150 s with an air chamber flow of 70 mL min<sup>-1</sup> and a flow injection of 60 mL min<sup>-1</sup>. The sample analysis was performed randomly, leaving a time of 450 s for cleaning the sensors between each cycle of analysis in order to avoid the interference and undesirable effects on the responses resulting from the dragging of volatile residues from the previous sample (Zuluaga *et al.*, 2011).

**Vitamin C.** The extraction was carried out with distilled water; before injection, the solution was purified with a C<sub>18</sub> cleaning cartridge. For the detection and quantification, HPLC chromatography was employed (Jasco™ Pump PU980, Detector UV/VIS975) with a Phenomenex Rezex™ ROA-Organic Acid H+ 8% ion exchange column, at a flow of 0.5 mL/min and with a mobile phase of H<sub>2</sub>SO<sub>4</sub> 4 mM at room temperature. The employed wavelength was 254 nm. The data were reported as µmol of vitamin C g<sup>-1</sup> of fresh fruit (Shui and Leong, 2002; Castañeda *et al.*, 2009). 20 g of fruit were prepared and homogenized (Ultra-Turrax™ IKA T18 Basic), 20 mL of extraction solution were added and, then, vortex agitation and heating in

**Table 1.** Chemical compounds associated with the sensor array of the Airsense Analytics GmbH PEN3 electronic nose.

Sensor	Detected chemical compounds
W1C	Aromatic compounds
W5S	Wide range of compounds
W3C	Aromatic compounds. Ammonia
W6S	Mainly Hydrogen
W5C	Aromatic Compounds. Alkanes
W1S	Wide range. Methane
W1W	Sulfur compounds. Terpenes
W2S	Alcohol detection. Wide range
W2W	Aromatic compounds. Organic compounds
W3S	Methane

a magnetic stirring plate were carried out. The mixture was centrifuged for 10 min (Tecnovetro™ 4235); this procedure was repeated twice with the solid. The extracts were brought to a volume of 100 mL by using the extraction solution. Each sample was performed in triplicate and stored at -18 °C until analysis.

**Total anthocyanin content.** The differential pH method was employed. A.O.A.C. Official Method 2005.02. Results were reported as mg of cyaniding-3-glycoside (cyn-3-glu) g<sup>-1</sup> of fresh fruit.

**Total phenolic content.** The Folin-Ciocalteu reagent was used (Panreac, Spain). This is a method described by Vasco *et al.* (2008) and Oliveira *et al.* (2011). The wavelength at which the reading was taken was 765 nm and the quantification was done by reference to a calibration curve of caffeic acid, Sigma Aldrich, (R<sup>2</sup> = 0.995). The data were reported as mg of caffeic acid g<sup>-1</sup> of fresh fruit.

**Antioxidant capacity.** In TEAC, radical discoloration was read at 734 nm after a 6min reaction. Each extract was read in triplicate and the results were reported as μmol trolox g<sup>-1</sup> of fresh fruit with respect to a trolox 97% calibration curve, Sigma Aldrich™, (R<sup>2</sup> = 0.989) (Murcia *et al.*, 2009).

In FRAP, 330 μL of extract were mixed with 10 mL of a solution made from buffer acetate pH 3.6 (300 nM), TPTZ diluted in HCl (40 mM) and FeCl<sub>3</sub> (20 mM) at a 10:1:1 ratio, respectively. The reaction was held for an hour and the reading was taken at 593 nm. Each extract was read in duplicate and the results were reported according to the Trolox calibration curve (R<sup>2</sup> = 0.996) in mmol trolox g<sup>-1</sup> of fresh fruit (Gorinstein *et al.*, 2009; Müller *et al.*, 2010).

**Statistical analysis.** The data were analyzed using one-way analysis of variance ANOVA, in conjunction with a Tukey-Kramer's multiple comparison test, using an alpha of 0.05. In addition, the principal component analysis-PCA was performed as a multivariate technique for exploration of the data obtained with the electronic nose. The statistical analyses were carried out with the MATLAB software, v. 7.9 (MathWorks™).

## RESULTS AND DISCUSSION

The "mora de Castilla" is a fruit whose maturity is determined by changes in color, size, percentage of acidity and soluble solids content, according to NTC 4106. For this fruit, seven stages of maturation can be differentiated. The ° Brix of the samples evaluated correspond to the zero stage, a state corresponding to green; stage three: an intermediate state; and stage five: the harvest state. It could be observed (Table 2) that, as the fruits matured, humidity increased and the percentage of acidity decreased. No pH changes were noticeable in these states.

In the bilberry, in addition to the obvious changes of color, parameters such as pH and concentration of soluble solids were indicators of fruit maturation (Table 3). The study of the bilberry is gaining strength given its wild characteristic. Ávila *et al.* (2007) reported, for the harvesting stage, acidity values close to 1.44%. The behavior of this fruit is similar to that of the cranberry, which belongs to the same genus (Castrejón *et al.*, 2008).

**Aromatic profile.** The Release of aromatic compounds, together with the loss of the green color and acid

**Table 2.** Physical-chemical behavior of the blackberry during ripening.

Ripening stage <sup>†</sup>	Moisture (%)	pH	Acidity (% malic acid)	Total soluble solids (°Brix)
G	81.96±1.69 a	2.88±2.83 e <sup>-2</sup> a	2.53±4.19 e <sup>-1</sup> ab	5.49±8.13 e <sup>-2</sup> a
I	87.30±3.40 e <sup>-1</sup> ab	2.76±9.90 e <sup>-2</sup> a	3.09±5.53 e <sup>-2</sup> a	6.60±1.07 e <sup>-2</sup> b
H	88.34±1.89 b	2.92±1.48 e <sup>-1</sup> a	1.86±6.04 e <sup>-2</sup> b	7.36±1.17 e <sup>-2</sup> c

<sup>†</sup>G: green, I: intermediate, H: harvesting. Mean ± SD; SD = Standard deviation. Means with different letters in a same row show significant differences; \*significance level P<0.05.

flavor, is one of the changes that occur during the ripening of any fruit.

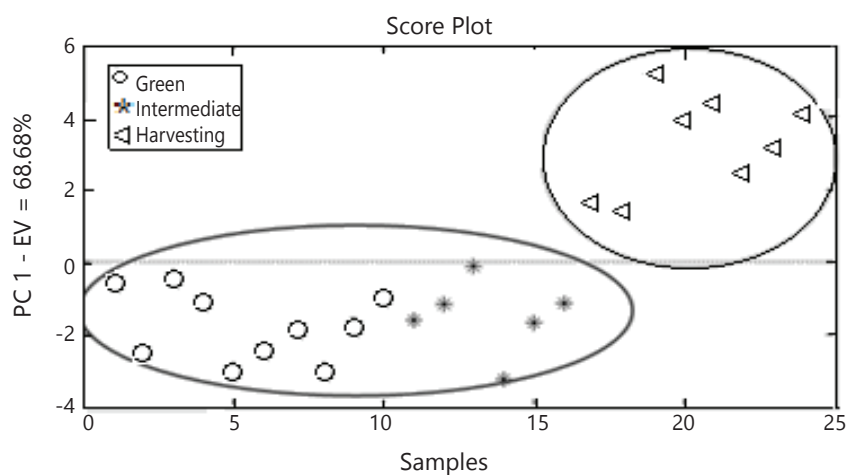
The principal component analysis for the blackberry (Figures 1 and 2) showed that the green and intermediate maturation stages are distinguished from the harvesting stage by the response of a group of sensors that perceive the presence of aromatic functional groups of low polarity: W1C, W5S, W3C and W5C (Table 1). In the harvesting

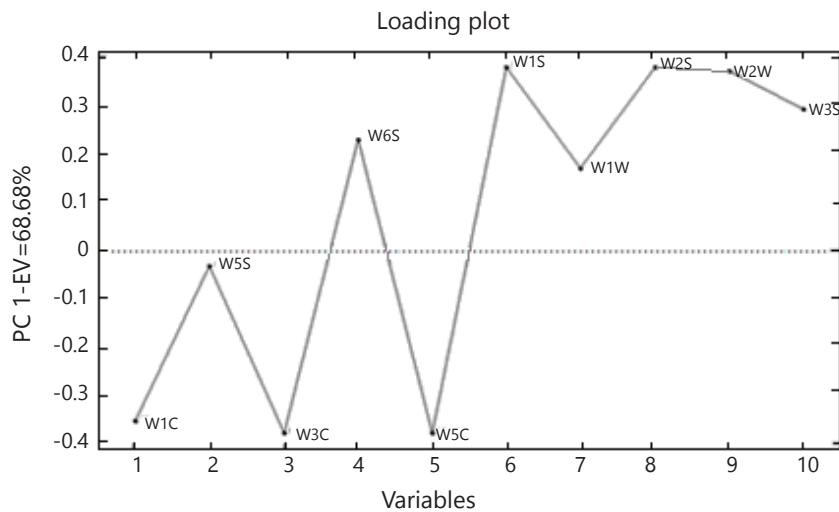
stage, the presence of terpenes, alcohols, and aromatic compounds such as methanol can be considered because the group of sensors that shows responses includes: W6S, W1S, W2S, W3S, W1W and W2W. These results agree with the findings of a study conducted with volatiles of ready-to-eat blackberries, where the presence of benzoic acid, 2-heptanol, terpene-4-ol, ethyl and methyl benzoate was reported (Meret *et al.*, 2011); also, in the *Rubus* sp. genus, during maturation, the production of

**Table 3.** Physical-chemical behavior of the bilberry during ripening.

Ripening stage <sup>†</sup>	Moisture (%)	pH	Acidity (% Malic acid)	Total soluble solids (°Brix)
G	84.13±3.27 e <sup>-1</sup> a	2.99±0.0 a	1.84±2.71 e <sup>-4</sup> a	7.2±7.07 e <sup>-2</sup> a
I	83.73±6.35 e <sup>-1</sup> a	2.96±7.07 e <sup>-3</sup> b	1.85±1.33 e <sup>-2</sup> a	9.6±7.07 e <sup>-2</sup> b
H	83.34±6.53 e <sup>-1</sup> b	3.05±7.07 e <sup>-3</sup> c	1.72±1.10 e <sup>-1</sup> a	9.9±1.41 e <sup>-1</sup> b

<sup>†</sup>G: green, I: intermediate, H: harvesting. Mean ± SD; SD = Standard deviation. Means with different letters in a same column show significant differences; \*significance level P<0.05.

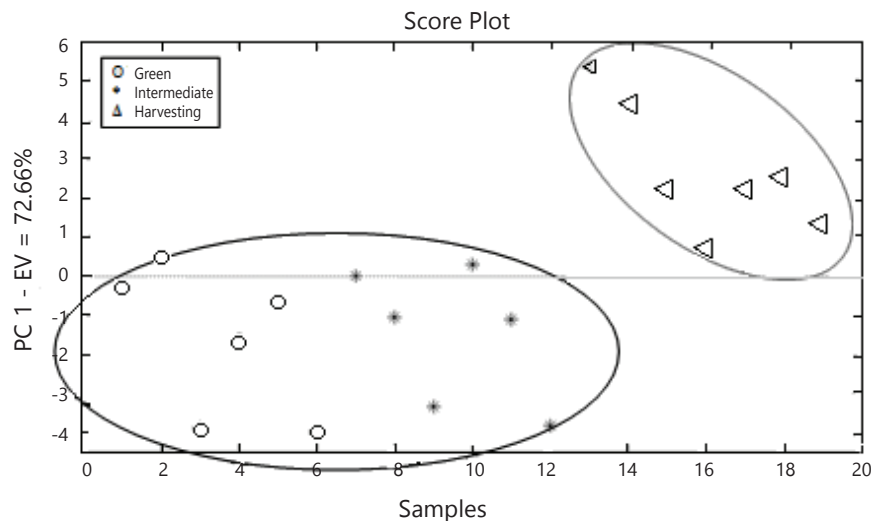
**Figure 1.** Score plot of the blackberry samples, calculated PCA explains 68.68% of the variance.



**Figure 2.** Loading plot of the blackberry samples, each variable represents an E-nose sensor.

alcohols, aldehydes, esters and ketones is favored (Perkins-Vezzie *et al.*, 2000). The bilberry (Figures 3 and 4) exhibited the same behavior as that of the blackberry; fruits in the harvesting stage can be differentiated from the other two stages by sensors

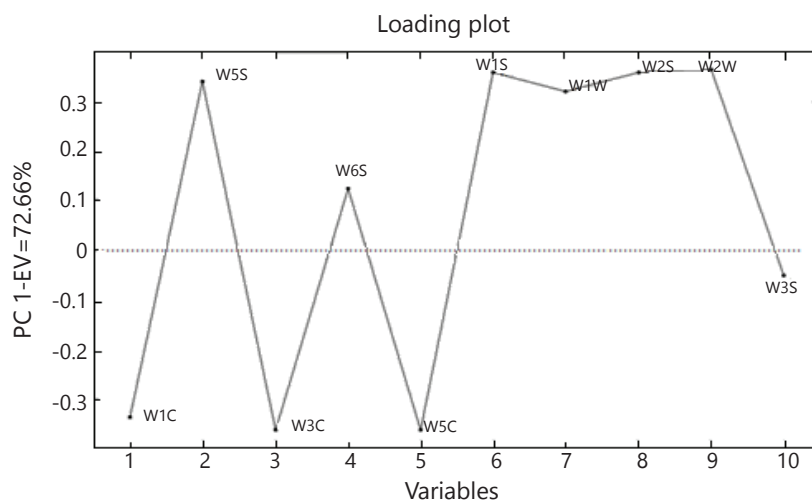
that respond to volatile aromatic functional groups, alcohols and terpenes. Studies on the *Vaccinium* sp. genus report the presence of 1-hexanol, 2-hexen-1-ol, 2-hexenal, 2-butyl-1-octanol, hexadecanol, etc. (Diban, 2008).



**Figure 3.** Score plot of the bilberry samples, calculated PCA explains 72.66% of the variance.

Both score plots explain more than 50% of the variance of the groups in each case. In the loading plots, a difference between the aromatic profile of the blackberry and the bilberry is seen due to different responses from the group of sensors that actuates during ripening.

**Vitamin C content.** Vitamin C has biological activity as an antioxidant because the double bond and substitutions have two hydroxyl groups within the chemical structure, is synthesized in plants in response to stress conditions, and, additionally, its concentration is determined by genetic factors (Asard *et al.*, 2004).



**Figure 4.** Loading plot of the bilberry samples, each variable represents an E-nose sensor.

The evaluated samples contained between 13.46 and 22.48  $\mu\text{g}$  of vitamin C  $\text{g}^{-1}$  of blackberries and 23.09 and 25.06  $\mu\text{g}$  of vitamin C  $\text{g}^{-1}$  of bilberries (Table 4); there were no significant differences in either case, but the p value of the blackberries was close to the alpha value ( $P=0.057$ ), suggesting that, for one of the stages of maturity, the content is different, statistical

analysis according to the value found corresponds to the first stage of maturation, where it presented high variability. Contrary to expectations, the vitamin C content of the evaluated blackberry sample increased from state zero to state three. This could be explained by the non-climacteric behavior and the variability in agricultural practices and environmental conditions.

**Table 4.** Antioxidant properties of the blackberry and the bilberry during ripening.

	Vitamin C $\mu\text{g g}^{-1}$	Total anthocyanins $\text{mg g}^{-1}$	Total phenolics $\text{mg g}^{-1}$	TEAC $\mu\text{mol trolox g}^{-1}$	FRAP $\text{mmol trolox g}^{-1}$
<b>Ripening state of the blackberry</b>					
G	13.46 $\pm$ 2.07 a	1.89e-2 $\pm$ 9.03e-3 a	3.33 $\pm$ 7.30e-2 a	33.29 $\pm$ 5.56a	42.37 $\pm$ 1.42 a
I	21.99 $\pm$ 7.61e-1 a	5.67e-2 $\pm$ 2.69e-2 a	2.92 $\pm$ 2.79e-1 b	32.43 $\pm$ 7.24a	37.11 $\pm$ 5.55 b
H	22.48 $\pm$ 3.60 a	2.57e-1 $\pm$ 4.35e-2 b	2.68 $\pm$ 2.48e-1 b	30.22 $\pm$ 9.42a	17.30 $\pm$ 0.71 c
<b>Ripening state of the bilberry</b>					
G	24.97 $\pm$ 7.99e-1 a	3.49e-2 $\pm$ 7.72e-3 a	4.69 $\pm$ 7.57e-2 a	30.71 $\pm$ 2.96a	39.70 $\pm$ 5.00 a
I	25.06 $\pm$ 3.20 a	3.21e-1 $\pm$ 2.27e-2 b	4.74 $\pm$ 1.01e-1 a	34.81 $\pm$ 1.24b	39.52 $\pm$ 1.28 a
H	23.09 $\pm$ 2.53e-1 a	1.59 $\pm$ 9.34e-2 c	5.57 $\pm$ 2.67e-1 b	44.40 $\pm$ 3.33c	54.45 $\pm$ 3.56 b

G: Green, I: Intermediate, H: Harvesting. Mean  $\pm$  SD; SD = Standard deviation. Means with different letters in a same column show significant differences; \*significance level  $p<0.05$

It has been reported that there is an accumulation of vitamin C and phenolic compounds (Cocetta *et al.*, 2012) in the *Vaccinium* sp genus. during ripening, a behavior corresponding to that found in the bilberry. Rodriguez *et al.* (2007) reported a higher content of vitamin C for this fruit in the

harvesting stage, about 80  $\mu\text{g}$  of vitamin C  $\text{g}^{-1}$  of fresh fruit.

The vitamin C content of the blackberry and the bilberry was considerably lower than that reported in the composition table of the USDA (2010); for the



blackberry: 209.72  $\mu\text{g}$  of vitamin C  $\text{g}^{-1}$  of fresh fruit and for the blueberry: 97.24  $\mu\text{g}$  of vitamin C  $\text{g}^{-1}$  of fresh fruit. On the other hand, some Colombian fruits have a vitamin C content between 0.53 and 257  $\text{mg}$   $100 \text{g}^{-1}$  of fresh fruit (Contreras *et al.*, 2011); the present data are outside this range, so it is important to note the differences that exist between families and the ecophysiological conditions of fruits, which determine composition and affect behavior patterns.

**Anthocyanins content.** The blackberry and the blueberry are sources of anthocyanins, the most important flavonoids responsible for attracting insects to plants and protecting them from various diseases and predators (Brown *et al.*, 2006; Chemler *et al.*, 2009). In general, the anthocyanin structure is linked to a sugar and can even be copolymerized, which affects the availability and quantification (AOAC, 2005) Different authors report that predominate anthocyanin in blackberry and blueberry is the cyn-3-glu (Dai *et al.*, 2009; Garzón *et al.*, 2010).

During maturation, the increased anthocyanin content, in this way is synthesized from shikimic acid and malonic acid. (Chemler *et al.*, 2009). In the case of the blackberry, there are no significant differences between the green and intermediate stages but an increase of nearly 14 times more than the initial value has been observed in the harvesting stage. This is comparable to that described by Chen *et al.* (2012) for the early stages of ripening, reporting a value of 0.106  $\text{mg}$  cyn-3-glu  $\text{g}^{-1}$  of fresh fruit and, for the subsequent maximum peak in the maturing state, 1.46  $\text{mg}$  cyn-3-glu  $\text{g}^{-1}$  of fresh fruit. The bilberry presents significantly different contents in the three stages, with the harvesting stage having 45 times more anthocyanins than the green stage.

The contents of anthocyanins found in the blackberry and the bilberry allow for the comparison of the blackberry stage of harvesting with the intermediate stage of the bilberry, leading to the conclusion that, among mature fruits, the bilberry is an important source of these bioactive compounds.

**Total phenolic content.** The concentrations of phenolic compounds and nutrients, vitamins, and minerals, among others, respond to multiple environmental and genetic factors. The composition of a food is never constant and it is not uncommon to find differences between fruits, even in the same crop. For example, it has been found that the leaves and fruits that are in direct contact with solar radiation have increased contents of phenolic

compounds, vitamin C and carotenoids, in comparison to those found in the shade (Brown *et al.*, 2006; Vicente *et al.*, 2009). The total phenolic content in the case of the bilberry, at any stage of maturity, was greater than that of the blackberry and, while polyphenols increased in the bilberry, in the blackberry, they decreased. The variety of compounds present in each fruit may influence the final result, especially considering that the Folin-Ciocalteu method may not necessarily respond to polyphenolic substances such as aromatic amines or even ascorbic acid (Magalhães *et al.*, 2008).

**Antioxidant capacity.** The antioxidant power of a fruit can be measured by different *in vitro* methods, which give an idea of the ability of the sample to scavenge free radicals (Niki, 2010). Non-competitive methods employing ABTS $^{\bullet+}$  and the complex  $[\text{Fe}(\text{II})(\text{TPTZ})_2]^{+2}$  introduce a radical in the sample and quantify how much was inhibited by the substances present. During ripening, the bilberry increased its antioxidant capacity (Table 4), whereas the blackberry presented no evidence of significant changes (such as TEAC), similar to the findings of Livani *et al.* (2013), who studied two ripening stages of *Rubus anatolicus* with the DPPH radical. However, the FRAP data of the blackberry showed that the stage of harvesting had a lower antioxidant capacity than the green stage. Acosta-Montoya *et al.* (2011) evaluated samples of *Rubus adenotrichus* Schltdl with H-ORAC during ripening, finding that the antioxidant capacity increased while the total phenol content remained unchanged; these results show the differences that can be found among the same family and that might be associated with genetic characteristics or crop and postharvest conditions.

## CONCLUSIONS

The behavior of the antioxidant properties during ripening in the blackberry and the bilberry are independent. In the blackberry, there was a decrease but, in the bilberry, an increase was evidenced, a better characteristics that makes the latter fruit an attractive food. The electronic nose allowed for the formation of the aromatic profile in different stages of maturity, revealing differences in the harvesting stage for both fruits.

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