

The occurrence of *Septoria nodorum* Berk. and associated mycoflora in seeds of wheat cultivated in the Szczecin voivodeship

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In the years 1982-1984, the occurrence of *Septoria nodorum* and associated mycoflora in 48 and 23 samples of winter and spring cultivars of wheat were examined, respectively. The following species of fungi were most frequently recovered: *Alternaria alternata*, *Aureobasidium pullulans*, *Epicoccum purpurascens*, species of the genera *Cladosporium*, *Fusarium*, *Helminthosporium* and *Penicillium*, *S. nodorum*, and non-sporulating fungi. The occurrence of *S. nodorum* was higher in seeds coming from the wetter years 1982 and 1984. The level of seed infestation by *S. nodorum* was higher in winter cultivars. *Septoria nodorum* was more frequently isolated from shrunken than normally developed seeds.

INTRODUCTION

Septoria nodorum Berk., the cause of wheat glume blotch is considered to be important parasite of wheat all over the world (Brönnimann, 1968; Tyldesley, Thompson, 1980; Weber, 1922). Severe attacks of *S. nodorum* can cause yield losses greater than 50 % (Brönnimann, 1968; Jones, Oddebunmi, 1971; Weber, 1922). One of the sources of *S. nodorum* inoculum is a seed lot (Weber, 1922). However, the incidence of *S. nodorum* in seeds differs considerably depending on the plant resistance to this pathogen, climatic conditions in the vegetative period, site of cultivation, and conducted agrochemical practices (Nelson, Morey, Brown, 1974; Sharp, Brönnimann, McNeal, 1972; Tyldesley, Thompson, 1980). The associated mycoflora plays an important role in influencing the activity of *S. nodorum* (Dickinson, Skidmore, 1976; Skidmore, Dickinson, 1976).

The aim of this study was to determine the occurrence of *S. nodorum* and associated mycoflora in seeds of wheat cultivated in the Szczecin voivodeship.

MATERIAL AND METHODS

In the years 1982-1984, a total of 71 300-400 g seed samples were obtained from the Szczecin Voivodeship Station of Seed Estimation (Tab. 2). Forty-eight and 23 samples represented winter and spring wheat cultivars, respectively. The mean proportion of shrunken seeds in each sample based on 4 subsamples consisting of 100 seeds was determined.

Forty normally developed seeds and 40 shrunken seeds were selected randomly and surface desinfected in a solution of $HgCl_2$ prior to fungal isolation. The mycoflora was investigated by placement of five seeds in each 10 cm Petri dish containing potato glucose agar (PGA). The Petri dishes were incubated at room conditions for 10-14 days. At the end of this period, fungal colonies growing out of each seed were transferred individually to PGA slants and recognized.

Fungi were identified according to Arx (1970), Barnett (1960), Booth (1971), De Vries (1959), Domsch (1970), Drechsler (1923), Ellis (1971), Gams (1971), Gilman (1945), Raper, Thom (1949), Raper, Fennel (1965), and Zycha, Siepmann, Linnemann (1969). Except for *S. nodorum*, representatives of each the other species were grown from single conidia in Petri dishes of PGA at room temperature with a 12 h photoperiod under cool white fluorescent lamps located 40 cm above cultures. Cultures were grown for 10-14 days. *Septoria nodorum* was cultured on oatmeal agar, since this medium produces distinctive colonies with abundantly sporulating pycnidia.

Table 1
Monthly sums of rainfall in the years 1982-1984

Station		Month											
		IX	X	XI	XII	I	II	III	IV	V	VI	VII	VIII
Swinoujście	a*	40	100	80	45	53	8	38	21	59	61	12	22
	b	5	36	25	37	56	34	45	57	81	37	5	47
	c	46	34	41	34	78	29	7	21	61	107	43	29
Resko	a	36	112	108	64	67	12	25	36	57	60	29	20
	b	6	37	34	58	94	44	72	68	70	19	22	50
	c	72	43	31	29	82	22	8	20	80	167	82	62
Nowogard	a	51	110	106	52	**	-	20	6	40	-	33	26
	b	9	39	31	53	83	29	57	63	85	6	8	23
	c	-	-	-	-	-	-	-	-	-	-	-	-
Łobez	a	22	97	108	57	59	10	25	38	40	65	32	12
	b	-	-	-	-	-	-	-	-	-	-	-	-
	c	-	-	-	-	-	-	-	-	-	-	-	-
Szczecin-Dąbie	a	28	83	83	49	52	10	30	12	46	57	22	18
	b	5	35	25	34	65	28	37	77	85	7	12	53
	c	26	36	32	23	53	26	5	19	68	88	83	26
Lipki	a	23	89	81	51	47	11	44	23	63	60	13	16
	b	4	32	19	57	62	16	36	86	89	23	23	49
	c	33	31	34	12	52	21	5	23	73	125	100	32

* a - 1982, b - 1983, c - 1984; ** - data not available

RESULTS

Monthly sums of rainfall recorded in some selected meteorological stations of the Szczecin voivodship in the years 1982-1984 are presented in Table 1. Both autumn and the ripening period, June-August, were much wetter in 1982 and 1984 than in 1983.

The proportion of shrunken seeds in the seed samples examined ranged from 2.0 % in 1982 to 43 % in 1983 (Tab. 2). In winter cultivars, the shrunken seeds constituted 8.7 % (in 1982) to 16.0 % (in 1984), whereas the occurrence of shrunken seeds in spring cultivars ranged from 10.1 to 17.1 % in 1982 and 1984, respectively. Samples harvested in 1984 (av. 16.5 %) contained the highest proportion of shrunken seeds independent of cultivars.

Of the 3840 examined seeds of winter wheat cultivars harvested in the years 1982-1984, a total of 5157 fungal colonies representing 37 species from 25 genera and 9 forms of non-sporulating fungi were isolated (Tab. 3). The normally developed seeds coming from the harvests in 1982-1984 harboured 21, 20, and 22 fungal species and 5, 5 and 7 forms of non-sporulating fungi, respectively. The shrunken seeds from the 1982-1984 harvests were contaminated by 27, 19, and 20 species and 5, 4, and 7 forms of non-sporulating fungi, respectively.

The 1840 investigated seeds of spring cultivars yielded a total of 1997 colonies representing 37 species from 20 genera and 9 forms of non-sporulating fungi (Tab. 4). The mycofloras of normally developed seeds from the 1982-1984 harvests comprised of 19, 13, and 19 species and 5, 7, and 6 forms of non-sporulating fungi. The fungal communities recovered from shrunken seeds during the three-year study were represented by 22, 25, and 19 species and 5, 7, and 3 forms of non-sporulating fungi, respectively.

The fungi most frequently recovered from the seed samples investigated were as follows: *A. alternata*, *A. pullulans*, *E. purpurascens*, species of the genera *Cladosporium*, *Fusarium*, *Helminthosporium* and *Penicillium*, *Septoria nodorum*, and non-sporulating fungi (Tabs. 3, 4). The mean percentages of seeds from which these fungi were isolated are presented in Table 5.

Septoria nodorum was most frequently isolated in 1982 (273 isolates). The lowest number of colonies of this fungus was recorded in 1983 (38) (Tabs. 3, 4). Among of the wheat cultivars examined, the highest proportion of seeds infested by *S. nodorum* in 1982 was found in Grana, Saga, and Liwilla (Tab. 6). In 1983, the seeds of cultivars which were colonized by *S. nodorum* were those of Gama and Saga. In 1984, the highest proportion of seeds with *S. nodorum* occurred in Liwilla, Saga, and Jana.

Septoria nodorum was more frequently isolated from shrunken (0.8-38.1 %) than normally developed seeds (1.5-36.3 %) (Tabs. 3, 4). Of the other fungi most frequently recovered in this study, only members of the genera *Fusarium* and *Penicillium* were more frequently associated with shrunken seeds. In contrast, *A. alternata* and non-sporulating fungi were more frequently isolated from normally developed

seeds. No correlations were found between the frequency of occurrence of *A. pullulans*, *E. purpurascens*, and *Cladosporium* spp. and the degree of seed development.

The other fungi listed in Tables 3 and 4 sporadically contaminated the investigated seed samples.

Table 2

Wheat cultivars from which seed samples were collected

Year of harvest	Cultivar	Degree of reproduction	% of shrunked seeds	Site of cultivation
1982	Liwilla	original	7.9	Tapadły
		1st reprod.	8.0	Bielicz
	Jana	original	6.8	Lubanów
		1st reprod.	3.8	Grzybno
		1st reprod.	11.3	Trzyglów
	Beta	1st reprod.	4.8	Koszewo
		1st reprod.	12.0	Żuków
	Maris Huntsman	original	13.8	Barkowo
	Grana	original	12.8	Laskowo
		basic seed	5.3	Cerkwica
		1st reprod.	12.5	Wrzosowo Kamień
	Saga	1st reprod.	8.3	Przelewice
		1st reprod.	13.0	Wyszobór
		original	10.3	Sienno-Dolne
		original	5.3	Przeclaw
	Salwa	original	11.8	Mobtwo
		original	7.0	Topolek
		1st reprod.	3.8	Nowielice
		original	3.5	Ostoja
	1st reprod.		7.5	Skrzykocin
	Jara	1st reprod.	18.3	Żabów
		original	6.0	Stróżewo
original		14.8	Orle	
original		25.0	Kolbacz	
original		12.5	Dobropole	
original		2.0	Koszewo	
Kolibri	1st reprod.	5.5	Witnica	
	1st reprod.	6.0	Witnica	
	1st reprod.	7.5	Boniewice	
	1st reprod.	3.0	Gardno	
1983	Beta	1st reprod.	7.3	Karsko
	Maris Huntsman	1st reprod.	43.8	Borkowo

			cont. Tab. 2	
1983	Liwilla	original	7.5	Reclaw
		elita	6.5	Lubiechowo
		1st reprod.	8.3	Skarbimierzyce
		1st reprod.	12.0	Cerkwica
	Gama	1st reprod.	16.0	Prusinowo
	Saga	1st reprod.	7.0	Kłodzino
		1st reprod.	11.8	Dargosław
	Salwa	original	16.5	Krzemlin
		elita	3.7	Żabów
	Jana	1st reprod.	9.5	Skalin
	Grana	1st reprod.	5.0	Krzemlin
		1st reprod.	7.5	Krzemlin
	Sawa	original	9.3	Ostoja
	Jara	1st reprod.	18.5	Barzkowice
		1st reprod.	14.0	Drzemień
		original	12.0	Cerkwica
		original	12.3	Kluczewo
original		11.5	Tagadły	
Williamia	original	7.8	Witnica	
Kolibri	1st reprod.	12.5	Witnica	
	1st reprod.	14.6	Gardno	
1984	Liwilla	1st reprod.	25.5	Lubanowo
		1st reprod.	10.0	Witnica
		original	18.5	Cerkwica
		original	22.5	Siemieczyn
	Gama	1st reprod.	9.0	Płońsko
		1st reprod.	13.5	Strzykocin
		1st reprod.	6.0	Skarbimierzyce
		1st reprod.	10.5	Przeclaw
		basic seed	11.0	Strzyżno
	Jana	1st reprod.	15.0	Molstkowo
		1st reprod.	20.0	Koszewo
		1st reprod.	21.5	Samolino
	Saga	basic seed	12.5	Bielin
		1st reprod.	28.0	Węgorzyno
	Jara	1st reprod.	16.5	Drzemien
		1st reprod.	14.5	Cerkwica
		1st reprod.	18.5	Karnice
1st reprod.		23.0	Krasne	
	1st reprod.	13.0	Krasne	

Table 3

Fungi recovered from seeds of winter wheat cultivars harvested in 1982-1984

Fungus	1982		1983		1984		Total
	a	b	a	b	a	b	
<i>Absidia glauca</i> Hagem	2	1	-	-	-	-	3
<i>Acremonium atrum</i> Sacc.	-	2	-	-	1	-	3
<i>Acremonium</i> sp.	-	-	2	2	-	-	4
<i>Alternaria alternata</i> (Fr.) Keissler	354	279	314	279	375	330	1931
<i>Aspergillus ruber</i> (Konig, Speieckermann et Bremer) Thom et Church	-	1	-	-	-	-	1
<i>Aureobasidium pullulans</i> (de Bary) Arn.	21	33	15	24	2	-	95
<i>Botrytis cinerea</i> Pers.: Fr.	4	8	3	9	5	9	38
<i>Chaetomium globosum</i> Kunze	1	2	1	2	-	-	6
<i>C. olivaceum</i> Cooke et Ellis	-	1	-	-	-	-	1
<i>Circinella rigida</i> Smith	-	-	-	-	1	1	2
<i>Cladosporium cladosporioides</i> (Fres.) de Vries	13	16	8	10	1	-	48
<i>C. herbarum</i> Link.: Fr.	15	12	11	9	3	4	54
<i>C. macrocarpum</i> Preuss	-	-	3	-	1	-	4
<i>Epicoccum purpurascens</i> Link	26	26	17	28	105	65	267
<i>Fusarium avenaceum</i> (Corda: Fr.) Sacc.	1	-	1	-	2	3	7
<i>F. culmorum</i> (W. G. Smith) Sacc.	-	3	1	2	5	3	14
<i>F. graminearum</i> Schwabe	-	3	-	2	-	1	6
<i>F. lateritium</i> Nees	-	-	-	1	-	3	4
<i>F. nivale</i> (Fr.) Ces.	7	7	-	-	6	38	58
<i>F. poae</i> (Peck) Wollenw.	5	3	5	14	4	41	72
<i>Fusidium</i> sp.	-	1	-	-	1	-	2
<i>Gonatobotrys simplex</i> Corda	3	9	4	8	1	-	25
<i>Helminthosporium sativum</i> P. K. B.	3	3	1	-	1	-	8
<i>H. triseptatum</i> Drechs.	32	17	1	-	-	1	51
<i>Mucor hiemalis</i> Wehmer.	2	7	1	1	1	2	14
<i>M. racemosus</i> Fres.	-	-	-	-	1	2	3
<i>M. strictus</i> Hagem	8	15	1	1	1	-	26
<i>Papularia arundinis</i> (Corda) Fr.	4	3	-	1	-	1	9
<i>P. sphaerosperma</i> (Pers.) Hohn.	-	1	-	-	-	-	1
<i>Penicillium notatum</i> Westling	-	-	-	-	6	6	12
<i>Penicillium</i> spp.	23	26	24	34	5	4	116
<i>Phoma eupyrena</i> Sacc.	1	-	-	-	-	-	1
<i>Phoma</i> sp. 1	1	-	-	-	2	-	3
<i>Phoma</i> sp. 2	-	-	-	-	1	-	1
<i>Rhizopus nigricans</i> Ehrenb.	-	-	4	8	6	5	23
<i>Sclerotium</i> sp.	-	1	-	-	-	-	1
<i>Septoria odororum</i> Berk.	124	149	14	24	131	141	583
<i>Stemphylium botryosum</i> Wallr.	9	5	11	8	2	2	37
<i>Trichocladium asperum</i> Harz	-	1	-	-	-	-	1
<i>Trichoderma viride</i> Pers.: Fr.	2	-	-	-	-	-	2
<i>Trichothecium roseum</i> Link	-	-	-	2	-	-	2
<i>Ulocladium atrum</i> Preuss	1	3	1	2	-	-	7
<i>U. botrytis</i> Preuss	-	1	1	1	-	-	3
<i>Verticillium indicum</i> (Petch) Gams	-	1	-	-	-	-	1
<i>Verticillium</i> sp.	-	1	-	-	-	-	1
Yeast-like pink	5	19	5	5	3	2	39
Non-sporulating	341	275	376	293	154	134	1573
Total	1006	934	824	769	827	797	5157

Table 4

Fungi recovered from seeds of spring wheat cultivars harvested in 1982-1984

Fungus	1982		1983		1984		Total
	a	b	a	b	a	b	
<i>Acremoniella atra</i> Sacc.	-	-	-	-	-	2	2
<i>Alternaria alternata</i> (Fr.) Keissl.	125	92	134	111	134	95	691
<i>Aspergillus niger</i> v. Tieghem	1	4	-	-	1	1	7
<i>A. versicolor</i> (Vuill.) Tiraboschi	-	1	-	-	-	-	1
<i>Aspergillus</i> sp.	-	1	-	-	-	-	1
<i>Aureobasidium pullulans</i> (de Bary) Arn.	10	11	15	15	1	1	53
<i>Botrytis cinerea</i> Pers.: Fr.	1	4	6	2	2	1	16
<i>Chaetomium globosum</i> Kunze: Fr.	-	-	1	-	-	-	1
<i>C. indicum</i> Corda	1	2	-	-	-	-	3
<i>Circinella muscae</i> (Sorok.) Berl. et de Toni	-	-	-	1	-	-	1
<i>C. rigida</i> Smith	-	-	-	2	-	-	2
<i>Cladosporium chadosporioides</i> (Fres.) de Vries	11	25	2	4	1	-	43
<i>C. herbarum</i> Link: Fr.	17	22	2	1	3	-	45
<i>C. macrocarpum</i> Preuss	2	-	-	1	-	-	3
<i>Epicoccum purpurascens</i> Link	4	3	-	3	43	28	81
<i>Fusarium avenaceum</i> (Corda: Fr.) Sacc.	-	-	-	-	-	1	1
<i>F. culmorum</i> (W. G. Smith) Sacc.	-	-	-	2	1	2	5
<i>F. graminearum</i> Schwabe	-	2	-	1	-	2	5
<i>F. lateritium</i> Nees	-	1	-	1	-	1	3
<i>F. nivae</i> (Fr.) Ces.	-	3	-	-	1	4	8
<i>F. oxysporum</i> Schl.	-	-	1	-	-	-	1
<i>F. poae</i> (Peck) Wollenw.	1	4	-	1	-	6	12
<i>F. sporotrichioides</i> Sherb.	-	-	-	1	-	-	1
<i>Gonotobotrys simplex</i> Corda	2	4	-	2	1	-	9
<i>Helminthosporium sativum</i> P. K. B.	3	1	-	1	1	-	6
<i>H. triseptatum</i> Drechs.	1	-	-	-	-	1	2
<i>Mucor hiemalis</i> Wehmer	1	1	1	3	1	1	8
<i>M. plumbeus</i> Bonorden	-	-	-	1	-	-	1
<i>M. strictus</i> Hagem	3	1	3	1	1	-	9
<i>Papularia arundinis</i> (Corda) Fr.	-	6	-	2	-	-	8
<i>P. sphaerosperma</i> (Pers.) Hohn.	-	1	-	-	-	-	1
<i>Penicillium notatum</i> Westl.	-	-	-	-	4	4	8
<i>Penicillium</i> spp.	20	25	5	9	2	2	63
<i>Phoma</i> sp.	1	-	-	-	-	-	1
<i>Rhizopus nigricans</i> Ehrenb.	-	-	2	7	-	-	9
<i>Septoria nodorum</i> Berk.	14	18	6	9	17	31	95
<i>Stemphylium botryosum</i> Walk.	6	9	3	1	2	-	21
<i>Trichothecium roseum</i> Link	1	-	-	1	-	-	2
<i>Ulocladium atrum</i> Preuss	1	-	-	-	-	-	1
<i>U. botrytis</i> Preuss	-	1	1	1	-	-	3
Yeast-like pink	4	3	9	8	2	1	27
Non-sporulating B-1	191	129	180	173	42	23	738
Total	421	374	371	365	260	206	1997

Table 5

Mean percentage of seeds of winter and spring cultivars from which the most frequently occurring fungi were isolated in the years 1982-1984

Fungus		Winter cultivars			Spring cultivars		
		1982	1983	1984	1982	1983	1984
<i>Alternaria alternata</i>	a	42	52	65	32	42	65
	b	37	44	57	27	37	48
<i>Aureobasidium pullulans</i>	a	3	3	4	3	4	1
	b	4	4	0	3	5	1
<i>Cladosporium</i> spp.	a	4	4	2	8	2	3
	b	4	4	2	12	2	0
<i>Epicoecum purpurascens</i>	a	4	4	19	2	0	19
	b	4	4	10	1	2	13
<i>Fusarium</i> spp.	a	2	1	4	1	2	3
	b	2	2	18	3	3	8
<i>Helminthosporium</i> spp.	a	4	1	1	3	0	1
	b	2	0	0	1	2	1
<i>Penicillium</i> spp.	a	4	2	1	6	3	4
	b	2	2	2	7	4	4
<i>Septoria nodorum</i>	a	13	2	23	4	3	8
	b	19	2	27	5	4	18
Non-sporulating fungi	a	30	63	27	48	55	18
	b	31	51	24	30	52	10

a - normally developed seeds; b - shrunken seeds

Table 6

Mean percentage infestation of seeds of winter (A) and spring (B) cultivars by *Septoria nodorum*

Cultivar	Year						
	1982		1983		1984		
	a	b	a	b	a	b	
(A)	Beta	8	15	0	0	-	-
	Gama	-	-	14	18	10	16
	Grana	22	28	2	2	-	-
	Jana	13	9	0	0	24	23
	Liwilla	17	20	2	3	35	37
	Maris Huntsman	14	10	0	0	-	-
	Saga	17	23	3	9	25	29
	Salwa	11	12	0	0	-	-
	Sawa	-	-	0	0	-	-
(B)	Jara	5	3	2	1	8	16
	Kolibri	3	1	4	8	-	-
	Sappo	0	0	-	-	-	-
	Williana	-	-	0	0	-	-

a - normally developed seeds; b - shrunken seeds

DISCUSSION

Most of the fungal species isolated during the study discussed in the present paper are known to occur on cereal seeds, including *T. aestivum* (e.g., Flannigan, 1971; Hänni, 1980; Hewett, 1965).

The predominance of *A. alternata*, *A. pullulans*, *E. purpurascens*, species of the genera *Cladosporium*, *Fusarium*, *Helminthosporium* and *Penicillium*, *S. nodorum*, and non-sporulating fungi in the fungal populations isolated fits the results of e.g., Flannigan (1971) and Łacicowa (1964).

Except for members of *Penicillium* and non-sporulating fungi, the other fungi listed above are known to be called "field fungi" (Neergaard, 1977). According to Chełkowski (in Gabińska, Narkiewicz-Jodko, Schneider, 1991) and Narkiewicz-Jodko (1991), the presence of *A. alternata*, *Cladosporium* spp., and *E. purpurascens* suggests a good quality of a seed lot. Bateman (1979) found these fungi to inhabit *S. nodorum*. *Aureobasidium pullans* also showed antagonistic properties to *S. nodorum* (Fokema, Van Der Meulen, 1976).

Species of the genera *Fusarium* and *Helminthosporium* are known to produce seed rot either in the crop or during germination (Colhoun, Park, 1964; Hänni, 1980; Łacicowa, 1963).

Penicillium spp. are "storage fungi" (Neergaard, 1977) that spread rapidly at a higher moisture content (Hill, Lacey, 1983) and usually replace "field fungi" (Lutey, Christensen, 1963). *Penicillium* spp. decrease germination of seeds (Neergaard, 1977).

The role of non-sporulating fungi in mycoflora of wheat seeds is unknown.

The seeds harvested in 1982 and 1984 yielded many more colonies of *S. nodorum* than those from the 1983 harvest. Weather records from meteorological station suggest that this may have resulted from the wetter vegetative periods in 1982 and 1984. Seed infection by *S. nodorum* is favoured by wet weather (Leiteritz, Focke, 1977).

The proportion of seeds with *S. nodorum* was higher in winter than spring cultivars. This supports the opinion of Obst (1980) that winter cultivars grow in more favourable conditions for establishment of *S. nodorum* infections.

The higher incidence of *S. nodorum* in shrunken than normally developed seeds conforms the findings of many investigators (e.g., Cooke, Jones, 1970; Jones, Odubunmi, 1971) that this fungus is a severe pathogen highly reducing the yield of wheat.

REFERENCES

- ARR J. A. von, 1970. The genera of fungi sporulating in pure culture. J. Cramer, Lehre.
BARNETT H. L., 1960. Illustrated Genera of Imperfect Fungi. Minneapolis.
BATEMAN G. L., 1979. Relationships between *Fusarium nivale* and other microorganisms on seed of wheat and barley. Trans. Br. Mycol. Soc. 72: 245-249.

- Booth C., 1971. The genus *Fusarium*. Commonwealth Mycol. Inst. Kew, Surrey.
- Brönnimann A., 1968. Zur Kenntnis von *Septoria nodorum* Berk., dem Erreger der Spelzenbräune und einer Blattfäule des Weizens. *Phytopath. Z.* 61: 101-146.
- Cooke B. M., Jones D. G., 1970. The epidemiology of *Septoria tritici* and *S. nodorum*. II. Comparative studies of head infection by *Septoria tritici* and *S. nodorum* on spring wheat. *Trans. Br. Mycol. Soc.* 54: 395-404.
- Colhoun J., Park D., 1964. *Fusarium* diseases of cereals. I. Infection of wheat plants with particular reference to the effects of soil moisture and temperature on seedling infection. *Trans. Br. Mycol. Soc.* 47: 559-572.
- De Vries G. A., 1959. Contribution to the knowledge of the genus *Cladosporium*. Baarn, Dickinson.
- Dickinson C. H., Skidmore A. M., 1976. Interactions between germinating spores of *Septoria nodorum* and phylloplane fungi. *Trans. Br. Mycol. Soc.* 66: 45-56.
- Domsch N. K., Gams W., 1970. *Pilze aus Agrarböden*. Stuttgart.
- Drechsler C., 1923. Some graminicolous species of *Helminthosporium*. *J. Agric. Res.* 24: 641-740.
- Ellis M. B., 1971. *Dematiaceous Hyphomycetes*. C. M. I., Kew, Surrey.
- Flannigan B., 1971. Distribution of seed-borne micro-organisms in naked barley and wheat before harvest. *Trans. Br. Mycol. Soc.* 62: 51-58.
- Fokkema N. J., Van Der Meulen F., 1976. Antagonism of yeast-like phyllosphere fungi against *Septoria nodorum* on wheat leaves. *Neth J. Pl. Path.* 82: 13-16.
- Gabińska K., Narkiewicz-Jodko M., Schneider J., 1991. Wpływ wieloletniego przechowywania na wartość siewną pszenżyta ozimego. *Biul. IJAR* 180: 43-52.
- Gams W., 1971. *Cephalosporium-artige Schimmelpilze Hyphomycetes*. Stuttgart.
- Gilman I. C., 1945. *A manual of soil fungi*. Ames - Iowa.
- Häni F., 1980. Über Getreidefusariosen in der Schweiz: Saatgutbefall, Ährenbefall und Bodenkontamination. *Z. Pflkrankh. Pflsch.* 87: 257-280.
- Hewett P. D., 1965. A survey of seed-borne fungi of wheat. I. The incidence of *Leptosphaeria nodorum* and *Griphosphaeria nivalis*. *Trans. Br. Mycol. Soc.* 48: 59-72.
- Hill R. A., Lacey J., 1983. The microflora of ripening barley grain and the effects of pre-harvested fungicide application. *Ann. Appl. Biol.* 102: 455-465.
- Jones D. G., Odebumi K., 1971. The epidemiology of *Septoria tritici* and *S. nodorum*. V. Effect of mixed inocula on disease symptoms and yield in two spring wheat varieties. *Trans. Br. Mycol. Soc.* 57: 153-159.
- Leiteritz R., Focke L., 1977. Occurrence of winter wheat glume blotch (*Septoria nodorum* Berk.) and *Fusarium ear rot* [*Fusarium culmorum* (W. G. Sm)] in the German Democratic Republic as influenced by rainfall. *Arch. Phytopath. Pflanzenesch.* 13: 407-418.
- Lutey R. W., Christensen C. M., 1963. Influence of moisture content, temperature, and length of storage upon survival of fungi in barley kernels. *Phytopathology* 53: 713-717.
- Łacikowa B., 1963. Badania nad morfologią i biologią *Fusarium poae* (Pk.) Wr. oraz patogenicznością tego gatunku względem siewek pszenicy. *Ann. Univ. M. Curie-Skłodowska* 18: 419-439.
- Łacikowa B., 1964. Badania mikoflory materiału siewnego pszenicy uprawianej na obszarze woj. lubelskiego uwzględniające szczególnie grzyby patogeniczne. *Ibid.* 19: 381-406.
- Narkiewicz-Jodko M., 1991. Wpływ warunków zbioru na mikoflorę przechowywanego ziarna pszenżyta ozimego. *Biul. IJAR* 180: 33-41.
- Nelson L. R., Morey D. D., Brown A. R., 1974. Wheat cultivar responses to severe glume blotch in Georgia (*Leptosphaeria nodorum*). *Plant. Dis. Repr.* 58: 21-23.
- Obst A., 1980. In search of a simple method for the prognosis of *Septoria nodorum* glume blotch of wheat. *Protect. Ecol.* 2: 275-279.
- Raper K. B., Fennel D., 1965. The genus *Aspergillus*. Baltimore.
- Raper K. B., Thom Ch., 1949. *A manual of the Penicillia*. Baltimore.
- Sharp E. L., Brönnimann A., McNeal F. H., 1972. Reaction of selected spring wheat varieties to infection by *Septoria nodorum*. *Plant Dis. Repr.* 56: 761-764.
- Skidmore A. M., Dickinson C. H., 1976. Colony interactions and hyphal interference between *Septoria nodorum* and phylloplane fungi. *Trans. Br. Mycol. Soc.* 66: 57-64.
- Tyldesley J. B., Thompson N., 1980. Forecasting *Septoria nodorum* on winter wheat in England and Wales. *Pl. Pathol.* 29: 9-20.
- Weber G., 1922. *Septoria* diseases of wheat. *Phytopathology* 12: 537-585.
- Zycha H., Siepmann R., Linneemann G., 1969. *Micorales*. Lehre.