

**Studies on certain aspects of root surface fungi
IV. Succession of fungi on *Pennisetum typhoides*
in fertilized soils***

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The mycoflora and its succession on decomposing roots of *P. typhoides* of plots fertilized with urea, ammonium sulphate, superphosphate, organic manure, fresh leaves and twigs of *Ipomoea fistulosa* and irrigated has been investigated. Fungi were higher on fertilized plots, both a root surface and away from it (the control). Highest and lowest population were recorded soils fertilized with organic manure and *I. fistulosa*. The amino acids and sugars, cellulose, hemicellulose and lignin components of the roots from fertilized and control plots has been estimated and a correlation between mycoflora succession and the above components has been established.

INTRODUCTION

Waksman and Starkey (1924), Zacharias (1949), Kaufman and Williams (1964) and Kanauija (1977) reported that the addition of various chemical fertilizers the soil improved fungal growth. The effect of fertilization on the rhizosphere mycoflora of various crop plant also been studied (Bagyaraj and Rangaswami 1967; Mishra 1971a, b, 1972; Maurer and Baker 1965; Naim and Shaaban 1967). However, the effect of fertilization on the succession of fungi on recomposing underground plant parts is least understood (K-

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naaujia 1973) and this forms the bulk of the present study. In this study the effect of urea, ammonium sulphate, superphosphate, organic manure, fresh leaves and twigs of *Ipomoea fistulosa* and irrigation practice on the fungi and their succession on the decomposing *Pennisetum typhoides* (Burm f.) Stapf Hubb. roots has been examined.

MATERIALS AND METHODS

To investigate the effect of various fertilizers, organic manure, plant parts on the succession pattern of fungi on decomposing roots of *P. typhoides*, the experimental plot where the crop was raised was ploughed over after removing the above-ground plant parts in January 1971 and divided into seven equal parts. Urea (U), ammonium sulphate (AS), superphosphate (SP), organic manure (OM) and fresh leaves and twigs of *Ipomoea fistulosa* (IF) were added separately at the rate of 60 kg/acre on January 1, 1971. One part of the plot was regularly irrigated fortnightly (IR) by a spraying method to bring its moisture content to of $25 \pm 1\%$ and other was not treated (control). The first sampling of the roots and soil was made January 25, 1971 and subsequent ones were done once in a month till the total decomposition of the roots. The mycoflora associated with root (RS — rhizosphere, RPC — cortical and RPS — stelar part of rhizoplane) and away from it (NR) was investigated by the method adopted by Kanaujia (1973). The amino acid and sugar contents of root on various sampling dates was determined by paper chromatography and calorimetry as suggested by Smith (1960) and Peach and Tracey (1955) and was expressed as $\mu\text{g}/2\text{ g}$ fresh roots. The cellulose, hemicellulose and lignin components were estimated on the basis of initial dry weight of roots (Wise et al. 1945). The pH and moisture content of the plots was determined every time using the methods described by Piper (1966).

RESULTS

Mycoflora of amended plots

Forty-three, 37, 36, 39, 42, 30 and 37 fungal species were isolated from U, AS, SP, OM, I, IF amended and control sets respectively. Eighteen species were commonly isolated from different root regions of all the sets whereas 8 species were specifically present in NR of different sets (Table 1). *Cladosporium epiphyllum* (Pers.) Mart. and *C. herbarum* (Pers.) Link were quantitatively dominant in all the sets

Table 1

Distribution of fungi of different taxonomic groups in non-rhizosphere (NR), rhizosphere (RS), and rhizoplane (RPC cortical and RPS steler) regions of fertilized plots

Amendments		Taxonomic fungal groups									Total No. of spp.	
		Phycomycetes		Ascomycetes	Deuteromycetes							colony Sterile
		Rhizopus spp.	Others		Chaetomium spp.	Aspergillus spp.	Penicillium spp.	Cladosporium spp.	Fusarium spp.	Others		
Urea	NR	2	3		9	3	2	2	7	4	32	
	RS	2	2		8	3	2	3	4	1	25	
	RPC		2	1	5	2	2	2	2	2	18	
	RPS		2		3	1	2	2	1	2	13	
Ammonium sulphate	NR	2	1		9	4	2	2	3	2	25	
	RS	1	1		9	2	2	1	4	1	21	
	RPC		2		4	2	2	3	2	2	17	
	RPS		1		2	1	2	2	1	3	12	
Superphosphate	NR	2	2		11	3	2	2	5	3	30	
	RS	1	3		8	2	2	2	4	1	23	
	RPC		2	1	3	1	2	1	3	1	14	
	RPS		1		3		2	1	1	2	10	
Organic Manure	NR	2	3		10	2	3	2	4	4	30	
	RS	1	2		10	2	2	2	4	2	25	
	RPC		2		5	2	2	2	3	2	18	
	RPS		2		4	1	2	1	1	2	13	
Irrigated	NR	2	3	1	11	2	3	3	5	4	34	
	RS	2	3	1	9	1	2	3	4	2	27	
	RPC	1	2		6		2	3	3	2	19	
	RPS		1		4	1	2	3	3	2	16	
Ipomoea fistulosa	NR	1	3		6	3	3	1	4	2	23	
	RS	1	2		5	2	2	2		1	15	
	RPC		2	1	2	1	2	1	1	2	12	
	RPS		1	1	2	1	2	1	1	1	9	
Control	NR	2	1	1	6	4	2	2	4	3	25	
	RS	1	3		9	1	2	2	3	3	24	
	RPC		2		3	1	2	3	3	1	15	
	RPS		1		1	2	2	2	1	1	10	

during January to March. Other dominant fungi viz., *Zygorhynchus heterogamus* (Vuill.) Vuill. (AS, IF), *M. hiemalis* Wehm. (U, OM, I),

Rhizopus stolonifer (Ehr.) Vuill. (control), *Aspergillus flavus* Link (W, IF), *A. niger* v. Tiegh. (U, AS, SP, OM, IR and control), *Penicillium chrysogenum* Thom (SP, IF), *Fusarium nivale* (Fr.) Ces. (OM and control) and white sterile fungus — W1 (AS, SP and control) were obtained from plots indicated in parentheses. Out of 59 fungi isolated in the present study deuteromycetes and among them *Aspergilli* were in majority in

Table 2

Distribution pattern of number of fungal species in non-rhizosphere (NR), rhizosphere (RS) and rhizoplane (RPC cortical and RPS steller) regions of fertilized plots at different sampling periods

Amendments		Sampling period				
		Jan.	Febr.	March	April	May
Urea	NR	13	12	13	14	7
	RS	18	7	11	5	6
	RPC	5,7*	5,8*	6,6*	4,3*	7,3*
	RPS	4,4*	4,6*	4,4*	3,3*	5,2*
Ammonium sulphate	NR	7	8	13	13	7
	RS	17	6	9	6	
	RPC	13,8*	5,5*	5,8*	5,6*	
	RPS	3,3*	4,4*	3,3*	4,3*	
Superphosphate	NR	6	11	15	19	7
	RS	17	8	10	10	
	RPC	4,5*	6,7*	6,3*	6,3*	
	RPS	2	4,4*	3,4*	4,3*	
Organic Manure	NR	13	7	15	13	7
	RS	18	6	8	9	
	RPC	9,6*	5,6*	5,9*	6,3*	
	RPS	2,5*	4,6*	5,6*	4,3*	
Irrigated	NR	16	16	19	19	13
	RS	19	7	13	12	
	RPC	7,6*	6,7*	7,6*	10,6*	
	RPS	10,3*	5,7*	4,5*	6,6*	
Ipomoea fistulosa	NR	11	8	9	12	7
	RS	11	6	6	6	
	RPC	2,4*	5,4*	5,4*	3,3*	
	RPS	1,4*	5,4*	2,3*	3,3*	
Control	NR	8	9	13	13	7
	RS	16	7	10	11	
	RPC	8,7*	6,6*	5,4*	5,3*	
	RPS	3,3*	5,3*	3,4*	3,3*	

* Represents the average number of colonies/plate in the rhizoplane region.

all the plots. The details are given in Table 1. The number of fungal species in RS, RPC and RPS regions of root and NR in amended plots varied differently, however, a gradual decrease to the number of fungal species from NR, RS, RPC, RPS regions was obtained in all the cases. Highest and lowest number of species was obtained in irrigated sets and those fertilized with *Ipomoea fistulosa*, respectively (Table 1). A similar trend of species on different sampling periods was observed (Table 2). Species in U, SP and OM stets were nearly equal and the same was true with AS and control sets (Table 1).

The population in NR region of all the sets was highest in January and decreased gradually onwards till the end. It was highest in the OM and the lowest in the IF set. Population at the root-surface exhibited varying trends. It was maximum in urea, superphosphate and control sets during February. In remaining cases maximum and minimum population was obtained during March and April respectively. In the set fertilized with IF the population was comparatively low during January and February, however, it increased considerably afterwards (Fig. 1). The differences in the fungal population on fertilized plots as compared with control were statistically significant (Table 3).

Table 3

Analysis of variance for fungal population caused due to the supplements and age of the decomposition

Variation due to	SS	df	Variance	F Calculated	F Tabulated
Amendments	3179.04	6	529.84	3.00*	2.66 4.01
Age	507.69	3	169.23	0.958	3.16 5.09
Exp. error	3179.04	18	176.61		

* Significant at 1% level.

Amino acids and sugars in the roots on fertilized plots

A total of 15 amino acids (Table 4) was detected from the root extracts of all the sets. Alanine, arginine, glutamic acid, leucine, proline, threonine and valine were frequently present in the roots obtained from different plots whereas histidine and serine were present only in control and irrigated sets, respectively. Amino acids were nearly exhausted in April in the majority of sets (Table 4). The amount was generally highest in January and decreased in subsequent months (See

Table 4
Amino acids/sugars in the root extracts of amended plots

Sampling months 1971	Amino / Sugars															No.	Amount ug/22
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15		
January																	
U	75	-/+	-/+	-/+	-/+	90	60/+	40/+	170/+	-/+	150	88	45			6/4	618/450
AS	140/+	30/+	30/+	-/+	75/+	70	60/+	40/+	-/+	140	90	90	45			8/6	645/500
SP	85	100	-/+	-/+	-/+	45	-/+	-/+	125/+	170	50	60	60			7/5	635/300
OM	105	80	45/+	-/+	80/+	80	-/+	-/+	130/+	90	60	75	75			9/4	740/300
I	50	-/+	60/+	-/+	80/+	40	-/+	-/+	50/+	90	90	100	80			8/6	600/240
IF	125	55	-/+	-/+	-/+	40	-/+	-/+	180/+	225	125	125	30			6/3	750/410
C	150	100	-/+	T	-/+	30	-/+	170/+	170/+	225	100	30	30			8/3	825/240
February																	
U	25	-/+	-/+	-/+	-/+	25	60/+	25	90/+	125	40	30				4/3	120/300
AS	50	90	-/+	-/+	45/+	-/+	60/+	25	90/+	75	75	70				6/3	430/300
SP	75	-/+	-/+	-/+	60/+	-/+	-/+	-/+	25/+	50	50	70				4/6	285/280
OM	70/+	60	40/+	25	50/+	25	-/+	-/+	50/+	75	75	60				7/4	380/240
I	100	50	30/+	-/+	50/+	25	-/+	-/+	125/+	150	75	25				6/4	510/180
IF	100	60	-/+	-/+	-/+	25	-/+	-/+	125/+	150	75	25				5/4	205/330
C	100	60	-/+	-/+	-/+	25	-/+	-/+	125/+	150	75	25				5/3	510/200
March																	
U	25/+	T	-/+	-/+	-/+	40	-/+	-/+	-/+	25	35	45				3/4	45/290
AS	25/+	40	-/+	-/+	30/+	T	-/+	-/+	-/+	40	T	T				4/2	110/280
SP	25	40	-/+	-/+	45/+	-/+	-/+	-/+	-/+	T	25	25				3/4	55/260
OM	25	40	-/+	-/+	25/+	-/+	-/+	-/+	-/+	T	25	40				4/4	135/220
I	75	25	-/+	-/+	25	-/+	-/+	-/+	-/+	T	25	45				4/4	135/100
IF	75	25	-/+	-/+	25	-/+	-/+	-/+	-/+	T	25	40				3/2	40/240
C	75	25	-/+	-/+	25	-/+	-/+	-/+	25/+	45	45	T				4/2	170/150

cd. tab. 4 — Table 4 cont.

April									
U	-/+	-/+	-/+	+/+	-/3	-/280			
AS	-/+	-/+	-/+	-/+	-/2	-/240			
SP	-/+	-/+	-/+	-/+	-/3	-/210			
OM	-/+	-/+	-/+	-/+	1/3	20/110	T		
I	-/+	-/+	-/+	-/+	-/4	-/150			
IF	-/+	-/+	-/+	-/+	-/3	-/100			
C	-/+	-/+	-/+	-/+	2/2	40/150			
May									
Urea	-/+	-/+	-/+	-/+	-/3	-/100			

Legend: 1 — alanine/arabinose, 2 — arginine/deoxyribose, 3 — citrulline/glucose, 4 — cysteine/raffinose, 5 — glutamic acid/rhamnose, 6 — glycine/ribose, 7 — histidine/sucrose, 8 — isoleucine/trehalose, 9 — leucine/xylose, 10 — methionine, 11 — proline, 12 — serine, 13 — threonine, 14 — tryptophane, 15 — valine, T — amino acid present in traces, U — urea, AS — ammonium sulphate, SP — superphosphate, OM — organic manure, I — irrigated, IF — *Ipomoea fistulosa*, C — control (unamended).

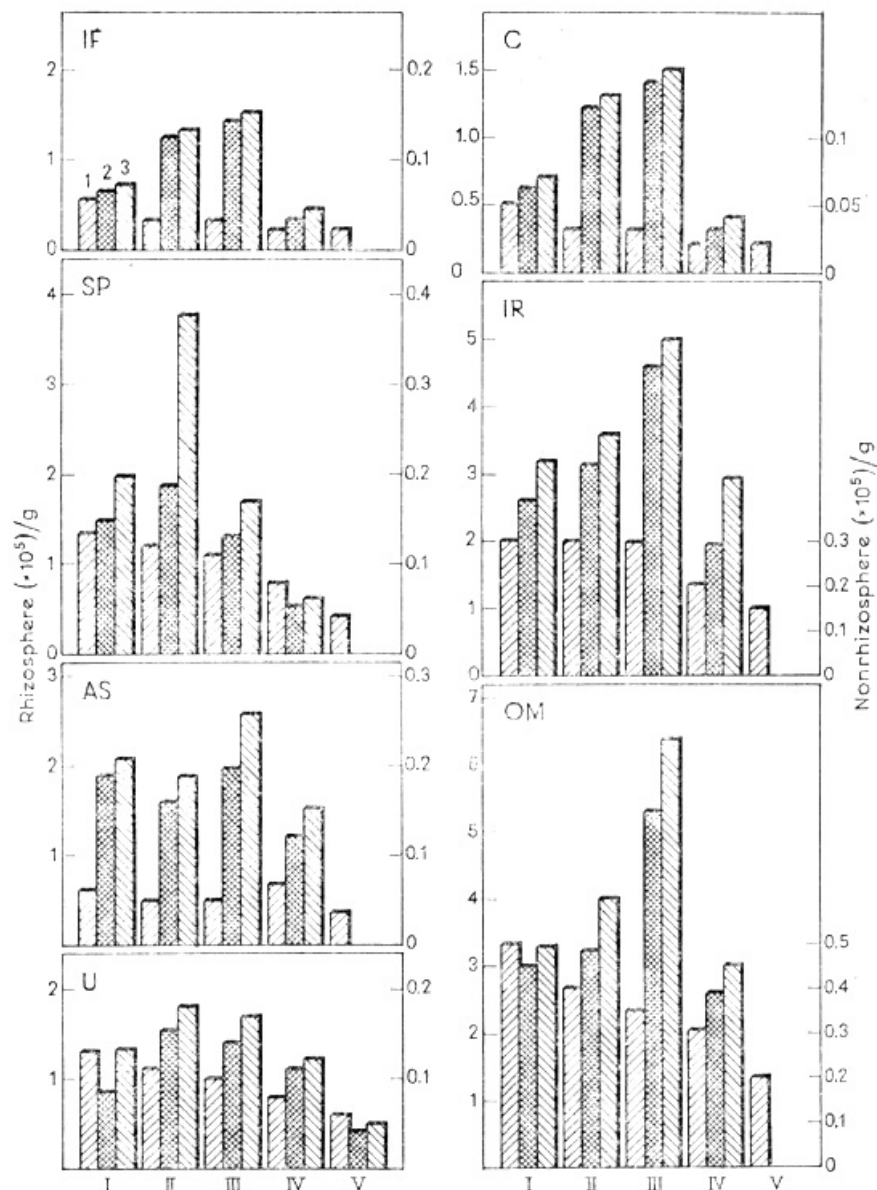


Fig. 1. Periodic variation in nonrhizosphere and rhizosphere fungal population of *Pennisetum typhoides* roots in fertilized plots in 1971

1 — nonrhizosphere fungal population ($\times 10^5$) g dry soil; 2 — rhizosphere fungal population ($\times 10^5$) g dry root; 3 — rhizosphere fungal population ($\times 10^5$) g dry soil
 IF — *Ipomoea fistulosa*; SP — superphosphate; AS — ammonium sulphate; U — urea;
 C — control; IR — irrigated; OM — organic manure

details in Table 4). Nine sugars were obtained from roots-extracts of different plots. Glucose, and xylose were observed in all the sets during different stages of root decomposition. No regular pattern of distribu-

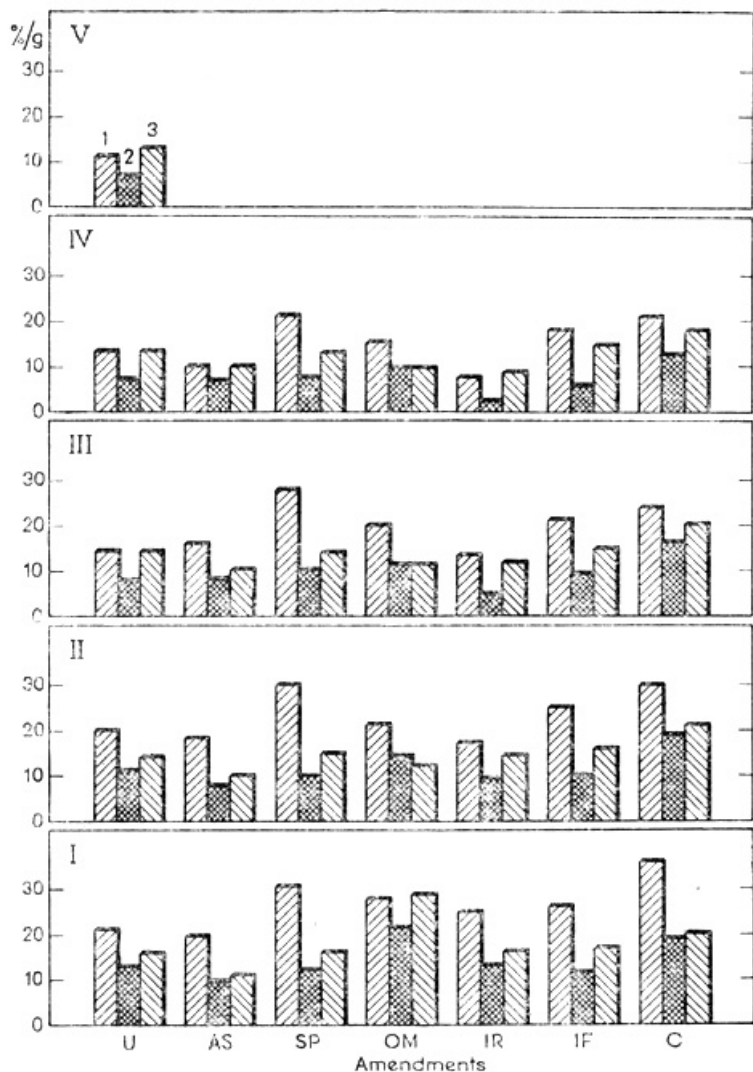


Fig. 2. Celluloses, hemicelluloses and lignins of *Pennisetum typhoides* roots decomposing in amended plots in 1971 (Amount initial dry WT of root)
 U — urea; AS — ammonium sulphate; SP — superphosphate; OM — Organic manure;
 IR — irrigated; IF — *Ipomoea fistulina*; C — control

tion of sugars in different sets was obtained. The amount of total sugars exhibited a trend similar to that for amino acids (Table 4).

Cellulose, hemicellulose and lignin components of roots

In all the sets the amount of the above three components was highest in January and then decreased gradually. Cellulose and hemicellulose decomposed more rapidly than lignin. Highest and lowest decomposition of the above components of the root was always recorded in irrigated and control sets respectively (Fig. 2).

Organic matter content and pH moisture content of fertilized plots

There existed no considerable differences in organic matter content and the pH of various plots except that OMC and MC was higher in the plots where organic matter content and water themselves were

Table 5

Organic matter content (OMC—%), moisture content (MC—%) and pH of fertilised plots

Sampling months 1971		Supplements						
		U	AS	SP	OM	I	IF	C
Jan.	OMC	2.5	2.61	2.47	3.9	2.6	2.4	2.52
	MC	15.6	16.3	15.8	14.9	25.7	15.0	15.0
	pH	6.7	6.9	6.8	6.7	6.9	7.1	6.9
Febr.	OMC	2.3	2.4	2.32	3.5	2.47	2.2	2.45
	MC	15.0	16.0	15.3	14.5	25.9	14.7	14.5
	pH	6.6	6.9	6.9	6.6	6.9	7.1	6.9
March	OMC	2.0	2.25	2.20	3.2	2.30	2.5	2.40
	MC	14.6	15.7	15.0	14.0	25.2	14.3	14.3
	pH	6.7	6.9	6.9	6.7	6.9	6.9	6.9
April	OMC	1.9	2.0	2.0	3.0	2.0	2.78	2.0
	MC	14.0	15.3	14.6	13.8	24.9	14.0	13.5
	pH	6.9	6.9	6.9	6.7	6.9	6.9	6.9
May	OMC	1.75	1.9	1.85	2.9	1.95	2.95	1.90
	MC	13.5	15.0	14.3	13.5	24.3	13.5	13.2
	pH	6.9	6.9	6.9	6.7	6.9	6.9	6.9
June	OMC	1.60						
	MC	13.1						
	pH	6.9						

Total decomposition was observed in these plots

separately supplemented (Table 5). Except in the plot fertilized with urea plot, roots in other sets decomposed completely before 25th May and no data were obtained in these sets, however, in urea, these were exhausted during June.

DISCUSSION

The mycoflora of the plots containing different commercial fertilizers, organic manure and *Ipomoea fistulosa* varied differently. Irrigated and non-irrigated plots also showed considerable variation. The number of fungal species in all the fertilized sets except those supplemented with superphosphate and *I. fistulosa* was always greater than the control (Table 1). This is probably due to the availability of more nutrients in the supplemented soils.

Waksman (1922) reported that plots receiving commercial fertilizers, applied separately or in combination, gave higher fungal counts than controls. Stimulation in fungal population by nitrate-nitrogen, phosphate and potassium has been reported by Waksman and Starkey (1924). The beneficial effects of potassium chloride, calcium phosphate, ammonium sulphate and wood-ash-extract on soil mycoflora has been reported by Zacharias (1949).

In the beginning the number of fungal species was generally higher in rhizosphere then away from it. On subsequent sampling dates, however, the pattern was reversed (Table 2). The greater number of species in this region may be ascribed to the increased level of nutrition. In rhizosphere the fungal population was influenced both by decomposing root tissues as well as supplements. Recently Mishra (1971 a, b, 1972) and Kanaujia (1973) have reported the beneficial effect of commercial fertilizers on rhizosphere mycoflora of *Oryza sativa* and *P. typhoides*. They observed that fungal species were more numerous in fertilized plots than in control ones.

Comparatively high fungal population in plots supplemented with organic manure and in irrigated at the root surface and away from it. Increased nutritional status in the former and suitable moisture in the latter possibly accounted for higher fungal flora. Higher fungal counts in the organic matter rich soils has been emphasized by many workers (Dwivedi 1966; Kanaujia and Singh 1977; Mishra and Kanaujia 1972 a, b, 1973).

A higher fungal population during the month of January from the root surface of *P. typhoides* which gradually decreased with ageing in the summer months may be attributed to the high amino acid and

sugar content (Table 4). The breakdown of cellulose, hemicellulose and lignin (Fig. 2) supported the higher fungal population in the rhizosphere region. The lowest population in May in nearly all the sets was due to the high temperature and low moisture content of the soil.

A comparatively smaller population has been recorded in the plot supplemented with *I. fistulosa* (Fig. 1) which is possibly due to the presence of some antifungal substance(s) in *I. fistulosa*. During early stages of decomposition of the plant material the low fungal population may be due to the release of higher amounts of an antifungal substance (or substances) into the soil environment. At later stages, the concentration of antifungal substance(s) possibly decreased and the plant remains consequently harboured more fungi. The effect of added fresh plant material on soil mycoflora is poorly understood and whatever little is known, is concerned with mycoflora of root diseased. Bhakuni et al. (1969) and Dhar et al. (1968) while screening Indian plants for biological activity observed that *Ipomoea illustris*, *I. leari*, *I. reptans* and *I. pestigridis* possessed antifungal, antibacterial, antiviral, antiprotozoal and antifertility activities.

A greater number of fungal species were commonly associated with different plots and yet many were restricted to only one plot the former phenomenon may be due to the similar edaphic and environmental conditions and common substrate available for colonization and the latter due to the variation in amendments and status and quality of nutrition, (Table 4).

There was no remarkable variation in the successional pattern of fungal species on the roots of *P. typhoides* plots supplemented in various ways. Few forms especially the species of *Aspergillus* and *Fenicillium*, few other *Deuteromycetes* along with some sterile fungi were found to be associated with roots at all the stages. Many of them appeared as dominants which may be due to their wide range of nutritional acceptance and greater tolerance (Cochrane 1958).

The presence of *Cladosporium* species with high frequency during winter months may be due to the low temperature prevailing in this part of the country which is in accordance with our previous studies (Mishra and Kanaujia 1972, 1973). The dominance of sterile fungi during summer months may be due to the low moisture of the soil and high temperature. Dominance of other forms at various stages of root decomposition may be due to selective response to fertilizations and quality of the nutrition (Mishra 1971 a, b, 1972 and Kanaujia 1973).

The pH of the soils (Table 5) in this study showed a poor correlation with soil fungal population and played a negligible role except for the fertilized with organic matter and for irrigated plots.

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