

**Macromycetes of oak-lime-hornbeam woods
in the Niepolomice Forest near Kraków (S Poland)
— monitoring studies**

WŁADYSŁAW WOJEWODA, ZOFIA HEINRICH, and HALINA KOMOROWSKA

Mycology Laboratory, W. Szafer Institute of Botany, Polish Academy of Sciences
Lubicz 46, PL-31-512 Kraków, Poland

Wojewoda W., Heinrich Z., Komorowska H.: *Macromycetes of oak-lime-hornbeam woods in the Niepolomice Forest near Kraków (S Poland) — monitoring studies.* Acta Mycol. 34 (2): 201–266, 1999.

In the years 1994–1996 studies on macromycetes of the Niepolomice Forest near Kraków were made in four plots designated in deciduous forests (*Tilio-Carpinetum stachydotum*) with a population of *Carpinus betulus*, *Quercus robur* and *Tilia cordata* (the size of each plot was 1000 m²). The observations were made through an international project "Mycological monitoring in European oak forests". As many as 274 species were recorded, including 234 saprobic, 33 mycorrhizal, and 7 parasitic fungi. Moreover, 15 species of fungi are connected with oak, 24 species of fungi are threatened, and 16 species are new to Poland.

Key words: *Ascomycota, Basidiomycota, ecology of fungi, oak, mycological monitoring, threatened fungi.*

INTRODUCTION

In 1992 the European Council for the Conservation of Fungi initiated a project to monitor the disappearance of macromycetes in European oak forests. The project was included in the Copernicus Programme organized by the European Union. Its contributors were mycologists from the Czech Republic (coordinator Dr. Rostislav Fellner, Prague), Italy (coordinator Dr. Claudia Perini, University in Siena, which represented the project for the European Union in Brussels), and Poland (coordinator Prof. Maria Ławrynowicz, University of Łódź). A series of mycosociological studies made in 60's and 70's on permanent plots in the oak forests of Northern Poland favored Poland's participation in the project (Nespia k 1959; Lisiewska 1965; Lisiewska and Bujakiewicz 1976). It was expected that studies repeated after 20 years would provide data on the changes caused by anthropopression in the specific composition of fungi.

In Poland, the studies including different regions of the country were performed: in the Iński Landscape Park (NW Poland) by Stasińska, in the Kraków–Częstochowa Upland (S Poland) by Ławrynowicz, in Wielkopolska (W Poland) by Lisiewska and Połczyńska (1998), in the Białowieża National Park by Skirgielło (1998b) and in the Świętokrzyskie Mts. (Łuszczyska 1998). The results from the Niepołomice Forest near Kraków (S Poland) by Wojewoda and team are presented in this paper.

We are indebted to the following persons for their help: Prof. M. Ławrynowicz for coordination of these investigation Mrs. B. Pleban for her participation in the field studies and technical work as well as preparation of this manuscript for publication, Prof. J. Jakubowska-Gabara for making phytosociological tables, B. Szczepanowicz, M.Sc., for performing soil analyses, J. Cichocki, M.Sc. and S. Jurzecki, M.Sc. for delivering climatic data, Dr. A. Chlebicki for identification of some *Pyrenomyces* species, M. Piątek, M.Sc. for assistance in field studies and graphic work performed for this paper, M. Matyjaszkiewicz, M.Sc. for preparing the drawings.

METHODS

The subject of the study were macromycetes (fungi whose fructifications are greater than 5 mm in diameter), *Asco-* and *Basidiomycota*.

Monitoring was aimed at the study of the population of macromycetes in an oak-lime-hornbeam wood in the Niepołomice Forest near Kraków, its specific composition, phenology, abundance, ecological function, and dependence on anthropopression.

Four study plots were selected (each of 1000 m²). They were delineated in the field and each was divided into 10 subplots (each of 100 m²). Prof. Jakubowska-Gabara (1996) performed phytosociological studies on all plots in Poland, including the Niepołomice Forest. On August 27, 1997 B. Szczepanowicz, M.Sc. collected soil samples from each plot from the horizons: A₀L, A₀H, and A1, and determined pH.

The Kraków branch of the Institute of Meteorology and Water Management provided annual mean temperature values and monthly rainfall totals for the period 1994–1996 from the records kept by the Climatic Station in Igołomia, located 8 km NE of study plots.

In 1994 the studies were launched in July and completed in November. Over the years 1995–1996 observations were made once or twice a month from spring to autumn. Altogether, 26 individual observations were performed. In each plot the following were sampled: the number of fructifications of selected species, substrate (terrestrial, lignicolous, and litter decomposing fungi), and trophism.

The fungi were determinated on the basis of monographic books. The authors are mentioned on the list of references. Dark-spore *Agaricales* were determinated by Z. Heinrich, white-spore *Agaricales* – by H. Komorowska, other groups of fungi – by W. Wojewoda. The names of orders were used mainly according to Hawksworth et al. (1995), names of species – according to Hansen and Knudsen (1992), Ryvarden and Gilbertson (1993–1994), Kreisel (1979, 1987), and Michael et al. (1986).

The herbarium collection of fungi has been deposited in the KRAM herbarium in the W. Szafer Institute of Botany, Polish Academy of Sciences in Kraków.

STUDY AREA

Description of the Niepolomice Forest. The selected four study plots were located in the northern forest complex of the Niepolomice Forest, 30 km from the center of Kraków (Fig. 1) in the subprovince of the Sandomierz Valley (Kondracki 1978, 1994). It is the largest forest in Kraków's neighborhood, a remnant of the primeval forests that once extended from the Carpathians to Małopolska Province. Originally, these were simply the forests growing between Kraków and the Dunajec river which, as a result of intensive logging, towards the end of the Middle Ages shrunk into a small forest area located at the junction of the Wisła and Raba rivers (Smolski 1981).

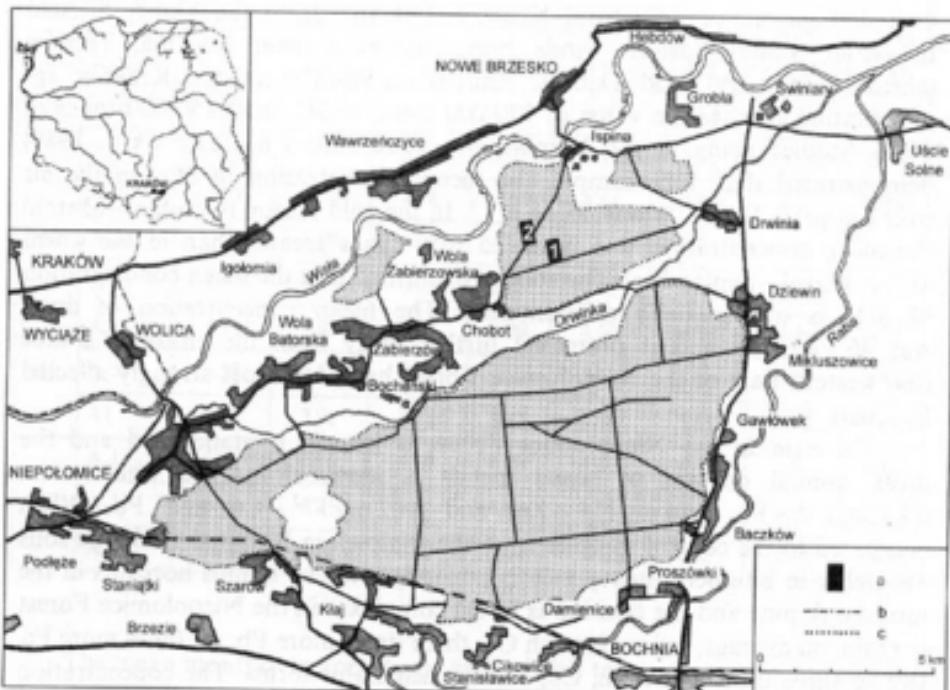


Fig. 1. Location of investigated plots in the Niepolomice Forest near Kraków: a — localities; 1 — forest section 460, the Lipówka reserve, locality 1 (plots I and II), 2 — forest section 447, locality 2 (plots III and IV), b — boundaries of Kraków city, c — boundaries of the Niepolomice Forest

Today's Niepołomice Forest is a forest area totally transformed in comparison to its primeval character and considerably rebuilt (Dziewiolski 1976). The major forest-forming species are: *Pinus sylvestris* (71%), *Quercus robur*, *Q. petraea* (17%), and *Alnus glutinosa* (9%). A greater part of the forest area is occupied by mixed woods growing in the southern part of the complex. *Tilio-Carpinetum* associations, with only a few relics of primeval nature, grow in the north. These forests include a proportion of *Quercus robur*, *Tilia cordata*, *Carpinus betulus*, little mixture of *Pinus sylvestris* and fairly rare specimens of *Sambucus nigra*. In the forest undergrowth occur, among other things, *Aegopodium podagraria*, *Asperula odorata*, and *Stellaria holostea* (Dziewiolski 1978; Ferchmin and Medwecka-Kornas 1976; Myczkowski 1981).

The age-old harmful activity of man in the given area intensified in the last 40 years by industrial emission by the agglomeration of Kraków. Many industrial works are located in that area. The most harmful are the T. Sendzimir Steelwork (the former Lenin Steelwork) and a huge electric power station. Apart from these two, thousands of chimneys and small coal-fuelled boiler houses in Kraków, Niepołomice, and other settlements pollute the environment. Winds blowing mainly from the west transport large quantities of dust and gases not only from Kraków, but also from the Upper Silesian industrial center. Eastern winds bring pollution from Bochnia, Tarnów (chemical industry), and Dębica. Annual emission from the Kraków agglomeration reaches the value of 170 000 tones of SO₂ and 150 000 tones of dusts. Studies made in the Niepołomice Forest (Grodziska 1980) demonstrated that, for example, the mean concentration of SO₂ in the air over the years 1977–78 was 25 µg m⁻³. In the cold season (October–March) the mean concentration was from 2.5 to 7 times greater than in the warm season (April–September). The studies confirmed that the mean concentration of SO₂ is not spatially differentiated. The mean concentration of dusts was 46.2 µg m⁻³ and it decreased further away from the emission source (the western part of the Niepołomice Forest has been most strongly affected by dust).

The rain in the Niepołomice Forest is heavily contaminated and the mean annual content of heavy metals is identical for the whole area: 0.13 mg l⁻¹ – Fe, 0.86 mg l⁻¹ – Zn, and 0.02 mg l⁻¹ – Cu and Pb. When compared to the control samples from the Białowieża National Park, the soils are richer in bitumens, heavy metals and sulphur. The humus horizons in the mixed oak-pine and the deciduous forest complexes in the Niepołomice Forest contain, on average, twice as much Cd, three times more Pb, six times more Fe, two to three times more Zn, Cu, Mn in assimilable forms. The concentration of sulphate (S – SO₄) is 2 to 4.5 times more than in the control samples (Grodziska 1982).

Economic changes occurring in Poland after 1989 caused a major decrease in the emission of gas and dust. However, permissible values for

the dustfall have been continuously exceeded (Grodzinska 1994). For example, in 1995 the Tadeusz Sendzimir Steelwork (the former Lenin Steelwork) emitted 10 000 tones of dusts and 13 000 tones of SO₂ into the air (Garścja 1996).

Macromyces of this area were examined by Wojewoda (1978, 1979) over the period 1974–1976, and Komorowska (1980, 1986, 1991) over the period 1976–1986. These investigations overlap the study area of the project "Mycological monitoring in European oak forests".

Description of the study plots. All study plots are located in the northern part of the Niepołomice Forest, 30 km E of Kraków. The subassociation of *Tilio-Carpinetum stachyetosum* where study plots were delineated occupy the gleyed brown soils also called old warp soils (Adamczuk 1984). Characteristic of soils on investigated plots I–IV are shown in Table 1.

Table 1
Characteristics of soils on the plots I–IV

Plots	Horizons	pH in H ₂ O				pH in KCL			
		sample 1	sample 2	sample 3	average	sample 1	sample 2	sample 3	average
I	A ₀ L	4.72	5.08	4.71	4.83	4.42	4.71	4.33	4.52
	A ₀ H	4.31	4.25	3.9	4.15	3.66	3.92	3.35	3.64
	A1	3.97	3.76	3.39	3.7	3.47	3.41	3.13	3.33
II	A ₀ L	5.03	5.14	4.98	5.05	4.54	4.67	4.57	4.59
	A ₀ H	—	4.09	4.08	4.85	—	3.7	3.8	3.75
	A1	4.15	3.78	3.73	3.88	3.41	3.47	3.46	3.44
III	A ₀ L	4.98	4.98	5.01	4.99	4.56	4.54	4.49	4.53
	A ₀ H	4.58	4.17	4.35	4.36	3.91	3.8	3.72	3.81
	A1	3.98	3.91	3.72	3.87	3.68	3.55	3.47	3.56
IV	A ₀ L	4.85	4.96	4.88	4.89	4.4	4.6	4.57	4.52
	A ₀ H	4.19	4.21	4.07	4.15	3.76	3.84	3.81	3.8
	A1	3.78	3.69	3.57	3.68	3.49	3.43	3.41	3.44

The mean monthly temperatures and rainfall totals of selected months for the period 1994–1996 are presented in Figs 2 and 3.

Locality 1 (plots I, II). This locality is situated in the strict reserve of Lipówka in the forest division 460 (Fig. 1) on the right side of the Niepołomice–Ispina road.

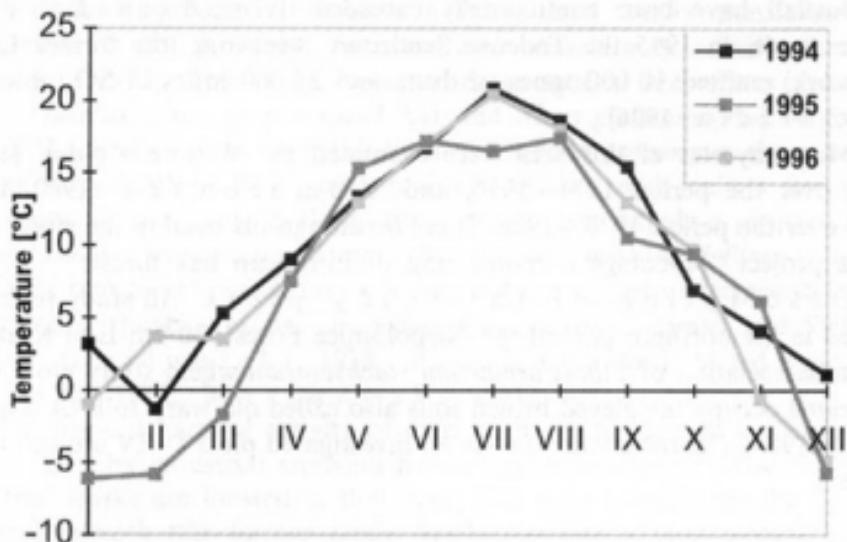


Fig. 2. Climatogram for the Igolomia meteorological station (8 km W of investigated plots) for the years 1994–1996: monthly average air temperatures

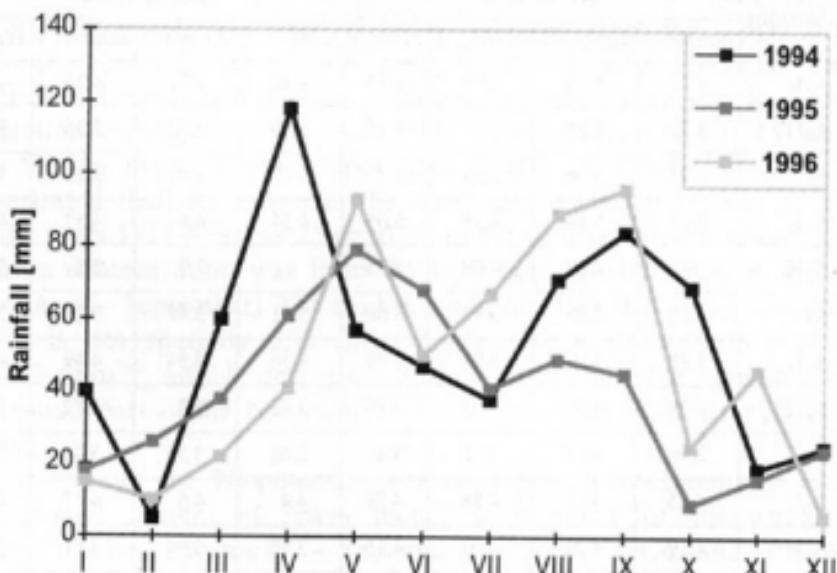


Fig. 3. Climatogram for the Igolomia meteorological station (8 km W of investigated plots) for the years 1994–1996: monthly rainfall total

The plots are adjacent. They are located in the forest with old trees of oak (*Quercus robur*), lime (*Tilia cordata*) and hornbeam (*Carpinus betulus*), marked with big empty circles in Fig. 4. The undergrowth with numerous young lime- and hornbeam-trees (with a trunk 2 to 15 cm in diameter, marked in Fig. 4 with small empty circles). The forest floor covered with numerous

trunks of dead trees (*Quercus*, *Tilia*, *Carpinus* and one *Pinus sylvestris* barkless trunk) and littler of branches and twigs (marked in grey in Fig. 4). Phytosociological analysis is given in Table 2.

The data on soil pH are shown in Table 1.

Locality 2 (plots III, IV). The locality is situated in an exploited forest area, forest division 447, on the left from the Niepolomice–Ispina road.

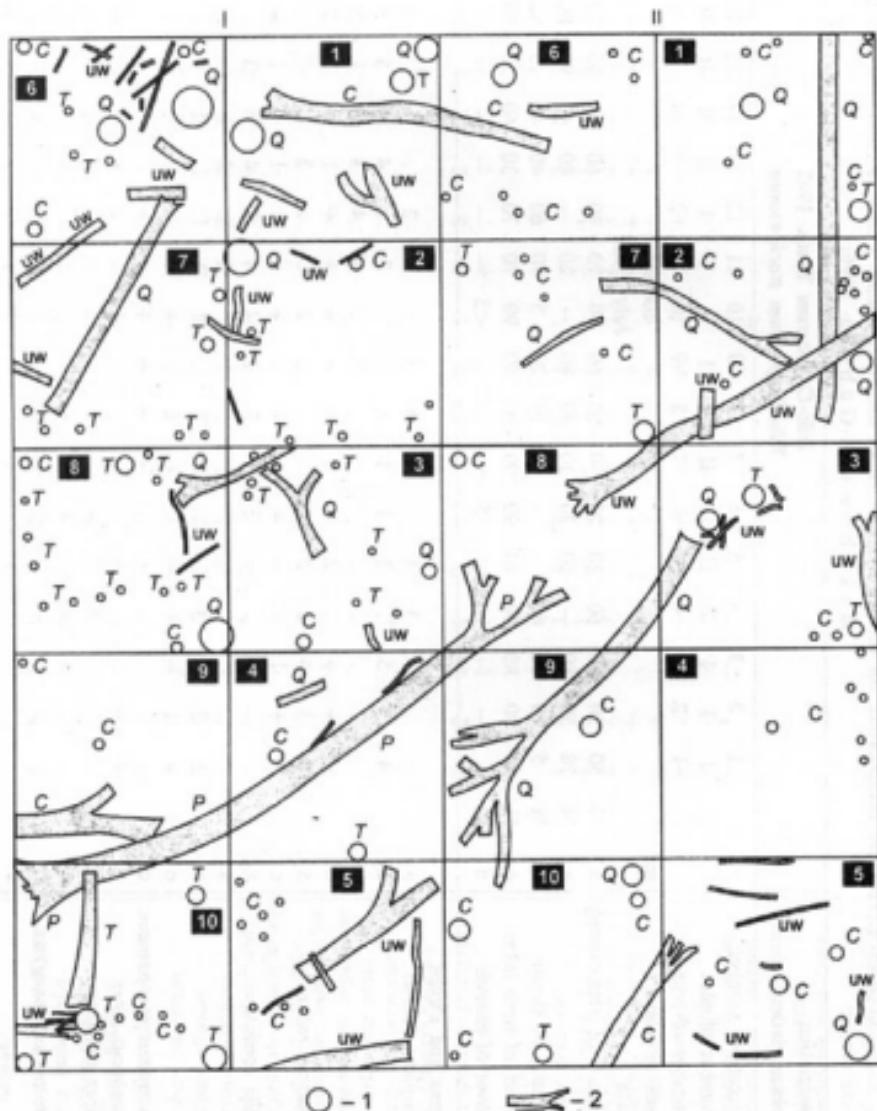


Fig. 4. Structure of the tree stand in the locality 1, plots I (subplots 1–10) and II (subplots 1–10). 1 – living trees, 2 – lying trunks and fallen branches, C – *Carpinus betulus*, P – *Pinus sylvestris*, Q – *Quercus robur*, T – *Tilia cordata*, uw – unidentified wood

Table 2
Locality 1. Floristic composition on the plots I and II, in the Niepolomice Forest, section 460, the Lipówka reserve
(Jakubowska-Gabar 1996)

These plots are located at a small distance from each other (Fig. 5). The forest consisting mainly of oak (*Quercus robur*), lime (*Tilia cordata*), and hornbeam (*Carpinus betulus*), is dominated by old oak-trees. Coniferous trees are missing, except old *Pinus sylvestris* specimens occurring individually. Unlike previous plots, these plots are located in the forest, which is still exploited (during the studies the traces of forest workers' activities were noticeable — single cuts, forest floor clearings). There are no dead trees in the forest floor; fallen branches

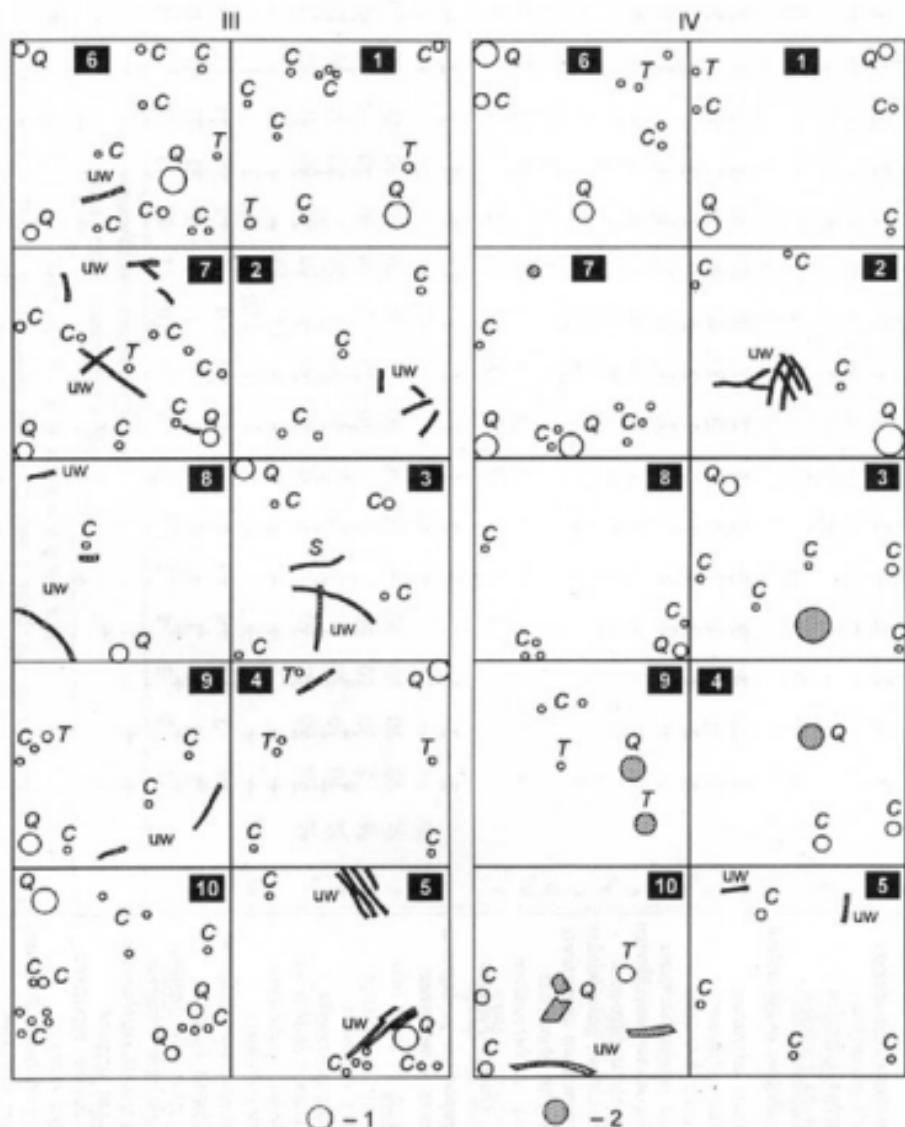


Fig. 5. Structure of the tree stand in the locality 2, plots III (subplots 1–10) and IV (subplots 1–10). 1 – living trees, 2 – dead stumps, fallen branches and twigs, C – *Carpinus betulus*, Q – *Quercus robur*, T – *Tilia cordata*, uw – unidentified wood

Table 3
Locality 2. Floristic composition on the plots III and IV, in the Niepolomice Forest, section 447 (Jakubowska-Gabara 1996)

Tilio-Carpinetum Tracz. 1962
Tilio-Carpinetum stackylosum

Tab. 3 cont.

	1	2	3
<i>Stachys sylvatica</i>	-	-	1
<i>Festuca gigantea</i>	+	1	2
<i>Ch. Fagellata sylvaticae</i>	-	-	1
<i>Milium effusum</i>	-	-	.
<i>Gallium odoratum</i>	-	-	V
<i>Viola reichenbachiana</i>	-	3	2
<i>Circaea latifolia</i>	-	2	2
<i>Polygonatum multiflorum</i>	-	1	IV
<i>Ranunculus lanuginosus</i>	-	1	II
<i>Carex sylvatica</i>	-	1	1
<i>Scrophularia nodosa</i>	-	1	1
<i>Brachypodium sylvaticum</i>	-	1	1
<i>Eurythodium angustifolium</i>	-	1	1
<i>Ch. Querceto-Fagetea</i>	-	1	V
<i>Aegopodium podagraria</i>	-	2	IV
<i>Anemone nemorosa</i>	-	2	IV
Others			
<i>Carex brizoides</i>	1	2	2
<i>Oxalis acetosella</i>	+	1	1
<i>Majanthemum bifolium</i>	+	1	1
<i>Meehringia trinervia</i>	+	1	1
<i>Urtica dioica</i>	+	1	1
<i>Athyrium filix-femina</i>	+	1	1
<i>Glechoma hederacea</i>	+	1	2
<i>Galeopsis bifida</i>	-	1	1
<i>Cardamine impatiens</i>	-	1	1
<i>Stellaria nemorum</i>	-	1	1
<i>Geum urbanum</i>	-	1	1
<i>Dryopteris carthusiana</i>	-	1	1
<i>Aluga reptans</i>	-	1	1
<i>Veronica montana</i>	-	1	1
<i>Galeopsis pubescens</i>	-	1	1
<i>Poa trivialis</i>	-	1	1
<i>Rubus plicatus</i>	-	1	1
<i>Laportea communis</i>	-	1	1

are scarce. In one of the subplots a heap of branches accumulated but it was removed after some time. Dead wood was present only in the form of small twigs or single stumps (from 55 to 110 cm in diameter — marked in Fig. 5 with grey circles). A phytosociological analysis is given in Table 3.

The data on soil pH are shown in Table 1.

RESULTS

S y s t e m a t i c a n a l y s i s. 274 species and one variety were collected from all study plots: 16 species of *Ascomycota* and 258 of *Basidiomycota*. The numbers of fungi in systematic groups (orders) are given in Table 4. The most abundant were the following genera: *Mycena* — 20, *Psathyrella* — 14, *Clitocybe*, *Pluteus* and *Phanerochaete* — 7 each, *Collybia*, *Hyphoderma* and *Russula* — 6 each, *Galerina*, *Athelia*, *Peniophora* — 5 each, *Hypholoma*, *Lepista*, *Marasmius*, *Stereum* — 4 species each. The list of species sampled in selected plots over 3 years is given in Tables 5 to 8.

Table 4
Fungi in systematic groups according to Hawksworth et al. (1995)

Phylum	Order	Number of species
<i>Ascomycota</i>	<i>Hypocreales</i>	2
	<i>Leotiales</i>	6
	<i>Pezizales</i>	2
	<i>Xylariales</i>	6
<i>Basidiomycota</i>	<i>Agaricales</i>	115
	<i>Atractiellales</i>	1
	<i>Boletales</i>	7
	<i>Cantharellales</i>	2
	<i>Cortinariales</i>	11
	<i>Dacryomycetales</i>	4
	<i>Fistulinales</i>	1
	<i>Ganodermatales</i>	1
	<i>Gomphales</i>	1
	<i>Hericiales</i>	1
	<i>Hymenochaetales</i>	6
	<i>Lycoperdales</i>	2
	<i>Nidulariales</i>	1
	<i>Phallales</i>	1
	<i>Porales</i>	15
	<i>Russulales</i>	10
	<i>Schizophyllales</i>	1
	<i>Sclerodermatales</i>	2
	<i>Stereales</i>	61
	<i>Thelephorales</i>	4
	<i>Tremellales</i>	9
	<i>Tulasnellales</i>	2
Total:		274

Table 5

Synoptical fungus table of the plot I
Dey—number of fruit-bodies, SF—spatial frequency, TF—total temporal frequency and MDev—maximum number of fruit-bodies during one visit
on one subplot

Years	Number of observations	1994			1995			1996			1994–1996		
		9	7	62	10	114	168	26	168	168	SF	TF	
<i>Agrocybe praecox</i>		Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	MDev	Dey	
<i>Amanita fulva</i>	1	1	1					15	2	2	15	12	2
<i>A. vaginata</i>								1	1	1	1	1	1
<i>Armillaria mellea</i> s. l.	1	1	1					5	1	1	6	5	2
<i>Ascocoryne sarcoides</i>	35	3	2					26	2	3	61	20	4
<i>Athelia decipiens</i>	3	2	1								3	2	1
<i>A. epiphylla</i>	1	1	1								1	1	1
<i>Bjerkandera adusta</i>				167	3	4	1073	4	10	1240	200	6	14
<i>Boletus pulverulentus</i>	1	1	1					8	2	4	9	2	3
<i>Botryosphaeridium laeve</i>	1	1	1	14	1	3	10	1	1	25	10	2	5
<i>B. stabcoronatum</i>	1	1	1	9	2	3	5	1	1	15	5	3	5
<i>Brevicillium olivaceum</i>	1	1	1								1	1	1
<i>Bulgaria inquinans</i>	3	1	1								3	3	1
<i>Bystroporus corynii</i>	59	5	2	30	1	2	64	2	3	153	40	5	7
<i>Calocera cornea</i>	85	1	5								85	50	1
<i>Calocybe carneola</i>	1	1	1								1	1	1
<i>Ciboria batschiana</i>								112	5	1	112	80	5

<i>Clavaria cinerea</i>	13	1	2			10	3	1	23	7	4	3	
<i>Clioocybe clavipes</i>	22	5	2	1	1	13	3	2	36	6	5	5	
<i>C. gibba</i>	17	4	3			18	6	3	35	9	7	6	
<i>C. lignatilis</i>	6	2	1						6	5	2	1	
<i>C. odora</i>	1	1	1						1	1	1	1	
<i>C. sp.</i>						4	3	1	4	2	3	1	
<i>Collybia butyracea</i> = <i>C. asteia</i>	295	9	5	6	2	1	98	9	3	399	77	10	9
<i>C. dryophila</i>	111	9	7	12	4	2	51	10	6	174	14	10	15
<i>C. erythropus</i> = <i>C. maramoides</i>	21	2	1						21	13	2	1	
<i>C. fuscopurpurea</i>						2	1	1	2	2	1	1	
<i>C. fusipes</i>	2	1	1			22	2	3	24	13	2	4	
<i>C. peronata</i>	4	1	2						4	3	1	2	
<i>Coniolephora arida</i>	9	2	4	2	1	1	10	1	1	21	10	3	6
<i>C. paleana</i>	3	2	2						3	2	2	2	
<i>Conocybe semiglobata</i>						1	1	1	1	1	1	1	
<i>C. tenera</i>						1	1	1	1	1	1	1	
<i>Corinarius</i> sp.	6	2	2						6	3	2	2	
<i>Crepidotus variabilis</i>	370	5	6	42	2	1	11	1	2	423	80	5	9
<i>Cyathus striatus</i>	92	2	6						92	22	2	6	
<i>Cylindrobasidium evolvens</i>	295	3	2			20	2	1	315	200	4	3	
<i>Dactylospores minor</i>	10	1	1	10	1	1	10	1	1	30	10	2	3
<i>D. stillans</i>	60	1	1						60	60	1	1	
<i>D. tortus</i>				3	1	1			3	3	1	1	

Tab. 5 cont.

Years	Number of observations	1994			1995			1996			1994-1996			
		Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	MDev	SF	TF
<i>Delicatula integriflora</i>								2	1	1	2	2	1	1
<i>Ecteoloma</i> sp.								18	5	2	18	5	5	2
<i>Exidia glandulosa</i> = <i>E. truncata</i>	48	1	5								48	16	1	5
<i>E. plana</i>				2	1	1					2	2	1	1
<i>Fistulina hepatica</i>							2	1	1	2	2	1	1	1
<i>Galerina cerina</i>							6	1	1	6	6	1	1	1
<i>G. fallax</i>							33	2	2	33	15	2	2	2
<i>G. triscopa</i>							11	1	1	11	11	1	1	1
<i>Grandospora litschense</i>	2	1	9	2	1	2	62	4	5	66	13	4	16	
<i>Hapalopilus rutilans</i>	1	1	1							1	1	1	1	1
<i>Hohenbuehelia myxotricha</i>	46	2	2				5	1	1	51	35	3	3	
<i>Hymenochaete erinaceomarginata</i>							15	1	2	15	10	1	2	
<i>H. rubiginosa</i>	800	2	9	1953	2	7	5230	2	10	7983	500	2	26	
<i>Hymenocystis calculus</i>	16	1	1							16	16	1	1	
<i>Hypoderma mutatum</i>	8	2	2							8	5	2	2	
<i>H. pallidum</i>							10	1	1	10	10	1	1	
<i>H. praetermittens</i>	1	1	1	10	1	2	10	1	1	21	10	2	4	
<i>H. piserium</i>	11	2	3	9	2	1				20	5	3	4	
<i>H. setigerum</i>				25	1	2				25	20	1	2	

Tab. 5 cont.

Years	Number of observations	1994						1995						1996						1994-1996					
		9			7			10			114			10			26			168					
Number of species		Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	M	Dey	SF	TF					
<i>Mycena crocea</i>								2	1	1	2	1	1	2	1	1	1	1	1	1	1	1	1		
<i>M. galericulata</i>					8	3	1	11	4	4	19	6	5	11	4	4	19	6	5	5	5	5	5		
<i>M. galopus</i>	54	6	4					61	8	6	115	24	9	10											
<i>M. galopus</i> var. <i>nigra</i>	10	1	1					2	2	1	2	1	2	1	2	1	2	1	2	1	2	1	1		
<i>M. haematocephalus</i>	54	2	2					26	2	1	80	50	30	3	26	2	1	80	50	30	3	3	3		
<i>M. leptocephala</i>								2	2	1	2	1	2	1	2	1	2	1	2	1	2	1	1		
<i>M. pellucida</i>	1	1	1	1	1	1	1	1	1	1	3	1	1	3	1	1	3	1	1	2	3	3	3		
<i>M. pura</i>	199	7	5	11	4	1	36	5	4	246	45	7	10	199	7	5	11	4	246	45	7	10	10		
<i>M. sanguinolenta</i>	51	7	3	8	2	1	15	3	4	74	10	8	8	51	7	3	8	2	74	10	8	8	8		
<i>M. stylobates</i>	133	9	3					1	1	1	134	50	9	4	133	9	3	8	2	134	50	9	4	4	
<i>M. subcaerulea</i>	2	2	1					4	2	2	6	3	3	3	2	2	6	3	3	3	3	3	3		
<i>M. vitilis</i>	3	3	1	2	1	1	20	4	3	25	10	6	5	3	20	4	3	25	10	6	5	5	5		
<i>M. zebrinus</i>	72	5	4	13	2	1	36	3	2	121	14	5	7	72	5	4	13	2	121	14	5	7	7		
<i>Nectria bohemica</i>								1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1		
<i>Nectria cinnabarina</i>	400	2	1								400	300	200	200	400	2	1	1	1	1	1	1	1		
<i>Oligoporus fragilis</i>								5	1	1	5	5	5	5	5	1	1	5	5	5	1	1	1		
<i>O. leucomallellus</i>								5	1	1	5	5	5	5	5	1	1	5	5	5	1	1	1		
<i>O. subcaesius</i>	18	1	1					12	2	2	30	18	3	3	12	2	2	30	18	3	3	3	3		

Tab. 5 cont.

Number of species	1994			1995			1996			1997			
	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	MDev	SF	TF
<i>Phlebia nemorum</i>				1	1	1				1	1	1	1
<i>Polyporus badius</i>				13	2	3	12	1	5	25	7	3	8
<i>P. brumalis</i>	33	3	4	6	1	1	3	1	1	42	13	3	6
<i>P. varius</i>							1	1	1	1	1	1	1
<i>Psathyrella gracilis</i>				1	1	1				1	1	1	1
<i>P. impexa</i>	3	1	1							3	3	1	1
<i>P. panamensisoides</i>	1	1	1							1	1	1	1
<i>P. spadiceogrisea</i>	3	1	1				4	1	1	7	4	2	2
<i>Psilocybe inquilina</i> var. <i>crotalaria</i>	1	1	1				14	2	2	15	9	3	3
<i>P. squamata</i>	19	6	3	15	6	1	33	6	1	57	12	10	5
<i>Ramicola haustellaris</i>							1	1	1	1	1	1	1
<i>R. laevigata</i>	2	1	1							2	2	1	1
<i>Rickenella fibula</i>	3	1	1	2	1	1	3	1	1	8	3	1	3
<i>Ripartites metzelli</i>	14	2	2							14	12	2	2
<i>R. tricholoma</i>	18	3	2				6	4	1	24	7	5	3
<i>Russula cyanoxantha</i>	4	1	1				10	6	4	14	4	6	5
<i>R. nigricans</i>	1	1	1				1	1	1	2	1	2	2
<i>R. ochroleuca</i>	12	3	2				13	5	1	25	7	6	3
<i>R. pectinatoides</i>							8	3	3	8	3	3	3

<i>R. violipes</i>	1	1	1				20	5	2	21	9	5	3
<i>Schizophyllum commune</i>	39	2	4	11	2	2					50	14	4
<i>Schizopora flavigpora</i>	5	1	1	2	1	1	35	3	2	42	20	3	4
<i>S. paradoxus</i> s. l.	132	4	9	6	2	2	227	7	5	365	100	10	16
<i>Scopuloides hydnoides</i>							2	1	1	2	2	1	1
<i>Steccherinum ochraceum</i>	15	1	1	20	1	1				35	20	2	2
<i>Stereum gracipinatum</i>	35	1	2							35	20	1	2
<i>S. hirsutum</i>	4790	6	9	3190	7	7	3731	8	10	11711	500	9	26
<i>S. rameale</i> = <i>S. ochraceoflavum</i>							280	1	3	280	120	1	3
<i>Sistropharia aeruginosa</i>				4	1	1				4	4	1	1
<i>S. cinnerea</i> = <i>S. cyanea</i>	2	1	1							2	2	1	1
<i>Thelophora terrestris</i>							3	1	1	3	3	1	1
<i>Tomentella fuscoferruginea</i>	2	1	1							2	2	1	1
<i>T. sp.</i>	2	2	1							2	1	2	1
<i>Trametes versicolor</i>	4	1	1				80	1	1	84	80	2	2
<i>Trechispora farinacea</i>							10	1	1	10	10	1	1
<i>Tremella mesenterica</i>	11	3	3	7	1	1	15	1	2	33	10	4	6
<i>Tulostoma thelephoreoides</i>				1	1	1				1	1	1	1
<i>Wulfenia conidea</i>	5	2	1				5	1	1	10	5	3	2
<i>Xerocomus chrysenteron</i>	25	7	6	36	6	2	15	9	4	76	18	10	12
<i>Xerula radicata</i>	1	1	1				1			1	1	1	2
<i>Xylaria hypoxylon</i>				41	3	2	68	3	2	109	40	4	4
<i>X. longipes</i>	442	4	8	430	4	5	173	4	7	1045	100	5	20
<i>Xylolobus frustulatus</i>	3	1	9	88	1	7	660	1	10	751	120	1	26

Table 6

Synoptical fungus table of the plot II
Dey - number of fruit-bodies, SF - spatial frequency, TF - total temporal frequency and MDev - maximum number of fruit bodies during one visit
on one subplot

Years	Number of species	1994			1995			1996			1994-1996		
		Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	MDev	SF	TF
<i>Agrocybe praetox</i>					3	1	1	11	3	2	14	8	4
<i>Amillaria mellea</i> s.l.	9	3	1	68	2	1	100	1	1	177	100	3	3
<i>Ascocoryne sarcoides</i>	20	2	1							20	10	2	1
<i>Athelia alnicola</i>	3	1	1							3	3	1	1
<i>A. decipiens</i>	2	1	1							2	2	1	1
<i>A. epiphylla</i>	1	1	1	3	1	1				4	3	2	2
<i>Basidiobolus caesiolichenum</i>							10	1	1	10	10	1	1
<i>Bjerkandera adusta</i>	13	3	4	27	3	3	87	4	5	127	20	5	12
<i>Bryothrixidium laeve</i>	3	2	2	9	3	3	7	3	2	19	5	5	7
<i>B. subcoronatum</i>				8	3	2	35	2	4	43	10	5	6
<i>Brevicillium olivaceum</i>							2	1	1	2	2	1	1
<i>Bystomanellus corium</i>	56	6	4	5	1	1	28	3	4	89	10	6	9
<i>Calocera cornea</i>	155	1	8	5	1	1	3	1	1	163	50	2	10
<i>Ciboria batschiana</i>	21	2	1				578	6	2	599	300	7	3
<i>Clavulinopsis cinerea</i>							47	7	3	47	11	7	3
<i>C. coralloides</i>								2	1	1	2	2	1
<i>Clitocybe canescens</i>									1	1	1	1	1

<i>Clitocybe clavipes</i>	18	4	2			6	3	3	24	7	5	5
<i>C. gibba</i>	24	5	3			25	5	4	49	10	7	7
<i>C. lignicola</i>	2	1	1						2	2	1	1
<i>C. odora</i>	1	1	1						1	1	1	1
<i>Collybia butyracea</i> = <i>C. aescina</i>	416	8	4	1	1	181	8	2	597	90	9	7
<i>C. dryophila</i>	80	9	6	33	7	3	102	10	6	215	26	10
<i>C. personata</i>	19	3	3	4	1	1	25	4	3	48	6	4
<i>Coniochaeta arida</i>	9	3	1	3	1	2	23	4	4	35	10	5
<i>C. pulicaria</i>							5	1	1	5	5	1
<i>Crepidotus variabilis</i>	331	3	7	116	3	3	74	2	4	521	58	4
<i>Cyathus striatus</i>	34	3	4				281	2	3	315	120	4
<i>Cylindrobasidium evolvens</i>	162	3	2	3	1	1	30	1	1	195	150	4
<i>Dacrymyces minor</i>	10	1	1	40	2	2				50	20	3
<i>Diodella querina</i>	9	1	9	5	1	1	7	1	1	21	9	1
<i>Dentipellis fragilis</i>							2	1	1	2	2	1
<i>Entoloma</i> sp.	1	1	1			10	4	2	11	3	5	3
<i>Exidia glandulosa</i> = <i>E. truncata</i>	13	1	2						13	10	1	2
<i>E. plana</i>	348	2	4	66	3	3	40	3	3	454	200	4
<i>Fomes fomentarius</i>	1	1	9						1	1	1	9
<i>Galerina alpinotincta</i>						17	2	2	17	12	2	2
<i>G. cerina</i>						40	2	1	40	25	2	1
<i>G. fallax</i>						7	1	1	7	7	1	1
<i>G. marginata</i>	2	1	1						2	2	1	1

Tab. 6 cont.

Number of species	Years			1994			1995			1996			1994-1996		
	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	SF	MDev	SF	TF	
<i>Gonosperma lippiense</i>		117			62			114					26		
<i>Hypolepis rutilans</i>	1	1	1				18	2	3	19	12	3	4		
<i>Hoheria beccariana myxotricha</i>	34	2	2							34	29	2	2		
<i>Hymenochete rubiginosa</i>	470	5	9	520	3	7	835	5	8	1825	250	7	24		
<i>Hymenoscyphus calyculus</i>	30	1	1				50	1	1	80	50	2	2		
<i>H. fructigenus</i>	10	1	1							10	10	1	1		
<i>Hypoderma praevertissum</i>	1	1	1				20	1	1	21	20	2	2		
<i>H. puberum</i>	1	1	1							1	1	1	1		
<i>H. setigerum</i>	2	1	1				12	2	2	14	10	2	3		
<i>Hypodontia quercina</i>							1	1	1	1	1	1	1		
<i>H. sambuci</i>							2	1	1	2	2	1	1		
<i>Hypoholoma fasciculare</i>				32	1	2	25	2	2	57	30	2	4		
<i>H. subviride</i>	420	3	8	214	3	4	379	4	7	1013	100	4	19		
<i>Hypochnium geogenium</i>	2	1	1							2	2	1	1		
<i>Hypoxylon deustum</i>							10	1	1	10	10	1	1		
<i>H. fasciatum</i>	230	2	2							230	100	2	2		
<i>H. howeanum</i>	380	1	7	140	2	2	180	3	4	700	80	5	13		
<i>Kuehneromyces mutabilis</i>										2	2	1	1		
<i>Laccaria laccata</i>	5	3	3				207	10	4	212	33	10	7		

<i>Lactarius camphoratus</i>							12	1	2	12	6	1	2
<i>L. quietus</i>	1	1	1				17	6	3	18	3	6	4
<i>Laetiporus sulphureus</i>	60	1	7							60	15	1	7
<i>Lepista flaccida</i> = <i>L. inversa</i> = <i>Clitocybe inversa</i>	100	8	4				2	2	2	102	14	9	6
<i>L. gilva</i>							3	2	1	3	2	2	1
<i>L. mada</i>	27	2	2	3	1	1	2	1	1	32	8	4	4
<i>Lycoperdon perlatum</i>	3	2	2	2	1	1	21	4	2	26	10	7	5
<i>L. pyriforme</i>	35	3	4	38	1	1	279	1	6	352	100	3	11
<i>Macrolepia proceria</i>	6	3	2	2	1	1	1	1	1	9	2	4	4
<i>M. rhinodes</i>	6	3	2	1	1	1				7	3	3	3
<i>Marasmius bulliardii</i>				34	1	1	1	1	1	35	34	2	2
<i>M. querophila</i>	10	1	1							10	10	1	1
<i>M. rotula</i>				5	2	1	7	3	3	12	4	4	4
<i>M. torquescens</i>	3	1	1				4	1	1	7	4	2	2
<i>Megacollybia platyphylla</i>	37	9	4	12	4	3	45	9	7	94	14	10	14
<i>Mycena abramissii</i>							3	1	1	3	3	1	1
<i>M. annicta</i>	1	1	1							1	1	1	1
<i>M. filipes</i>							2	1	1	2	2	1	1
<i>M. galericulata</i>				1	1	1	3	2	2	4	2	3	3
<i>M. galopus</i>	20	7	3	15	4	2	51	9	6	86	11	10	11
<i>M. haematocephalus</i>	9	2	2							9	8	2	2
<i>M. inclinata</i>	1019	4	4	333	3	2	174	4	3	1526	430	5	9
<i>M. para</i>	188	10	5	35	7	1	27	7	4	250	43	10	10

Tab. 6 cont.

Years	Number of observations	1994						1995						1996						1994-1996		
		Dcy	SF	TF	Dcy	SF	TF	Dcy	SF	TF	Dcy	SF	TF	Dcy	SF	TF	MDev	SF	TF			
<i>Mycena sanguinolenta</i>	47	9	4	4	2	1	11	5	3	62	11	9	8									
<i>M. stylobates</i>	62	7	2				2	4	1	64	39	9	3									
<i>M. subcaerulea</i>	16	3	1	1	1	1	17	5	4	34	8	6	6									
<i>M. vitilis</i>	8	4	2	6	3	2	13	6	3	27	3	9	7									
<i>M. zephirus</i>								2	1	1	2	2	1									
<i>Mycenella salicina</i>								2	1	1	2	2	1									
<i>Naucoria spadicea</i>	1	1	1					100	1	1	100	100	1									
<i>Necritia ctenopharina</i>											1	1	1									
<i>Oligoporus leucotrichellus</i>								44	1	4	44	27	1	4								
<i>O. subelegans</i>	3	1	1								3	3	1	1								
<i>Omphalina</i> sp.	1	1	1								1	1	1	1								
<i>Paxillus involutus</i>				2	1	1	1	1	1	3	2	1	2									
<i>Peniophora cinerea</i>	79	4	6	54	3	3	170	4	4	303	100	8	13									
<i>P. querina</i>	29	2	2	13	1	1	25	2	2	67	20	4	5									
<i>P. rufomarginata</i>	18	2	1				170	3	3	188	100	5	4									
<i>Peziza varia</i>	1	1	1							1	1	1	1									
<i>P.</i> sp.	3	1	1							3	3	1	1									
<i>Phallus impudicus</i>				1	1	1	2	2	2	3	1	2	3									
<i>Phanerochaete laevigata</i>							13	2	2	13	8	2	2									

<i>P. marcelliana</i>	4	1	1					8	1	1	12	8	2	2
<i>P. sordida</i>	4	1	1					14	3	2	14	8	3	2
<i>P. tuberculata</i>														
<i>P. velutina</i>	2	1	1	1	1	1	1	4	4	3	7	2	4	5
<i>Phellinus contiguus</i>	3	3	4	19	2	5	55	4	5	77	30	6	14	
<i>P. nigricans</i>	1	1	9	2	1	6	2	1	10	5	2	1	25	
<i>Phlebiella pseudosugaæ</i>	5	2	2	2	1	1	25	2	2	32	10	3	5	
<i>P. vaga</i>	5	1	1							5	5	1	1	
<i>P. sp.</i>				10	1	1				10	10	1	1	
<i>Phleogenia faginea</i>	230	1	3	300	1	1	300	1	3	830	300	1	7	
<i>Pholiota flammans</i>				2	1	1				2	2	1	1	
<i>Pluteus cervinus</i>	7	3	2	1	1	1	6	4	2	14	3	6	5	
<i>P. inquinans</i>	1	1	1					1	1	1	1	1	1	
<i>P. pellitus</i>								2	1	1	2	2	1	
<i>P. phlebophorus</i>										2	2	1	1	
<i>P. salicinus</i>	2	1	1							1	1	1	1	
<i>Polyporus badius</i>	2	1	2					1	1	1	3	1	2	3
<i>Psathyrella chondroderma</i>	3	1	1							3	3	1	1	
<i>P. fuliginea</i>									1	1	1	1	1	
<i>P. gossypina</i>									1	1	1	1	1	
<i>P. obtusata</i>	3	2	1							3	2	2	1	
<i>P. piliformis</i>	5	1	1							5	5	1	1	
<i>P. semex</i>	1	1	1							1	1	1	1	

Tab. 6 cont.

Number of species	1994			1995			1996			1994-1996		
	9	7	10	114	114	10	114	114	10	26	168	26
<i>Pseathyrella spadicogeisea</i>	117	62	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey
<i>P. spadicogeisea f. vernalis</i>			3	1	1				11	2	2	11
<i>Pseudodictyotye cyathiformis</i>	2	1	1									3
<i>Psilocybe ingens</i> var. <i>croatica</i>	1	1	1				1	1	1	2	1	1
<i>P. squamosa</i>	14	6	3	60	7	1	73	8	1	147	29	10
<i>Radulomyces confluens</i>	27	3	2				25	2	1	52	20	4
<i>R. molaris</i>							12	1	2	12	10	1
<i>R. rickii</i>				10	1	1				10	10	1
<i>Ramicola centunculus</i>				8	1	1				8	8	1
<i>Rickenella fibula</i>	19	2	2				3	1	1	22	8	3
<i>R. setipes</i> = <i>R. swartzii</i>							5	1	1	5	5	1
<i>Ripartites metrodii</i>	3	2	2							3	2	2
<i>R. tricholoma</i>	1	1	1							1	1	1
<i>Russula cyanoxantha</i>	17	4	4				23	6	4	40	6	6
<i>R. ochroleuca</i>	3	3	2				11	3	2	14	6	5
<i>R. pectinatoides</i>							11	3	3	11	7	3
<i>R. violaceipes</i>							5	4	3	5	1	4
<i>Schizophyllum commune</i>				23	1	1				23	23	1

<i>Schizopora flavigena</i>	1	1	1	5	1	1	30	2	3	36	10	3	5
<i>S. paradoxxa</i> s. l.	318	7	9	251	5	7	585	9	9	1154	100	10	25
<i>Sistotrema brinkmannii</i>	1	1	1						1	1	1	1	1
<i>Steccherinum ochraceum</i>	23	2	2	67	2	4			90	22	4	6	
<i>Stereum hirsutum</i>	1340	8	9	1460	7	7	3200	9	10	6000	200	10	26
<i>S. rameale</i> = <i>S. ochraceoflavum</i>	65	3	2	50	1	1			115	50	4	3	
<i>Stropharia aeruginosa</i>	3	2	1				3	2	1	6	2	3	2
<i>S. caerulea</i> = <i>S. cyanea</i>	6	3	3						6	3	3	3	
<i>Tomentella neoboudotii</i>							1	1	1	1	1	1	1
<i>Trametes versicolor</i>							500	1	2	500	300	1	2
<i>Trechispora farinacea</i>				3	1	1	5	1	1	8	5	2	2
<i>T. mollifexa</i>							19	3	4	19	5	3	4
<i>Tremella matenterica</i>	33	4	5				3	1	2	36	7	5	7
<i>Tubaria furfuracea</i>				6	1	1				6	6	1	1
<i>Tulostoma eichleri</i>	3	1	1							3	3	1	1
<i>Vullemnia comedens</i>							34	6	4	34	5	6	4
<i>Xerocomus chrysenteron</i>	47	10	6	63	10	3	30	8	4	140	10	10	13
<i>Xerula radicata</i>	1	1	1				1	1	1	2	1	2	2
<i>Xylaria hypoxylon</i>				332	6	3	525	7	5	857	150	10	8
<i>X. longipes</i>	417	6	9	323	9	4	187		5	927	84	9	18

Table 7

Synoptical fungus table of the plot III

Dey - number of fruit-bodies, SF - spatial frequency, TF - total temporal frequency and MDcv - maximum number of fruit-bodies during one visit on one subplot

Years	Number of observations	1994			1995			1996			1994-1996			
		Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	MDcv	SF	TF
Number of species		79		59		76		10		26		125		
<i>Armillaria mellea</i> s. l.	2	2	1								2	1	2	1
<i>Athelia epiphylla</i>				2	1	1					2	2	1	1
<i>Basidiocarpon radula</i>	3	1	2	35	2	1	1	1	1	39	30	3	4	
<i>Bjerkandera adusta</i>	4	1	2				23	2	3	27	15	2	5	
<i>Boletus pulverulentus</i>							2	2	2	2	1	2	2	
<i>Boryodasism compactum</i>			1	1	1						1	1	1	1
<i>Byssomerulius corium</i>	98	4	5				62	3	4	160	30	7	9	
<i>Ciboria batckiana</i>				9	1	1					9	9	1	1
<i>Cladulinia cinerea</i>	9	2	1				2	1	1	11	7	2	2	
<i>C. coralloides</i>							2	1	1	2	2	1	1	
<i>Clitocybe candidula</i>	1	1	1				3	2	1	4	2	3	2	
<i>C. clavipes</i>	3	2	1							3	2	2	1	
<i>C. gibba</i>							17	5	2	17	8	5	2	
<i>C. hygrophoroides</i> = <i>C. phaeophila</i>				3	2	1	100	8	1	103	37	8	2	
<i>C. metachroa</i>								1	1	1	1	1	1	
<i>C. sp.</i>								5	1	1	5	5	1	1
<i>Collybia butyracea</i> = <i>C. asema</i>	119	10	5	2	2	1	69	8	3	190	24	10	9	

<i>C. dryophila</i>	191	10	6	24	8	3	67	9	5	282	25	10	14
<i>C. fusipes</i>	1	1	1	47	1	2	13	1	1	61	35	2	4
<i>C. personata</i>	19	6	4	11	1	2	104	5	4	134	22	6	10
<i>Coniophora erida</i>	3	1	1				10	1	1	13	10	2	2
<i>Coprinus micaceus</i>							14	1	1	14	1	1	1
<i>C. sp.</i>	5	1	1							5	5	1	1
<i>Crepidotus variabilis</i>	72	2	2	20	1	1	35	1	2	127	60	2	5
<i>Cyathus striatus</i>	158	4	6	86	2	2				244	90	4	8
<i>Dacrymyces minor</i>				17	2	1				17	12	2	1
<i>Dacrydium quercina</i>	20	1	9				1	1	1	21	20	1	10
<i>Eatableia sp.</i>	2	2	2				2	2	1	4	1	4	3
<i>Exidia glandulosa</i> = <i>E. truncata</i>				5	1	1				5	5	1	1
<i>E. plana</i>							30	1	2	30	20	1	2
<i>E. sp.</i>				1	1	1				1	1	1	1
<i>Hymenochaete rubiginosa</i>				150	1	3	320	1	9	470	100	1	12
<i>Hymenocystus fructigenus</i>	10	1	1							10	10	1	1
<i>Hypoderma argillaceum</i>	1	1	1							1	1	1	1
<i>H. militaris</i>							25	1	2	25	20	1	2
<i>H. pteretmissum</i>	1	1	1	17	3	3	2	1	1	20	10	4	5
<i>H. puberum</i>	1	1	1							1	1	1	1
<i>H. setigerum</i>				56	2	2				56	30	2	2
<i>Hypodontia sambuci</i>	26	2	2	10	1	1	20	1	2	56	20	3	5
<i>Hypodoma fasciculare</i>				3	1	1				3	1	1	1

Tab. 7 cont.

Number of species	Years		1994		1995		1996		1994–1996			
	Number of observations		9		7		10		26			
	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	MDev	SF	TF
<i>Hypoleoma subviride</i>				7	1	1	2	1	1	9	7	1
<i>Hypoxyylon howeanum</i>	395	3	3	1110	3	6	960	1	10	2465	350	4
<i>Inonotus radiatus</i>	30	1	2							30	20	1
<i>Laccaria laccata</i>	13	5	4				56	9	4	69	10	9
<i>Lactarius quietus</i>	5	2	3				17	7	2	22	7	8
<i>Lepista flaccida</i> = <i>L. inversa</i> = <i>Clitocybe inversa</i>	48	5	4				13	4	2	61	9	7
<i>L. galva</i>							45	7	1	45	20	7
<i>L. meda</i>	62	3	3	1	1	1	32	5	1	95	26	6
<i>Lycoperdon perlatum</i>	7	2	3				2	1	1	9	2	2
<i>L. pyriforme</i>	14	2	2							14	10	2
<i>Macrocyptula proceria</i>	39	7	4	7	4	2	4	3	2	50	7	9
<i>M. rhacodes</i>	28	7	3	3	1	1	4	4	2	35	17	8
<i>Marasmius bulliardii</i>	332	5	3	11	1	1				343	134	5
<i>M. quercoophilus</i>	100	2	1							100	97	2
<i>M. rotula</i>							25	1	1	25	25	1
<i>Megacollybia platyphylla</i>	9	4	4	6	1	1	5	3	3	20	6	8
<i>Mollisia</i> sp.				500	1	1				500	500	1
<i>Mycena galericulata</i>							1	1	1	1	1	1

<i>M. galopus</i>	107	10	3	3	1	1	27	7	4	137	17	10	8
<i>M. galopus</i> var. <i>nigra</i>							1	1	1	1	1	1	1
<i>M. incisa</i>	710	2	2				1	1	1	710	490	2	2
<i>M. macror</i>													
<i>M. pelizaeana</i>	1	1	1	1	1	1				2	1	2	2
<i>M. pallidula</i>							1	1	1	1	1	1	1
<i>M. pura</i>	14	3	4				8	5	3	22	4	7	7
<i>M. rotunda</i>	1	1	1							1	1	1	1
<i>M. sanguinolenta</i>	140	10	2	4	2	2	13	5	2	157	22	10	6
<i>M. stylolobates</i>	127	10	3				2	2	2	129	23	10	5
<i>M. vitidis</i>	2	1	1	2	1	2	7	6	2	11	2	6	5
<i>M. zephirus</i>	2	1	2				2	1	1	4	2	1	3
<i>Nectria cinnabarina</i>	50	1	1				20	1	1	70	50	2	2
<i>N.</i> sp.							100	1	1	100	100	1	1
<i>Oligoporus subelegans</i>	2	1	2							2	1	1	2
<i>Peniophora cinerea</i>	205	7	4	226	6	4	360	7	5	791	100	8	13
<i>P. incarnata</i>							1	1	1	1	1	1	1
<i>P. quercina</i>	140	4	4	63	4	3	30	2	1	233	50	8	8
<i>P. rufomarginata</i>				10	1	1				10	10	1	1
<i>Phanerochaete laevigata</i>	1	1	1							1	1	1	1
<i>P. velutina</i>				8	1	1				8	8	1	1
<i>P.</i> sp.				4	1	1				4	4	1	1
<i>Phellinus contiguus</i>	5	3	8	75	6	5	5	2	2	85	10	7	15

Tab. 7 cont.

Number of species	1994			1995			1996			1994-1996		
	9	117	62	7	10	114	10	116	26	1994	1995	1996
<i>Phellinus robustus</i>	2	2	8	2	2	6				4	1	2
<i>Phlebia radiata</i>							40	2	1	40	20	2
<i>P. rufa</i>				5	1	1	5	1	1	10	5	2
<i>P. tremellosa</i>	45	1	3	1	1	1	1	1	1	47	22	2
<i>Phlebiella vagia</i>	9	1	2				10	1	1	19	10	2
<i>Pholiota lenta</i>	31	3	2				8	2	1	39	15	4
<i>Pluteus cervinus</i>	1	1	1				1	1	1	2	1	2
<i>Panzarella condolleana</i>				1	1	1				1	1	1
<i>P. obtusata</i>	4	1	1							4	4	1
<i>P. ocellata</i>	3	1	1							3	3	1
<i>P. spadiceogrisea</i>	5	2	2							5	2	2
<i>Patinocybe squamosa</i>	69	7	5	29	5	1	14	4	1	112	16	8
<i>Radulomyces confluens</i>	50	2	4	28	2	2	10	1	1	88	25	4
<i>R. molaris</i>	10	1	1	1	1	1				11	10	2
<i>R. rickii</i>				2	1	1				2	2	1
<i>Ripartites metrodii</i>	13	4	3							13	5	4
<i>R. tricholoma</i>							3	2	1	3	2	2
<i>Rosellinia aquila</i>	15	1	1							15	15	1
<i>Russula cyanoxantha</i>	9	2	2	1	1	1	1	1	1	11	8	4

<i>R. ochroleuca</i>							5	1	1	5	3	1	1
<i>R. petimbeoides</i>							4	3	2	4	1	3	2
<i>R. violaceus</i>	1	1	1	2	2	1	17	7	2	20	5	7	4
<i>Schizophyllum commune</i>							6	1	2	6	3	1	2
<i>Schizophora flavipora</i>				3	2	2				3	2	2	2
<i>S. paradoxus</i> s. l.	341	10	9	527	9	7	334	8	8	1202	90	10	24
<i>Scleroderma areolatum</i>							4	1	1	4	4	1	1
<i>Sebacina calospora</i>				1	1	1				1	1	1	1
<i>Sistotrema brinkmannii</i>	1	1	1							1	1	1	1
<i>S. oblongisporum</i>	7	2	3	2	1	1				9	3	3	4
<i>Sistotremastrum niveocreatum</i>	5	1	1	2	1	1				7	5	2	2
<i>Steccherinum ochraceum</i>	11	2	2				70	3	2	81	20	5	4
<i>Stereum griseopallidum</i>	65	1	3							65	30	1	3
<i>S. hirsutum</i>	30	2	2	22	3	3	10	1	2	62	20	4	7
<i>Stropharia aeruginosa</i>	3	1	1				2	1	1	5	3	2	2
<i>S. caerulea</i> = <i>S. cyanea</i>	6	3	2							6	2	3	2
<i>Subulicystidium longisporum</i>				4	1	2				4	3	1	2
<i>Stypella grilletti</i> = <i>Myxarium podlachicum</i>				2	1	1				2	2	1	1
<i>Trechispora farinacea</i>	3	1	1							3	3	1	1
<i>Tremella mesenterica</i>	2	1	1							2	2	1	1
<i>Tubaria conspersa</i>	1	1	1							1	1	1	1
<i>Vullemnia comedens</i>	42	7	4	18	5	2	70	5	4	130	20	9	10
<i>Xerocomus chrysenteron</i>	44	9	5	29	6	1	41	8	5	114	11	10	11
<i>Xylaria hypoxylon</i>	200	3	4	36	2	2	200	4	7	436	30	7	13
<i>X. longipes</i>	7	2	2	8	1	1	13	1	2	28	10	3	5

Table 8

Synoptical fungus table of the plot IV
Dey - number of fruit-bodies, SF - spatial frequency, TF - total temporal frequency and MDev - maximum number of fruit-bodies during one visit
on one subplot

Years	1994			1995			1996			1994-1996			
Number of observations	9			7			10			26			
Number of species	101			46			79			137			
	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	MDev	SF	TF
<i>Agaricus sylvicola</i> = <i>A. exetei</i> = <i>A. abribulbus</i>	3	1	1							3	3	1	1
<i>Armillaria mellea</i> s. l.	27	4	2							27	20	4	2
<i>Athelia acrospora</i>	3	1	1							3	3	1	1
<i>A. alnicola</i>	3	1	1							3	3	1	1
<i>A. salicinum</i>	1	1	1							1	1	1	1
<i>Bastardodendron caesiocineraceum</i>	6	1	2	12	1	2	5	1	1	23	10	1	5
<i>Botryobasidium laeve</i>	2	1	1	2	1	1				4	2	2	2
<i>B. subcoronatum</i>		4	1	1						4	4	1	1
<i>Byssomerulius cortium</i>	45	6	4				129	6	6	174	20	8	10
<i>Ciboria batschiana</i>	13	2	1	4	1	1	11	1	1	28	11	3	3
<i>Clavulina ciberea</i>	6	1	2				6	3	2	12	4	4	4
<i>C. coralloides</i>	1	1	1							1	1	1	1
<i>Clitocybe campestris</i>							2	1	1	2	2	1	1
<i>C. clavipes</i>	4	1	2				11	2	2	15	8	2	4
<i>C. gibba</i>	2	2	1				14	5	3	16	6	5	4

<i>C. metachroa</i>						14	3	1	14	8	3	1
<i>C. sp.</i>						72	7	1	72	22	7	1
<i>Collybia butyracea</i> = <i>C. astema</i>	337	10	5	1	1	141	9	3	479	50	10	9
<i>C. dryophila</i>	29	6	6	13	5	3	32	9	6	74	8	10
<i>C. fusipes</i>							4	1	1	4	4	1
<i>C. peronata</i>	43	7	5	26	2	2	69	5	3	138	23	7
<i>Conocybe tenera</i>	2	1	1							2	2	1
<i>Corinarius cfr. alnetorum</i>						2	1	1	2	2	1	1
<i>C. sp.</i>	22	3	1						22	10	3	1
<i>Crepidotus variabilis</i>	278	4	5	22	2	3	37	3	2	337	60	6
<i>Cyathus striatus</i>	75	2	5			10	1	1	85	10	3	6
<i>Dactylospores minor</i>	40	1	1	10	1	1			50	40	2	2
<i>D. stipitatus</i>	30	1	1						30	30	1	1
<i>Entoloma sp.</i>	3	2	1			4	2	2	7	2	4	3
<i>Gymnopilus strobilis</i>	53	1	1						53	53	1	1
<i>Haplopilus rutilans</i>						4	1	1	4	4	1	1
<i>Hebeloma claviceps</i>						1	1	1	1	1	1	1
<i>H. clavulipes</i>	2	2	1						2	1	2	1
<i>H. sacchariolens</i>	12	1	1						12	9	1	1
<i>Hymenochaete rubiginosa</i>				40	1	2	50	1	1	90	50	1
<i>Hymenocystiphorus calyculus</i>	45	2	2						45	25	2	2
<i>Hypoderma praetermicatum</i>	10	1	1	5	1	1	20	1	1	35	20	3
<i>H. puberum</i>	1	1	1	3	1	1			4	3	1	2

Tab. 8 cont.

Years	1994	1995	1996	1994–1996
Number of observations	9	7	10	26
Number of species	117	62	114	168
<i>Hyphodontia pallidula</i>				
<i>Hypholoma capnoides</i>	2	1	1	
<i>H. fasciculare</i>				
<i>H. lateritium</i>	9	2	2	
<i>H. subvirens</i>	106	1	100	
<i>Hypochnicium eriksonii</i>	3	1	2	
<i>H. geogenium</i>	7	1	1	
<i>Hypoxyylon howeanum</i>	365	2	7	
<i>Inocybe geophylla</i> var. <i>halimae</i>	3	2	2	
<i>I. quietoides</i>				
<i>Laccaria amethystina</i>				
<i>L. laccata</i>	87	6	4	
<i>Lactarius quietus</i>	3	2	2	
<i>Lepista flaccida</i> = <i>L. inversa</i> = <i>Clitocybe inversa</i>	16	4	2	
<i>L. gibba</i>				
<i>L. irina</i>	14	1	1	
<i>L. munda</i>	51	6	2	

<i>Lycoperdon perlatum</i>	2	1	1			16	3	3	18	8	4	4
<i>L. pyriforme</i>	1006	4	6	103	1	2	318	2	7	1427	200	5
<i>Macrolepista proceră</i>	3	3	6	5	1					9	2	5
<i>M. rhacodes</i>	5	2	3			6	2	1	11	5	3	4
<i>Marasmiellus ramulosus</i>						7	1	1	7	1	1	1
<i>Marasmius bellardii</i>	18	3	2						18	7	3	2
<i>Marasmius quercoophilus</i>	4	1	1						4	1	1	1
<i>M. rotula</i>				5	1	1			5	5	1	1
<i>Megacollybia platyphylla</i>	10	4	3	6	2	1	15	7	3	31	4	9
<i>Mycena amicta</i>	1	1	1						1	1	1	1
<i>M. halibut</i>						1	1	1	1	1	1	1
<i>M. filipes</i>						1	1	1	1	1	1	1
<i>M. galericulata</i>						10	1	1	10	10	1	1
<i>M. galopus</i>	94	10	3	12	2	1	22	8	4	128	13	10
<i>M. inclinata</i>	32	1	1	13	1	1				45	32	2
<i>M. leptocephala</i>	2	1	1							2	2	1
<i>M. petaliphila</i>	1	1	1						1	1	1	1
<i>M. pura</i>	14	6	4	1	1	1	2	2	17	4	6	7
<i>M. rorida</i>	5	3	1						5	3	3	1
<i>M. sanguinolenta</i>	120	9	2	1	1	48	4	3	169	32	9	6
<i>M. stylidites</i>	286	9	2			17	4	2	303	70	10	4
<i>M. vitilis</i>	21	8	4	3	2	2	7	3	2	31	3	9
<i>M. zephirus</i>	38	2	3			18	1	1	56	30	2	4

Tab. 8 cont.

<i>P. spadiceogrisea</i>			1	1	1			1	1	1	1	1
<i>Pilozyme inqualina</i> var. <i>crobelatus</i>			2	1	1	1	1	3	1	2	2	2
<i>P. squamulosa</i>	54	6	4	11	5	1	19	7	1	84	21	8
<i>Radiomyces confusa</i>	111	5	4	14	2	1	33	3	3	158	25	6
<i>R. molaris</i>	23	3	3							23	10	3
<i>Ramaria eumorpha</i>	3	1	1							3	1	1
<i>Ramicola centunculus</i>			11	2	1					11	9	2
<i>Ripartites metrodii</i>	1	1	1							1	1	1
<i>R. tricholoma</i>	7	4	2				2	2	1	9	2	6
<i>Russula cyanoxantha</i>	31	6	3				6	4	4	37	3	8
<i>R. foetens</i>							1	1	1	1	1	1
<i>R. nigricans</i>							3	1	1	3	1	1
<i>R. ochroleuca</i>							2	1	2	2	1	1
<i>R. pettinatoidea</i>							11	5	3	11	3	5
<i>R. violipes</i>	7	3	2	4	1	1	26	7	2	37	9	7
<i>Schizophyllum commune</i>	30	1	2				40	1	2	70	20	2
<i>Schizophyllum flavipora</i>	6	1	1							6	6	1
<i>S. paradoxata</i> s. l.	434	8	9	318	8	7	253	6	6	1005	55	9
<i>Scleroderma bovista</i>	1	1	1							1	1	1
<i>Scopuloides hydnoides</i>							1	1	1	1	1	1
<i>Sebastia incrustans</i>							33	3	1	33	20	3
<i>Sistotrema brinkmannii</i>	2	2	1							2	1	2
<i>S. oblongisporum</i>	1	1	1							1	1	1

Tab. 8 cont.

Number of observations	1994			1995			1996			1994-1995			
	Dey	SF	TF	Dey	SF	TF	Dey	SF	TF	Dey	MDcv	SF	TF
<i>Scletofucus nivea</i>	5	1	2							5	4	1	2
<i>Steccherinum ochraceum</i>	15	1	1	110	1	3	15	1	1	140	70	1	5
<i>Stereum gausapatum</i>							20	1	1	20	20	1	1
<i>S. sanguinolentum</i>	5	1	1							5	5	1	1
<i>Strobilurus tenacellus</i>							2	1	2	2	2	1	2
<i>Stropharia aeruginosa</i>							1	1	1	1	1	1	1
<i>Tomentella sp.</i>	2	2	1							2	1	2	1
<i>Trechispora farinacea</i>	4	1	1							4	4	1	1
<i>Tremella encephala</i>	5	1	1							5	5	1	1
<i>T. mesenterica</i>	14	3	2				3	1	1	17	7	3	3
<i>Tubaria conspersa</i>	4	1	1							4	4	1	1
<i>Vauillainia comedens</i>	16	4	4	14	3	2	53	7	5	83	13	8	11
<i>Xerocomus chrysenteron</i>	48	10	4	57	8	1	33	9	4	138	14	10	9
<i>X. rubellus</i>	1	1	1							1	1	1	1
<i>X. subtomentosum</i>							1	1	1	1	1	1	1
<i>Xerula padens</i>							1	1	1	1	1	1	1
<i>Xylaria hypoxylon</i>	100	3	4	90	2	2	102	2	3	292	60	5	9
<i>X. longipes</i>	12	1	1	12	1	1	47	3	4	71	12	4	6

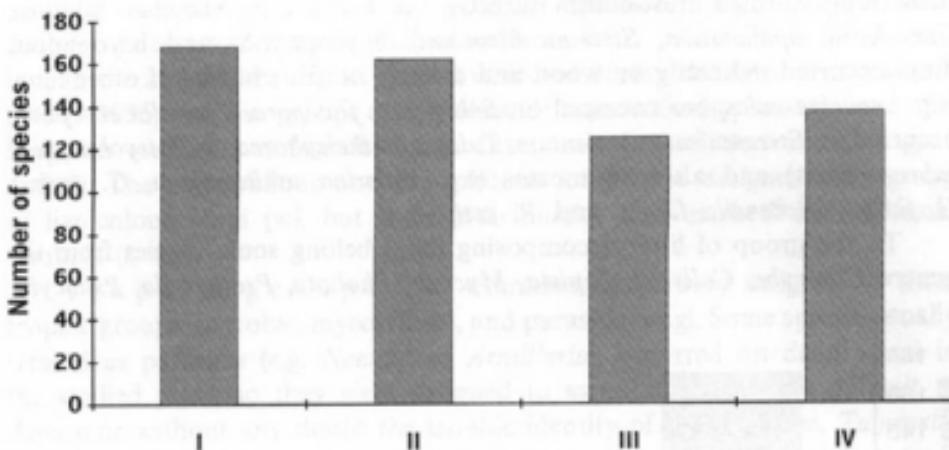


Fig. 6. Number of fungi collected during the years 1994–1996 on the plots I–IV

The richest in species were plots I (168 species) and II (162 species), slightly less abundant in species were plots III (125) and IV (137) – Fig. 6.

In the studied plots 16 species new to Poland were found. These were: *Athelia acrospora*, *A. decipiens*, *Galerina cerina*, *Hebeloma clavulipes*, *Inocybe quietiodor*, *Mycena subcaerulea*, *Mycenella salicina*, *Phanerochaete martelliana*, *Psathyrella impexa*, *P. ocellata*, *P. senex*, *Ramicola laevigata*, *Sebacina calospora*, *Sistotrema oblongisporum*, *Tomentella fuscoferruginea*, and *T. neobourdotii*.

A part of these species are rare or very rare species for Poland. *Mycena bulbosa* is known in Poland from the Niepolomice Forest only (Komorowska 1991). Some of them, e.g. *Athelia salicum*, *Stypella grilletii* and *Tulasnella eichleriana* had not been recorded in Poland for over one hundred years. These species were collected for the last time in the environs of Międzyrzec Podlaski in northeastern Poland by B. Eichler (Bresadola 1903).

E c o l o g i c a l a n a l y s i s . For each plot the following data were calculated: the total numbers of fruit-bodies of each species in the subsequent years (Dcy), the numbers of subplots in which fruit-bodies were recorded in each year (SF), the numbers of visits during which a species was recorded (TF), and maximum numbers of fruit-bodies recorded during one visit (MDcv). The results are given in Tables 5 to 8.

S u b s t r a t e . During each visit on the plot data on direct and indirect substrate of each fungus were recorded. Fungi formed fruit-bodies on wood, on litter (on plant fragments), on the ground, and some (infrequently) on fruit-bodies of other fungi and mosses growing on wood.

Lignicolous fungi occurred on standing, fallen, live, and dead logs and on hanging as well as fallen, dead or live, tree and shrub branches. The majority of these fungi formed fruit-bodies directly on wood (e.g. *Fistulina hepatica*, *Ganoderma applanatum*, *Stereum hirsutum*). Mycophilous and bryophilous fungi occurred indirectly on wood and directly on fruit bodies of other fungi (e.g. *Sebacina calospora* occurred on *Schizophora flavipora*, *Tremella encephala* occurred on *Stereum sanguinolentum*, *Tulasnella thelephorea* on *Botryobasidium subcoronatum*) and also on mosses (e.g. *Galerina atkinsoniana*, *G. cerina*, *G. fallax*, *Rickenella fibula*, and *R. setipes*).

To the group of litter-decomposing fungi belong some species from the genera *Clitocybe*, *Collybia*, *Lepista*, *Mycena*, *Pholiota*, *Psathyrella*, *Psilocybe*,

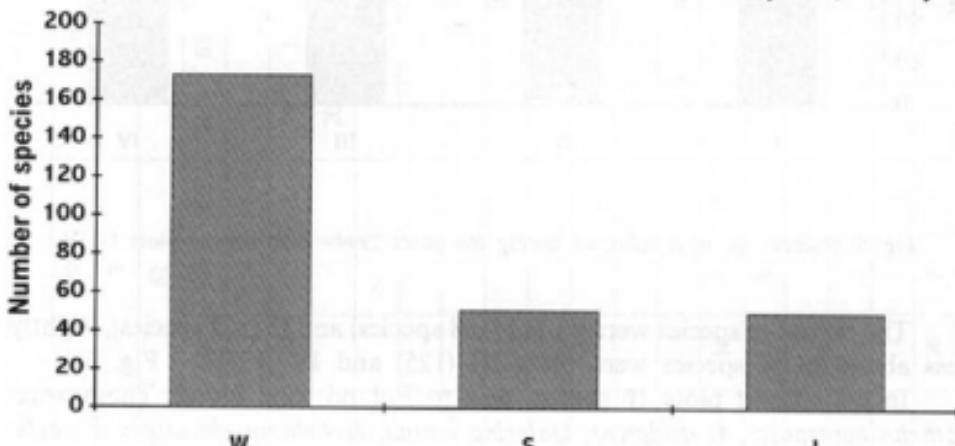


Fig. 7. Total number of fungal species on different substrates: I—litter, S—soil, W—wood

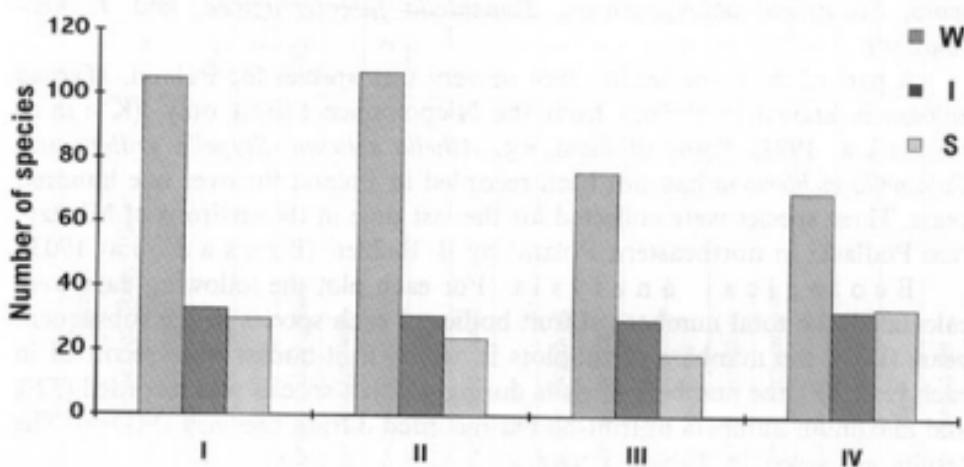


Fig. 8. Number of fungal species on each plot on different substrates: I—litter, S—soil, W—wood

and *Strobilurus*. The last genus in the present study has been assigned to litter and not wood.

On bare ground occur fruit-bodies of terrestrial fungi, e.g. *Hebeloma*, *Inocybe*, *Laccaria*, *Lactarius*, and *Russula*.

In the alphabetical list of species collected during the three-year research (1994–1996) trophic groups are indicated: mycorrhizal, parasitic, saprobic fungi, as well as a substrate: fungi, litter, mosses, soil and wood (Figs 7 and 8). The mycophilous and bryophilous fungi were assigned to a group of lignicolous fungi (w), but their individuality was marked with different symbols (f, m).

Trophic groups. The examined fungi were assigned to three trophic groups: saprobic, mycorrhizal, and parasitic fungi. Some species usually treated as parasites (e.g. *Nectria* or *Armillaria*), occurred on dead wood in the studied plots, so they were assigned to saprobic fungi. It is difficult to determine without any doubt the trophic identity of some species. *Tulasnella thelephorea* grows on wood mostly together with corticoid fungi (e.g. from the genus *Botryobasidium*), but it is unknown whether it is a parasite on these fungi or it decomposes their dying fruit-bodies, or at last, whether it decomposes wood.

The numbers of fungi occurring in the above-given groups in all plots are given in Fig. 9, while individual numbers of fungi for each of the four plots are given in Fig. 10.

Saprobes were the most abundant fungi in the collected material and their total number reached 234, there were also 33 mycorrhizal, and as few as 7 parasitic fungi found in this study. The most abundant in saprobes were plots I (144) and II (148), while mycorrhizal fungi were most numerous in plot IV (22) – Figs 9 and 10.

Weather impact. The numbers of recorded species were changing over the three years of studies. In the first year, in spite of the fact that studies were launched as late as in July, the highest number of 193 species was recorded. In the second year the number of species was 118, and in the third year it was 184 for all the plots. The richest in species was September 1994. The number of species for all the plots was 134, they were equally abundant in October: 105 (Fig. 11). In the plots I and II subsequently 64 and 68 species were recorded (Fig. 12). September was the richest in species also in the later years (Fig. 11). The numbers of fungi in particular plots differed from each other, for example in July 1996, the plots I and II had respectively 51 and 53 species, in September plot III had 41 species and in October plot IV had 36 species (Fig. 12). The comparison of these numbers with meteorological data demonstrates dependence of the number of species upon the amount of rainfall. Similar dependencies can be noticed between the numbers of fructifications and the amount of rainfall.

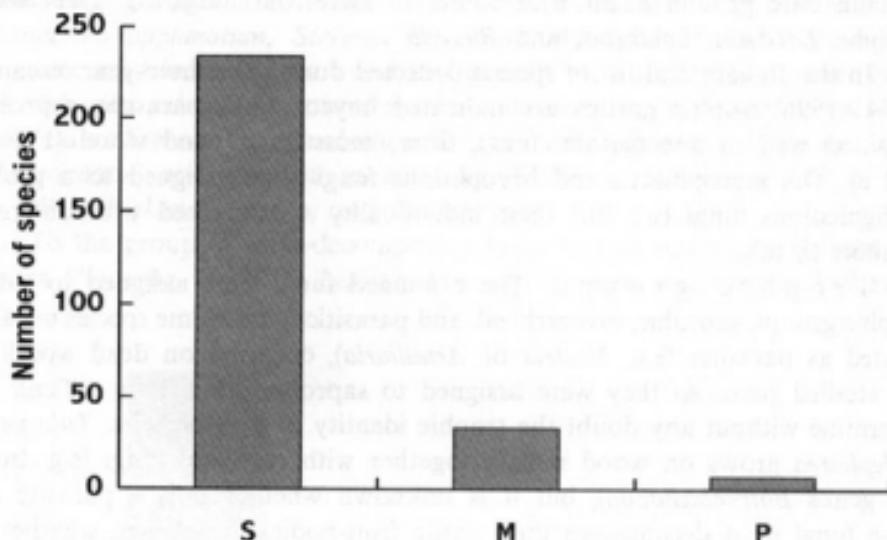


Fig. 9. Total number of species in the trophic groups of fungi: M—mycorrhizal, P—parasitic, S—saprobic

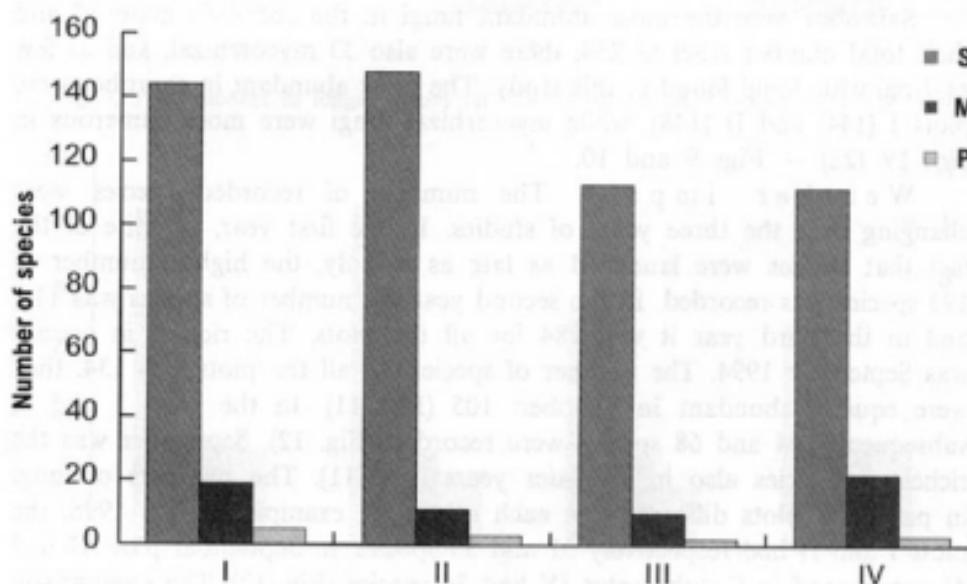


Fig. 10. Number of species on each plot in the trophic groups of fungi: M—mycorrhizal, P—parasitic, S—saprobic

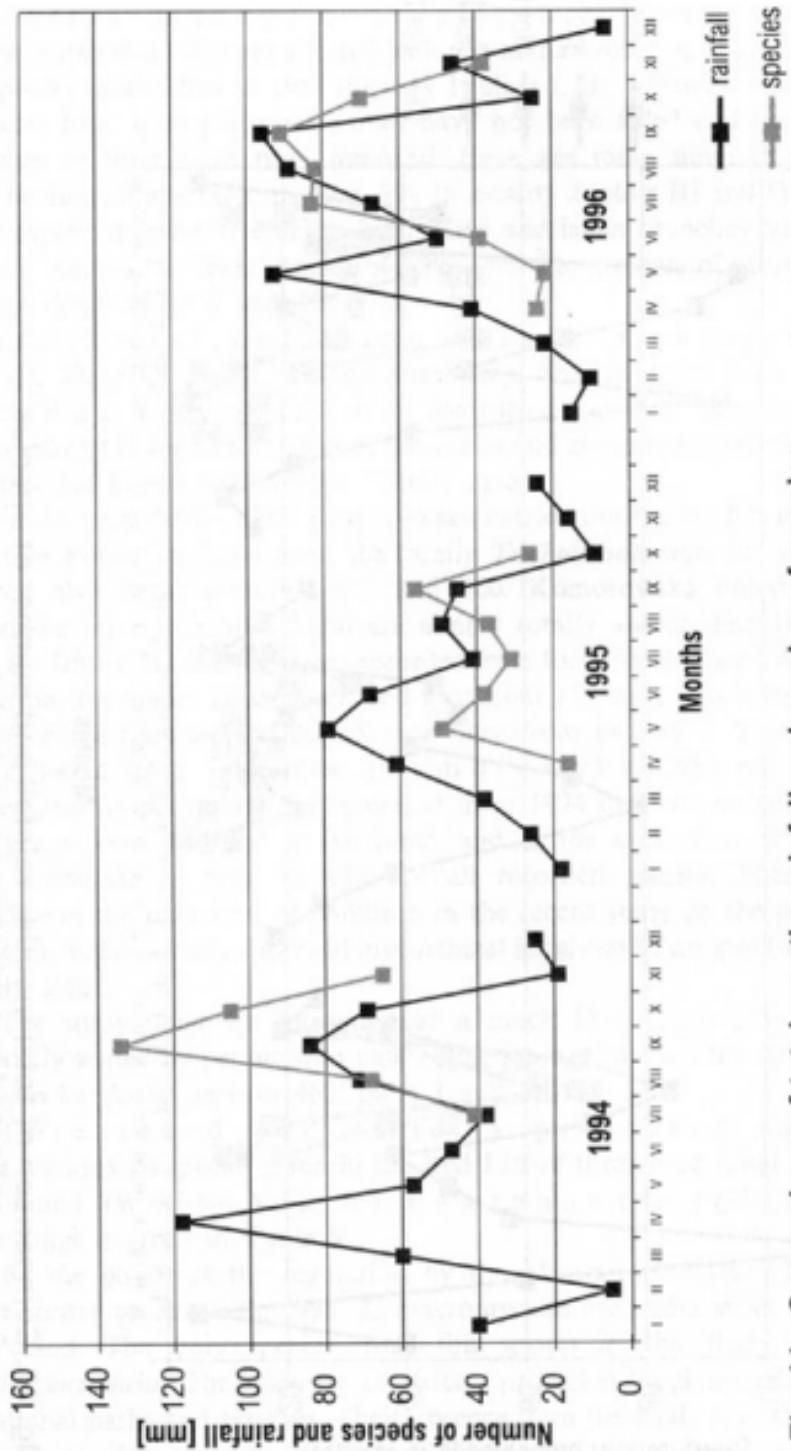


Fig. 11. Comparison of total months rainfall and number of species collected in each year on all plots.

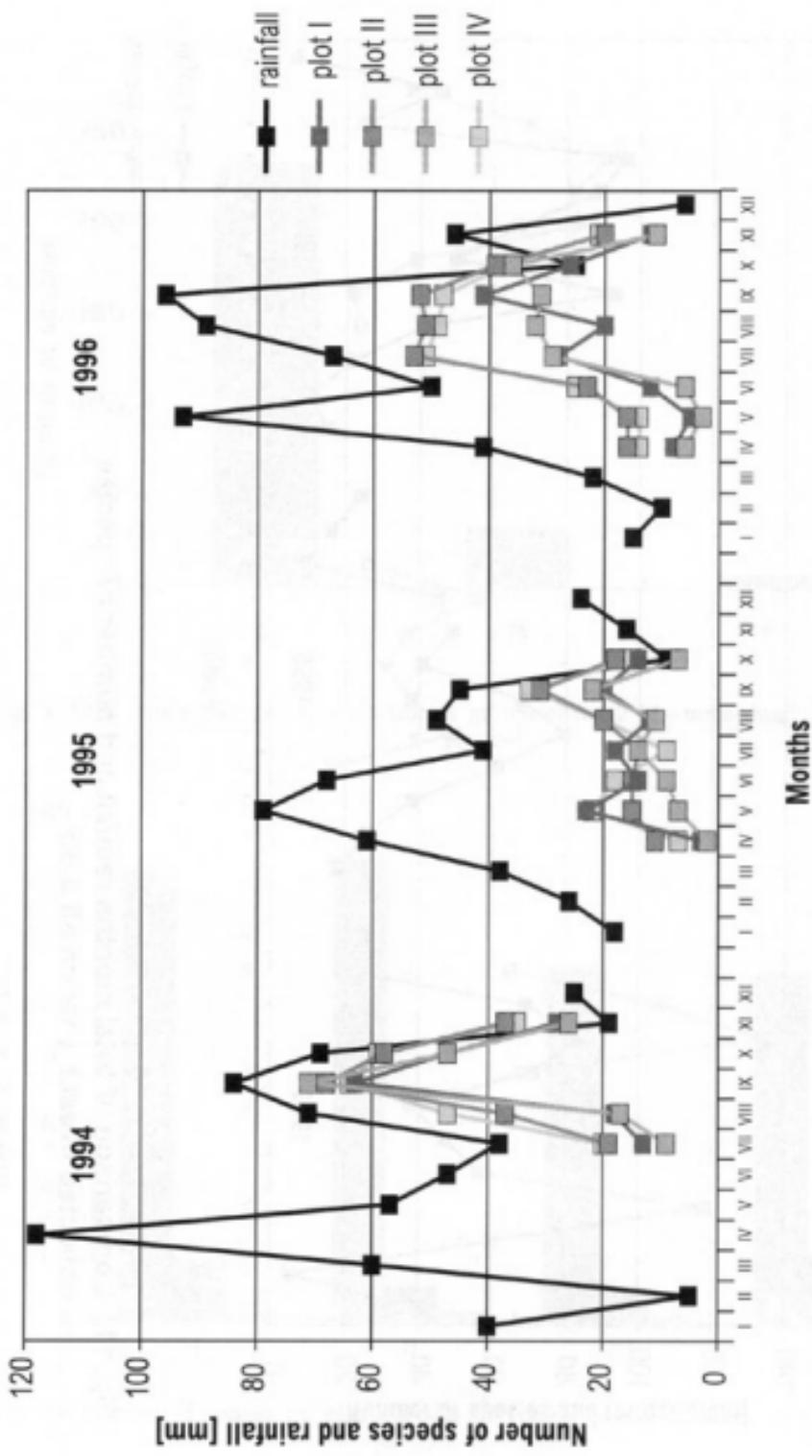


Fig. 12. Comparison of total monthly rainfall and number of species collected in each year on each plot.

F o r e s t m a n a g e m e n t e f f e c t. The present studies have demonstrated that fungi are affected by forest management. In the strict reserve of Lipówka established in 1958 (locality 1; plots I, II), where no management measures have been performed, trees have not been felled and neither logs, branches or litter have been removed, there are more fungi (Fig. 6) and saprobic fungi are in majority (Fig. 10). In locality 2 (plots III and IV), located in an exploited forest, trees have been felled and fallen branches removed. In this site there occur fewer species of fungi and the numbers of saprobic fungi are also fewer (Figs 6 and 10).

I n d u s t r i a l p o l l u t i o n v e r s u s p i c k i n g e d i b l e f u n g i. Only 33 species of mycorrhizal fungi were recorded from the study plots, which is as little as 12% of all recorded fungi. This situation may be explained by the long-term effect of industrial and communal pollution on the Niepolomice Forest (see chapter "Study area").

In the years 1976–1980 Komorowska carried out the studies in the Niepolomice Forest on fungi from the family *Tricholomataceae*, the study area covered also forest divisions 447 and 460. Komorowska noted that the fruit-bodies of mycorrhizal fungi are almost totally absent. For 166 species from the family *Tricholomataceae* recorded from the Niepolomice Forest, three were from the genus *Tricholoma*, and four from *Laccaria*. *Tricholoma fulvum* and *T. imbricatum* were collected only once from locality 1. *T. sulphureum* was collected from 7 localities (Komorowska 1986 msc.). During the monitoring performed in the period from 1994 to 1996, no species from that genus were recorded in the plots, and at the same time, mycorrhizal fungi constitute as little as 12% of all recorded species. Therefore, no influence of the reduction of pollution in the recent years on the production of spores, and especially spores of mycorrhizal fungi can be noticed (see chapter "Study area").

The study plots are located near a much busy road. They are also frequently visited by people who pick edible fungi, also from the strict reserve of Lipówka (forest section 460, plots I and II).

T h r e a t e n e d a n d s t r i c t l y p r o t e c t e d f u n g i. In the study area 24 species given in the Red List of threatened fungi in Poland were found (Woje woda and Ławrynowicz 1992). The list of these fungi is given in Table 9.

By the power of the regulation by the Minister of Nature Protection and Forestry on June 4th, 1995, 22 macromycetes are under strict protection in Poland. The only species from this group in the study area was *Phallus impudicus*. The majority of strictly protected fungi occur in Poland in national parks and reserves. Their absence from the study area demonstrates unfavourable changes in the environment of the Niepolomice Forest, as a result of which fungi with greater ecological demand are being eliminated.

Table 9

Fungi of the Red List of threatened fungi in Poland, collected on investigated plots in the Niepołomice Forest

Species	Plots				The red data book categories			
	I	II	III	IV	E	V	R	I
1. <i>Botryobasidium laeve</i>	+	+		+			+	
2. <i>Clitocybe candicans</i>		+	+	+				+
3. <i>Dacryomyces tortus</i>	+							+
4. <i>Fistulina hepatica</i>	+					+		
5. <i>Galerina triscopa</i>	+							+
6. <i>Hebeloma claviceps</i>				+			+	
7. <i>Hohenbuehelia myxotricha</i>	+	+					+	
8. <i>Hymenochaete cinnamomea</i>	+							+
9. <i>Macrolepiota procera</i>	+	+	+	+				+
10. <i>Marasmius bulliardii</i>	+	+	+	+				+
11. <i>Mycena abramsii</i>	+	+					+	
12. <i>M. crocata</i>	+						+	
13. <i>M. pelianthina</i>	+		+	+				+
14. <i>M. pullata</i>			+			+		
15. <i>Phlebia rufa</i>			+					+
16. <i>Phleogenia faginea</i>	+	+			+			
17. <i>Physisporinus vitreus</i>	+						+	
18. <i>Psilocybe squamosa</i>	+	+	+	+				+
19. <i>Radulomyces molaris</i>	+	+	+	+			+	
20. <i>Russula violeipes</i>	+	+	+	+			+	
21. <i>Stypella grilletii</i>			+		+			
22. <i>Trechispora mollusca</i>		+					+	
23. <i>Xerula pudens</i>				+		+		
24. <i>Xylobolus frustulatus</i>	+						+	
Total:	17	11	10	10	2	4	9	9

MONITORING DATA FROM OTHER REGIONS OF POLAND

Mycorrhizal fungi. As mentioned above, the studies performed in the Niepołomice Forest were a part of monitoring studies made in forests with the proportion of oak in different regions of Poland. Results of the present studies may be compared with the already published data. The comparison of mycorrhizal data seems especially interesting (Fig. 13). In the Białowieża National Park (Skirgielio 1998b) 45 species of mycorrhizal fungi were found in the study area, which makes about 21% of all fungi found there. In Wielkopolska (Lisiewska and Polczyńska 1998) in the Dębina reserve 43 mycorrhizal fungi were recorded, which makes 20% of all fungi found there. In the Świętokrzyskie Mountains (Łuszczynski 1998) the highest number of 58 species of mycorrhizal fungi was recorded which makes 31% of all fungi found there. In the Niepołomice Forest 33 species of mycorrhizal fungi were recorded, which makes as little as 12% of all fungi found there.

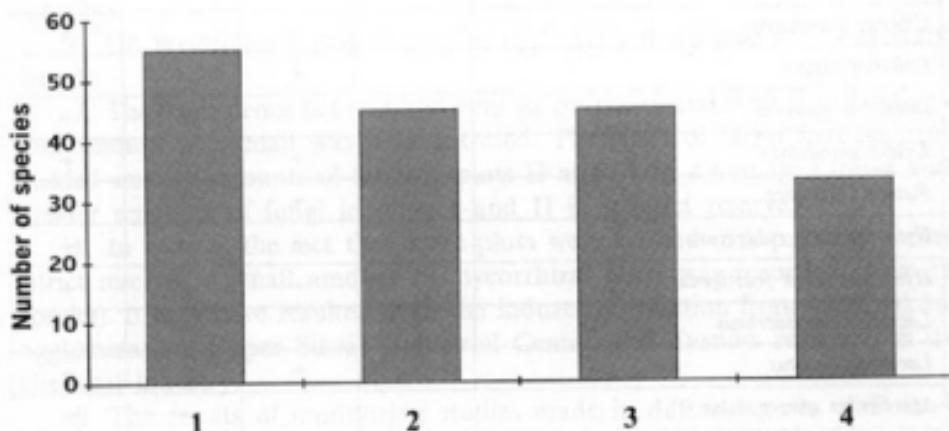


Fig. 13. Mycological monitoring in oak forests in Poland during the years 1994–1996. Comparison of mycorrhizal fungi in various localities: 1 – the Świętokrzyskie Mts., 2 – Wielkopolska, 3 – the Białowieża National Park, 4 – the Niepołomice Forest

Fungi connected with oak. Considering the topic of the present study, the authors were especially focused on the fungi connected with oak. Although oak occurred in the all study plots in the Niepołomice Forest, only 15 species connected exclusively or mainly with oak were recorded. Among the species listed here only two mycorrhizal fungi were found: *Lactarius quietus* and *Xerocomus rubellus*. The other fungi connected with

oak are mainly saprobic fungi, 2 are parasitic fungi: *Fistulina hepatica* and *Phellinus robustus*.

In the Białowieża National Park (Skirgiello 1998b) four species connected with oak were recorded. In Wielkopolska (Lisiewska and Połczyńska 1998) ten such species were found. In the Świętokrzyskie Mountains Łuszczynski (1998) listed 33 species connected with oak. Some of them (22) are not exclusively connected with oak, they can form a mycorrhiza with different tree species or they grow on wood of a variety of trees and shrubs. Only eleven species may be considered oak-associated. Only two species, *Hymenochaete rubiginosa* and *Lactarius quietus* were collected in the all monitoring plots in Poland (Table 10).

Table 10

List of species connected with oak on various investigated plots in Poland
 BNP — the Białowieża National Park (Skirgiello 1998b), W — Wielkopolska
 (Lisiewska and Połczyńska 1998), GS — the Świętokrzyskie Mts. (Łuszczynski 1998), NF — the Niepolomice Forest

Species	Regions			
	BNP	W	GS	NF
<i>Ciboria batschiana</i>		+		+
<i>Collybia fusipes</i>		+	+	+
<i>Daedalea quercina</i>			+	+
<i>Exidia glandulosa</i>		+	+	+
<i>Fistulina hepatica</i>		+	+	+
<i>Hymenochaete rubiginosa</i>	+	+	+	+
<i>Hymenoscyphus fructigenus</i>	+	+		+
<i>Lactarius chrysorrheus</i>			+	
<i>Lactarius quietus</i>	+	+	+	+
<i>Marasmius quercophilus</i>				+
<i>Mycena inclinata</i>		+	+	+
<i>Peniophora quercina</i>		+	+	+
<i>Phellinus robustus</i>			+	+
<i>Stereum gausapatum</i>				+
<i>Xerocomus rubellus</i>		+	+	+
<i>Xylobolus frustulatus</i>	+			+
Total: 16	4	10	11	15

SUMMARY OF RESULTS AND CONCLUSIONS

1. The present monitoring studies were carried out in the years 1994–1996 in four plots – each plot 1000 m² in size and divided into 10 subplots of 100 m² – in the *Tilio-Carpinetum stachygetosum* association in the Niepołomice Forest near Kraków (the Sandomierz Basin). As many as 26 observations were performed.

2. Altogether 274 species and one variety, including 16 species of *Ascomycota* and 258 species of *Basidiomycota* were recorded, 16 species are new to Poland (Table 11).

3. The applied method of detailed observations performed in small subplots, made it possible to sample many species with tiny fructifications which otherwise could have been easily overlooked, among other things, 13 species new to Poland and a few very rare species which have not been recorded from Poland for over 100 year.

4. The occurrence of 173 lignicolous, 51 terrestrial, and 50 species of litter-decomposing fungi was recorded. Four species of mycophilous fungi and four species of bryophilous fungi were assigned to lignicolous fungi.

5. On all plots 234 saprobic, 33 mycorrhizal, and 7 parasitic fungi were recorded.

6. On investigated area 15 species exclusively connected with oak were found.

7. The dependence between the number of species and their fructifications and amount of rainfall was demonstrated. The effect of forest management yielded smaller amounts of fungi in plots II and IV in a managed forest and greater amounts of fungi in plots I and II in a strict reserve.

8. In spite of the fact that some plots were located in a primeval forest (strict reserve), a small amount of mycorrhizal fungi was recorded (only 33 species). It may have resulted from the industrial pollution from the Kraków agglomeration, Upper Silesia Industrial Center, and Tarnów region with its chemical industry.

9. The results of monitoring studies made in different parts of Poland were compared. The smallest proportion of mycorrhizal fungi was recorded in the Niepołomice Forest: 12%, in Wielkopolska: 20%, in the Białowieża National Park: 21%, and in the Świętokrzyskie Mountains: 31%.

10. The studied plots of the Niepołomice Forest are located in the most polluted, heavily industrialized and densely populated region of Poland. This situation is undoubtedly affecting the specific composition of forest fungi. Saprobic fungi are relatively resistant to such a strong anthropopression, while mycorrhizal fungi much less so.

Table 11

List of species (in alphabetical order) collected during the three-year research (1994–1996)
 f – fungi, l – litter, m – mosses, M – mycorrhizal fungi, P – parasitic fungi, s – soil,
 S – saprobic fungi, w – wood

Species	Trophic Groups	Substrate
l	2	3
<i>Agaricus sylvicola</i> (Vitt.) Peck ss. auct. = <i>A. essetei</i> Bon = <i>A. abruptibulbus</i> Peck ss. auct. Europ., non Peck 1905	S	s
<i>Agrocybe praecox</i> (Pers.: Fr.) Fayod	S	s
<i>Amanita fulva</i> Sing.	M	s
<i>Amanita vaginata</i> (Bull.: Fr.) Vitt.	M	s
<i>Armillaria mellea</i> (Vahl: Fr.) Kumm. s. l.	S	w
<i>Ascocoryne sarcoides</i> (Jacq.: Fr.) Groves et Wilson	S	w
<i>Athelia acrospora</i> JüL.	S	w
<i>Athelia alnicola</i> (Bourd. et Galz.) JüL.	S	w
<i>Athelia decipiens</i> (Höhn. et Litsch.) J. Erikss.	S	w
<i>Athelia epiphylla</i> Pers.	S	w
<i>Athelia salicis</i> Pers.	S	w
<i>Basidiobolus caesiocinereum</i> (Höhn. et Litsch.) Luck-Allen	S	w
<i>Basidiobolus radula</i> (Fr.: Fr.) Nobles = <i>Hyphoderma radula</i> (Fr.: Fr.) Donk	S	w
<i>Bjerkandera adusta</i> (Willd.: Fr.) Karst.	S	w
<i>Boletus pulverulentus</i> Opat.	M	s
<i>Botryobasidium conspersum</i> J. Erikss.	S	w
<i>Botryobasidium laeve</i> (J. Erikss.) Parm.	S	w
<i>Botryobasidium subcoronatum</i> (Höhn. et Litsch.) Donk	S	w
<i>Brevicellicium olivascens</i> (Bres.) Larsson et Hjortst.	S	w
<i>Bulgaria inquinans</i> (Pers.: Fr.) = <i>B. polymorpha</i> (Lightfoot) Wettst.	S	w
<i>Byssomerulius corium</i> (Pers.: Fr.) Parm. = <i>Meruliodiplosis corium</i> (Pers.: Fr.) Gianni	S	w
<i>Calocera cornea</i> (Batsch: Fr.) Fr.	S	w
<i>Calocybe carneola</i> (Bull.: Fr.) Kumm.	S	l
<i>Ciboria batschiana</i> (Zopf in Zopf et Sydow) Buchw.	S	w
<i>Clavulinopsis cinerea</i> (Bull.: Fr.) Schroet.	S	s
<i>Clavulinopsis coraloides</i> (L.: Fr.) Schroet. = <i>C. cristata</i> (Holmsk.: Fr.) Schroet.	S	s
<i>Clitocybe candicans</i> (Pers.: Fr.) Kumm.	S	l
<i>Clitocybe clavipes</i> (Pers.: Fr.) Kumm.	S	l

<i>Clitocybe gibba</i> (Pers.: Fr.) Kumm. = <i>C. infundibuliformis</i> (Schaeff.) Quél.	S	I
<i>Clitocybe hydrogramma</i> (Bull.: Fr.) Kumm. = <i>C. phaeophthalma</i> (Pers.) Kuyper	S	I
<i>Clitocybe lignatilis</i> (Pers.: Fr.) Karst. = <i>Ossicaulis lignatilis</i> (Pers.: Fr.) Redhead et Ginns	S	w
<i>Clitocybe metachroa</i> (Fr.) Kumm. ss. Kuyper	S	I
<i>Clitocybe odora</i> (Bull.: Fr.) Kumm. = <i>Lepista odora</i> (Bull.: Fr.) Harmaja	S	I
<i>Clitocybe</i> sp.	S	I
<i>Collybia butyracea</i> (Bull.: Fr.) Kumm. var. <i>asema</i> (Fr.: Fr.) Quél. = <i>C. asema</i> (Fr.: Fr.) Kumm. = <i>Rhodocollybia butyracea</i> f. <i>asema</i> (Fr.: Fr.) Antonin et al.	S	I
<i>Collybia dryophila</i> (Bull.: Fr.) Kumm. = <i>Gymnopus dryophilus</i> (Bull.: Fr.) Murrill	S	I
<i>Collybia erythropus</i> (Pers.: Fr.) Kumm. = <i>C. marasmioides</i> (Britz.) Bresinsky et Stangl = <i>Gymnopus erythropus</i> (Pers.: Fr.) Antonin et al.	S	w
<i>Collybia fuscopurpurea</i> (Pers.: Fr.) Kumm. = <i>Gymnopus fuscopurpureus</i> (Pers.: Fr.) Antonin et al.	S	I
<i>Collybia fusipes</i> (Bull.: Fr.) Quél. = <i>Gymnopus fusipes</i> (Bull.: Fr.) S. F. Gray	S	w
<i>Collybia peronata</i> (Bolt.: Fr.) Kumm. = <i>Gymnopus peronatus</i> (Bolt.: Fr.) Antonin et al.	S	I
<i>Coniophora arida</i> (Fr.) Karst.	S	w
<i>Coniophora puteana</i> (Schum.: Fr.) Karst.	S	w
<i>Conocybe semiglobata</i> (Kühner) ex Kühner et Wall.	S	s
<i>Conocybe tenera</i> (Schaeff.: Fr.) Kumm.	S	s
<i>Coprinus micaceus</i> (Bull.: Fr.) Fr.	S	w
<i>Coprinus</i> sp.	S	s
<i>Cortinarius</i> cfr. <i>abietorum</i> (Vell.) Moser	M	s
<i>Cortinarius</i> sp.	M	s
<i>Crepidotus variabilis</i> (Pers.: Fr.) Kumm.	S	w
<i>Cyathus striatus</i> (Huds.) Willd.: Pers.	S	w
<i>Cylindrobasidium evolvens</i> (Fr.: Fr.) Jülich. = <i>C. laeve</i> (Pers.: Fr.) Chamuris	S	w
<i>Dacryomyces minor</i> Peck	S	w
<i>Dacryomyces stillatus</i> Nees: Fr.	S	w
<i>Dacryomyces tortus</i> (Willd.): Fr.	S	w
<i>Daedalea quercina</i> (L.) Pers.	S	w
<i>Delicatula integrella</i> (Pers.: Fr.) Fayod	S	I
<i>Dentipellis fragilis</i> (Pers.: Fr.) Donk	S	w
<i>Entoloma</i> sp.	M	s

Tab. 11 cont.

1	2	3
<i>Exidia glandulosa</i> (Bull.): Fr. = <i>E. truncata</i> Fr.	S	w
<i>Exidia plana</i> (Wiggers) Donk	S	w
<i>Exidia</i> sp.	S	w
<i>Fistulina hepatica</i> (Schaeff.): Fr.	P	w
<i>Fomes fomentarius</i> (L.: Fr.) Fr.	S	w
<i>Galerina atkinsoniana</i> A. H. Smith	S	w(m)
<i>Galerina cerina</i> Smith et Sing.	S	w(m)
<i>Galerina fallax</i> Smith et Sing.	S	w(m)
<i>Galerina marginata</i> (Batsch) Kühn.	S	w
<i>Galerina triscopa</i> (Fr.) Kühn.	S	w
<i>Ganoderma lipsiense</i> (Batsch) Atk. = <i>G. applanatum</i> (Pers.) Pat.	S	w
<i>Gymnopilus stabilities</i> (Weinm.) Kühn. et Romagn.	S	w
<i>Hapalopilus rutilans</i> (Pers.: Fr.) Karst. = <i>H. nidulans</i> (Fr.) Karst.	S	w
<i>Hebeloma claviceps</i> (Fr.) Kumm. = <i>H. crustuliniforme</i> (Bull.) Quél. ss. Bruchet	M	s
<i>Hebeloma clavulipes</i> Romagn.	M	s
<i>Hebeloma sacchariolens</i> Quél.	M	s
<i>Hohenbuehelia myxotricha</i> (Lév.) Sing. = <i>H. fluxilis</i> (Fr.) Orton ss. auct.	S	w
<i>Hymenochaete cinnamomea</i> (Pers.: Fr.) Bres.	S	w
<i>Hymenochaete rubiginosa</i> (Dickson: Fr.) Lév.	S	w
<i>Hymenoscyphus calyculus</i> (Sow.: Fr.) Phillips	S	w
<i>Hymenoscyphus fructigenus</i> (Bull.: Fr.) S. F. Gray	S	w
<i>Hyphoderma argillaceum</i> (Bres.) Donk	S	w
<i>Hyphoderma mutatum</i> (Peck) Donk	S	w
<i>Hyphoderma pallidum</i> (Bres.) Donk	S	w
<i>Hyphoderma praetermissum</i> (Karst.) J. Erikss. et Strid.	S	w
<i>Hyphoderma puberum</i> (Fr.) Wallr.	S	w
<i>Hyphoderma setigerum</i> (Fr.) Donk	S	w
<i>Hyphodontia pallidula</i> (Bres.) J. Erikss.	S	w
<i>Hyphodontia quercina</i> (Pers.: Fr.) J. Erikss.	S	w
<i>Hyphodontia sambuci</i> (Pers.) J. Erikss.	S	w
<i>Hypholoma capnoides</i> (Fr.: Fr.) Kumm.	S	w
<i>Hypholoma fasciculare</i> (Huds.: Fr.) Kumm.	S	w
<i>Hypholoma lateritium</i> (Schaeff.: Fr.) Schroet. = <i>H. sublateritium</i> (Fr.) Quél.	S	w

<i>Hypholoma subviride</i> (Berk. et Curt.) Dennis	S	w
<i>Hypochnicium erikssonii</i> Hallenb. et Hjorst. = <i>H. sphaerosporum</i> (Höhn. et Litsch.) J. Erikss. ss. J. Erikss. et Ryv.	S	w
<i>Hypochnicium geogenium</i> (Bres.) J. Erikss.	S	w
<i>Hypoxylon deustum</i> (Hoffm.: Fr.) Grev. = <i>Ustulina deusta</i> (Hoffm.: Fr.) Petrik	S	w
<i>Hypoxylon fuscum</i> (Pers.: Fr.) Fr.	S	w
<i>Hypoxylon howeanum</i> Peck	S	w
<i>Inocybe cookei</i> Bres.	M	s
<i>Inocybe geophylla</i> (Sow.: Fr.) Kumm. var <i>lilacina</i> Gill.	M	s
<i>Inocybe quietiodor</i> Bon	M	s
<i>Inonotus radiatus</i> (Sow.: Fr.) Karst.	S	w
<i>Kuehneromyces mutabilis</i> (Schaeff.: Fr.) Sing. et A. H. Smith	S	w
<i>Laccaria amethystina</i> Cooke	M	s
<i>Laccaria laccata</i> (Scop.: Fr.) Berk. et Br.	M	s
<i>Lactarius camphoratus</i> (Bull.) Fr.	M	s
<i>Lactarius mitissimus</i> (Fr.) Fr.	M	s
<i>Lactarius quietus</i> (Fr.) Fr.	M	s
<i>Lactarius</i> sp.	M	s
<i>Laetiporus sulphureus</i> (Bull.: Fr.) Murrill	P	w
<i>Lepista gilva</i> (Pers.: Fr.) Pat. = <i>Clitocybe gilva</i> (Pers.: Fr.) Kumm.	S	l
<i>Lepista flaccida</i> (Sow.: Fr.) Pat. = <i>L. inversa</i> (Scop.) Pat. = <i>Clitocybe inversa</i> (Scop.) Quéel.	S	l
<i>Lepista irina</i> (Fr.) Bigelow	S	l
<i>Lepista nuda</i> (Bull.: Fr.) Cke.	S	s
<i>Lycoperdon perlatum</i> Pers.: Pers.	S	s
<i>Lycoperdon pyriforme</i> Schaeff.: Pers.	S	w
<i>Macrolepiota procera</i> (Scop.: Fr.) Sing.	S	s
<i>Macrolepiota rhacodes</i> (Vitt.) Sing.	S	s
<i>Marasmiellus ramealis</i> (Bull.: Fr.) Sing.	S	w
<i>Marasmius bulliardii</i> Quéel.	S	l
<i>Marasmius quercophilus</i> Pouz. = <i>M. splachnoides</i> (Hornem.: Fr.) Fr. ss. auct.	S	l
<i>Marasmius rotula</i> (Scop.: Fr.) Fr.	S	l
<i>Marasmius torquescens</i> Quéel.	S	l
<i>Megacollybia platyphylla</i> (Pers.: Fr.) Kotl. et Pouz.	S	w
<i>Mollisia</i> sp.	S	w

Tab. 11 cont.

	1	2	3
<i>Mycena abramsii</i> (Murr.) Murr. = <i>M. praecox</i> Vel.	S	s	
<i>Mycena amicta</i> (Fr.) Quél.	S	s	
<i>Mycena bulbosa</i> (Cejp.) Kühn.	S	l	
<i>Mycena crocata</i> (Schrad.: Fr.) Kumm.	S	l	
<i>Mycena filipes</i> (Bull.: Fr.) Kumm.	S	l	
<i>Mycena galericulata</i> (Scop.: Fr.) Quél.	S	w	
<i>Mycena galopus</i> (Pers.: Fr.) Kumm. var. <i>galopus</i> et var. <i>nigra</i> ss. Rea	S	l	
<i>Mycena haematopus</i> (Pers.: Fr.) Kumm.	S	w	
<i>Mycena inclinata</i> (Fr.) Quél.	S	w	
<i>Mycena leptcephala</i> (Pers.: Fr.) Gill.	S	w	
<i>Mycena mucor</i> (Batsch: Fr.) Gill.	S	l	
<i>Mycena pellanthina</i> (Fr.) Quél.	S	s	
<i>Mycena pullata</i> (Berk. et Cke.) Sacc.	S	l	
<i>Mycena pura</i> (Pers.: Fr.) Kumm.	S	l	
<i>Mycena rorida</i> (Fr.) Quél.	S	l	
<i>Mycena sanguinolenta</i> (Alb. et Schw.: Fr.) Kumm.	S	l, w	
<i>Mycena stylobates</i> (Pers.: Fr.) Kumm.	S	l	
<i>Mycena subcaerulea</i> (Peck) Sacc.	S	w	
<i>Mycena vitilis</i> (Fr.) Quél.	S	l	
<i>Mycena zephyrus</i> (Fr.: Fr.) Kumm.	S	l	
<i>Mycenella salicina</i> (Vel.) Sing.	S	w	
<i>Naucoria bohemica</i> Vel.	M	s	
<i>Naucoria spadicea</i> Reid = <i>N. langei</i> Kühn. = <i>Abnicola langei</i> (Kühn.) Sing.	M	s	
<i>Nectria cinnabarinna</i> (Tode: Fr.) Fr.	S	w	
<i>Nectria</i> sp.	S	w	
<i>Oligoporus fragilis</i> (Fr.) Gilba. et Ryv. = <i>Postia fragilis</i> (Fr.) JüL.	S	w	
<i>Oligoporus leucomallellus</i> (Murr.) Gilba. et Ryv. = <i>Postia leucomallella</i> (Murr.) JüL.	S	w	
<i>Oligoporus subcaesius</i> (David) Ryv. et Gilba. = <i>Postia subcaesia</i> (David) JüL.	S	w	
<i>Omphalina</i> sp.	S	l	
<i>Panellus serotinus</i> (Schrad.: Fr.) Kühn. = <i>Sarcomyxa serotina</i> (Schrad.: Fr.) Karst.	S	w	
<i>Paxillus involutus</i> (Batsch.: Fr.) Fr.	M	s	

<i>Peniophora cinerea</i> (Pers.: Fr.) Cke.	S	w
<i>Peniophora incarnata</i> (Pers.: Fr.) Karst.	S	w
<i>Peniophora laeta</i> (Fr.) Donk	S	w
<i>Peniophora quercina</i> (Pers.: Fr.) Cke.	S	w
<i>Peniophora rufomarginata</i> (Pers.) Litsch. in Keissler	S	w
<i>Peziza varia</i> (Hedw.): Fr.	S	w
<i>Peziza</i> sp.	S	s
<i>Phallus impudicus</i> L.: Pers.	S	s
<i>Phanerochaete filamentosa</i> (Berk. et Curt.) Burds.	S	w
<i>Phanerochaete laevis</i> (Fr.) J. Erikss. et Ryv.	S	w
<i>Phanerochaete martelliana</i> (Bres.) J. Erikss. et Ryv.	S	w
<i>Phanerochaete sanguinea</i> (Fr.) Pouz. = <i>Ph. calotricha</i> (Karst.) J. Erikss.	S	w
<i>Phanerochaete sordida</i> (Karst.) J. Erikss. et Ryv.	S	w
<i>Phanerochaete tuberculata</i> (Karst.) Parm.	S	w
<i>Phanerochaete velutina</i> (DC.: Fr.) Karst.	S	w
<i>Phanerochaete</i> sp.	S	w
<i>Phellinus contiguus</i> (Pers.: Fr.) Fiasson et Niemelä	S	w
<i>Phellinus nigricans</i> (Fr.) Karst. ss. Černý, Vampola (1993) = <i>Ph. igniarius</i> (L.) Quél. s.l.	P	w
<i>Phellinus robustus</i> (Karst.) Bourd. et Galz.	P	w
<i>Phlebia radiata</i> Fr. = <i>Ph. merismoides</i> (Fr.): Fr.	S	w
<i>Phlebia rufa</i> (Pers.: Fr.) Christ.	S	w
<i>Phlebia tremellosa</i> (Schrad.: Fr.) Nakas. et Burds. = <i>Merulius tremellosus</i> Schrad.: Fr.	S	w
<i>Phlebiella pseudotsugae</i> (Burt.) Larrs. et Hjortst.	S	w
<i>Phlebiella vaga</i> (Fr.) Karst.	S	w
<i>Phlebiella</i> sp.	S	w
<i>Phleogena faginea</i> (Fr.: Fr.) Link	S	w
<i>Pholiota flammans</i> (Fr.) Kumm.	S	w
<i>Pholiota lenta</i> (Pers.: Fr.) Sing.	S	l
<i>Physporinus sanguinolentus</i> (Alb. et Schw.: Fr.) Pil.	S	w
<i>Physporinus vitreus</i> (Pers.: Fr.) Karst.	S	w
<i>Pluteus cervinus</i> (Schaeff.) Kumm. = <i>P. atricapillus</i> (Batsch) Fayod	S	w
<i>Pluteus inquilineus</i> Romagn. = <i>P. semibulbatus</i> ss. J. Lge.	S	w
<i>Pluteus leoninus</i> (Schaeff.: Fr.) Kumm.	S	w
<i>Pluteus nanus</i> (Pers.: Fr.) Kumm.	S	w

Tab. 11 cont.

1	2	3
<i>Phlebia pellitus</i> (Pers.: Fr.) Kumm.	S	w
<i>Phlebia phlebophorus</i> (Ditmar: Fr.) Kumm.	S	w
<i>Phlebia salicinus</i> (Pers.: Fr.) Kumm.	S	w
<i>Polyporus badius</i> (S. F. Gray) Schw.	S	w
<i>Polyporus brumalis</i> (Pers.): Fr.	S	w
<i>Polyporus varius</i> (Pers.): Fr.	S	w
<i>Psathyrella candolleana</i> (Fr.: Fr.) Maire	S	w
<i>Psathyrella chondroderma</i> (Berk. et Br.) Karst.	S	l
<i>Psathyrella fulvescens</i> (Romagn.) A. H. Smith	S	w
<i>Psathyrella gossypina</i> (Bull.: Fr.) Pearson et Dennis	S	w
<i>Psathyrella gracilis</i> (Fr.) Quéł.	S	l
<i>Psathyrella impexa</i> (Romagn.) Bon	S	l
<i>Psathyrella obtusata</i> (Fr.) A. H. Smith	S	l
<i>Psathyrella ocellata</i> (Romagn.) Mos.	S	w
<i>Psathyrella panaeoloides</i> (Maire) Arnolds	S	l
<i>Psathyrella piluliformis</i> (Bull.: Fr.) Orton	S	l
<i>Psathyrella prona</i> (Fr.) Gill.	S	l
<i>Psathyrella senex</i> (Peck) A. H. Smith	S	l
<i>Psathyrella spadiceogrisea</i> (Schaeff.) Maire	S	w
<i>Psathyrella spadiceogrisea</i> f. <i>vernalis</i> (Lge.) Mos.	S	w
<i>Pseudoclitocybe cyathiformis</i> (Bull.: Fr.) Sing.	S	w
<i>Psilocybe inquinina</i> (Fr.: Fr.) Bres. var. <i>crobulus</i> (Fr.) Hoiland = <i>Psilocybe crobulus</i> (Fr.) M. Lange ex Sing.	S	l
<i>Psilocybe squamosa</i> (Pers.: Fr.) Orton = <i>Stropharia squamosa</i> (Pers.: Fr.) Quéł.	S	l
<i>Radulomyces confluens</i> (Fr.: Fr.) M. P. Christ. = <i>Cerocorticium confluens</i> (Fr.: Fr.) Jülich et Stalp.	S	w
<i>Radulomyces molaris</i> (Chall.: Fr.) M. P. Christ. = <i>Cerocorticium molare</i> (Chaill.: Fr.) Jülich et Stalp.	S	w
<i>Radulomyces rickii</i> (Bres.) M. P. Christ. = <i>Cerocorticium rickii</i> (Bres.) Boid., Gilles et Hugueney	S	w
<i>Ramaria eumorpha</i> (Karst.) Corner	S	s
<i>Ramicola centunculus</i> (Fr.: Fr.) Watl. = <i>Simocybe centunculus</i> (Fr.) Sing.	S	s
<i>Ramicola haustellaris</i> (Fr.: Fr.) Watl. = <i>R. rubi</i> (Berk.) Watl. = <i>Simocybe rubi</i> (Berk.) Sing.	S	w
<i>Ramicola laevigata</i> (Favre) Watl. = <i>Simocybe laevigata</i> (Favre) Orton	S	l

<i>Rickenella fibula</i> (Bull.: Fr.) Raith. = <i>Gerronema fibula</i> (Bull.: Fr.) Sing. = <i>Omphalina fibula</i> (Bull.: Fr.) Quél.	S	w(m)
<i>Rickenella setipes</i> (Fr.: Fr.) Raith = <i>R. swartzii</i> (Fr.) Kuyper = <i>Omphalina setipes</i> (Fr.: Fr.) Quél.	S	w(m)
<i>Ripartites metrodii</i> Huijsman	S	I
<i>Ripartites tricholoma</i> (Alb. et Schw.: Fr.) Karst.	S	I
<i>Rosellinia aquila</i> (Fr.: Fr.) De Not.	S	w
<i>Russula cyanoxantha</i> (Schaeff.) Fr.	M	s
<i>Russula foetens</i> (Pers.): Fr.	M	s
<i>Russula nigricans</i> Fr.	M	s
<i>Russula ochroleuca</i> Pers.	M	s
<i>Russula pectinatoides</i> Peck	M	s
<i>Russula violeipes</i> Quél.	M	s
<i>Schizophyllum commune</i> Fr.: Fr.	S	w
<i>Schizopora flavigera</i> (Cooke) Ryv.	S	w
<i>Schizopora paradoxo</i> (Schrad.: Fr.) Donk s.l.	S	w
<i>Scleroderma areolatum</i> Ehrenb.	M	s
<i>Scleroderma bovis</i> Fr.	M	s
<i>Scopuloides hydnoides</i> (Cke. et Massee in Cke.) Hjortst. et Ryv. = <i>S. rimosa</i> (Cke.) Jülich	S	w
<i>Sebacina callospora</i> Bourd. et Galz.	S	w(f)
<i>Sebacina incrassans</i> (Pers.: Fr.) Tul.	S	w
<i>Sistotrema brinkmannii</i> (Bres.) J. Erikss.	S	w
<i>Sistotrema oblongisporum</i> M. P. Christ. et K. Hauersl.	S	w
<i>Sistotrema niveoacremeum</i> (Höhn. et Litsch.) J. Erikss.	S	w
<i>Skeletocutis nivea</i> (Jungh.) Keller	S	w
<i>Steccherinum ochraceum</i> (Pers.: Fr.) S. F. Gray	S	w
<i>Stereum gausapatum</i> (Fr.) Fr.	S	w
<i>Stereum hirsutum</i> (Willd.: Fr.) Pers.	S	w
<i>Stereum rameale</i> (Schw.) Burt = <i>S. ochraceoflavum</i> (Schw.) Ell.	S	w
<i>Stereum sanguinolentum</i> (Alb. et Schw.: Fr.) Fr.	S	w
<i>Strobilurus tenacellus</i> (Pers.: Fr.) Sing.	S	I
<i>Stropharia aeruginosa</i> (Curt.: Fr.) Quél.	S	I
<i>Stropharia caerulea</i> Kreisel = <i>S. cyanea</i> (Bolt.) Tuomikoski	S	s, I
<i>Stypella grilletii</i> (Bond.) P. Roberts = <i>Myxarium podlachicum</i> (Bres.) Raithv. in Parmasto	S	w
<i>Subulicystidium longisporum</i> (Pat.) Parm.	S	w

Tab. 11 cont.

1	2	3
<i>Thelephora terrestris</i> Ehrhart ex Willd.: Fr.	M	s
<i>Tomentella fuscoferruginea</i> (Bres.) Litsch.	S	w
<i>Tomentella neobourdotii</i> M. J. Larsen	S	w
<i>Tomentella</i> sp.	S	w
<i>Trametes versicolor</i> (L.: Fr.) Pil.	S	w
<i>Trechispora farinacea</i> (Pers.: Fr.) Liberta s. l.	S	w
<i>Trechispora mollusca</i> (Pers.: Fr.) Liberta	S	w
<i>Tremella encephala</i> Pers.	P	w(f)
<i>Tremella mesenterica</i> Retz.: Fr.	P	w(f)
<i>Tubaria conspersa</i> (Pers.: Fr.) Fayod	S	w
<i>Tubaria furfuracea</i> (Pers.: Fr.) Gill.	S	I
<i>Tulasnella eichleriana</i> Bres.	S	w
<i>Tulasnella thelephorea</i> (Juel) Juel = <i>T. inclusa</i> (M. P. Christ.) Donk	P	w(f)
<i>Vuilleminia comedens</i> (Nees: Fr.) Maire	S	w
<i>Xerocomus chrysenteron</i> (Bull.) Quél. = <i>Boletus pascuus</i> (Pers.) Krombh.	M	s
<i>Xerocomus rubellus</i> (Krombh.) Mos. = <i>Boletus rubellus</i> Krombh.	M	s
<i>Xerocomus subtomentosum</i> (L.: Fr.) Quél. = <i>Boletus subtomentosus</i> L.: Fr.	M	s
<i>Xerula pudens</i> (Pers.) Sing. = <i>Oudemansiella longipes</i> (Quél.) Mos.	S	w
<i>Xerula radicata</i> (Rehb.: Fr.) Dörfelt = <i>Oudemansiella pseudoradicata</i> Mos.	S	w
<i>Xylaria hypoxylon</i> (L.: Fr.) Grev.	S	w
<i>Xylaria longipes</i> Nitschke	S	w
<i>Xylobolus frustulatus</i> (Pers.: Fr.) Boid.	S	w

REFERENCES

- A d a m c z y k B. 1984. Jednostki glebowo-siedliskowe Puszczy Niepołomickiej i ich odporność na antropopresję. Stud. Ośrod. Dokum. Fizjogr. PAN Oddz. Kraków 12: 155–196.
- Antonin V., Noordeloos M. E. 1997. A Monograph of *Marasmius*, *Collybia* and related genera in Europe. Part 2. *Collybia*, *Gymnopus*, *Rhodocollybia*, *Crinipellis*, *Chaetocalathus*, and additions to *Marasmiellus*. Libri Botanici 17: 3–256.
- Bigelow H. E. 1982. North American Species of *Clitocybe*. Part I. Beih. Nova Hedwigia 72: 1–280.
- Bigelow H. E. 1985. North American Species of *Clitocybe*. Part II. Beih. Nova Hedwigia 81: 281–471.
- Bresadola J. 1903. Fungi polonici a cl. Viro B. Eichler lecti. Ann. Mycol. 1: 65–132.
- D o m a n s k i S. 1972. Fungi Polyporaceae I (resupinatae), Mucronoporaceae I (resupinatae). For. Sci. Publ. Departm. Nat. Cent. Sci., Techn. Econ. Inform. Warsaw.

- Domański S., Orłowski H., Skirgielło A. 1973. Fungi Polyporaceae II (pileatae), Mucronoporaceae II (pileatae), Ganodermataceae, Bondarzewiaceae, Boletopsidaceae, Fistulinaceae. For. Sci. Publ. Departm. Nat. Cent. Sci. Techn. Econ. Inform. Warsaw.
- Dzięwolski J. 1976. Zarys historii gospodarki leśnej w Puszczy Niepołomickiej. Studia Naturae Ser. A 13: 101–106.
- Dzięwolski J. 1978. Drzewostany rezerwatu Lipówka w Puszczy Niepołomickiej. Studia Naturae, Ser. A, 17: 119–133.
- Eriksson J., Hjortstam K., Ryvarden L. 1978. The Corticiaceae of North Europe. 5, Fungiflora. Oslo.
- Eriksson J., Hjortstam K., Ryvarden L. 1981. The Corticiaceae of North Europe. 6, Fungiflora. Oslo.
- Eriksson J., Hjortstam K., Ryvarden L. 1984. The Corticiaceae of North Europe. 7, Fungiflora. Oslo.
- Eriksson J., Ryvarden L. 1973. The Corticiaceae of North Europe. 2, Fungiflora. Oslo.
- Eriksson J., Ryvarden L. 1975. The Corticiaceae of North Europe. 3, Fungiflora. Oslo.
- Eriksson J., Ryvarden L. 1976. The Corticiaceae of North Europe. 4, Fungiflora. Oslo.
- Ferchmin M., Medwecka-Kornas A. 1976. Grądy północnej części Puszczy Niepołomickiej. Studia Naturae, Ser. A, 13: 143–169.
- Garścić E. 1996. Obłaskawianie hut. Aura 8: 37.
- Grodzińska K. 1980. Acidification of forest environment (Niepołomice Forest) caused by SO₂ emissions from steel mills. Instytut Bot. Pol. Acad. Sci. Cracow.
- Grodzińska K. 1982. Plant contamination caused by urban and industrial emissions in the region of Cracow city (Southern Poland). In: R. Bornkamm, J. A. Lee, M. R. S. Seaward (eds.), Urban Ecology. 149–160. The Second European Ecological Symposium (1980). Blackwell Sci. Publ. Oxford, London, Edinburgh, Boston, Melbourne.
- Grodzińska K. (ed.). 1994. Monitoring ekologiczny województwa krakowskiego w latach 1986–1992. Bibl. Monitor. Środowiska. Kraków.
- Halling R. E. 1983. The Genus *Collybia* (Agaricales) in the Northeastern United States and adjacent Canada. Mycologia Mem. 8. J. Cramer Publ. Braunschweig.
- Hansen L., Knudsen H. (eds.). 1992. *Polyporales, Boletales, Agaricales, Russulales*. In: Nordic Macromycetes, Vol. 2. Nordsvamp. Copenhagen.
- Hawksworth D. L., Kirk P. M., Sutton B. C., Pegler D. N. 1995. Ainsworth et Bisby's Dictionary of the Fungi. 8 ed. IMI, Univ. Press. Cambridge 404 pp.
- Heim R. 1931. Le genre *Inocybe*. Encyclopédie Mycologique 1. P. Lechevalier et Fils. Paris.
- Jakubowska-Gabara J. 1996. Phytosociological documentation of permanent plots in Poland. In: R. Fellner, M. Ławrynowicz, C. Perini. Mycological monitoring in European oak forest—the pilot project (msc).
- Jülich W. 1984. Die Nichtblätterpilze, Gallertpilze und Bauchpilze. In: H. Gams (ed.), Kleine Kryptogamenflora 2b/1. Basidiomyceten 1. Ed. 1. G. Fischer, Stuttgart—New York.
- Kits Van Waveren E. 1985. The Dutch, French and British species of *Psathyrella*. Persoonia suppl. 2: 1–300.
- Komorowska H. 1980. Mushrooms. In: K. Grodzińska (ed.). Acidification of forest environment (Niepołomice Forest) caused by SO₂ emissions from steel mills. Inst. Bot. Acad. Sci. Cracow.
- Komorowska H. 1986. Tricholomataceae Puszczy Niepołomickiej na tle zmian środowiska. Kraków (msc).
- Komorowska H. 1991 (1988). Tricholomataceae (Agaricales) Puszczy Niepołomickiej. Folia Soc. Sci. Lublin. 30, Biol. 1–2: 55–62.
- Kondracki J. 1978. Geografia fizyczna Polski. PWN. Warszawa.
- Kondracki J. 1994. Geografia Polski. Mezoregiony fizyczno-geograficzne. Wyd. Nauk. PWN. Warszawa.
- Kreisel H. (ed.) 1987. Pilzflora der Deutschen Demokratischen Republik. VEB G. Fischer Verl. Jena.

- Kreisel H. 1979. Zur Taxonomie von *Stropharia aeruginosa* sensu lato. Beih. Sydowia 8: 228–32.
- Kuyper T. W. 1986. A revision of the genus *Inocybe* in Europe. I. Subgenus *Inosperma* and the smooth-spored species if subgenus *Inocybe*. Persoonia suppl. 3. Rijksherbarium. Leiden.
- Lisiewska M. 1965. Udział grzybów wyższych w grądach Wielkopolski. Acta Mycol. 1: 169–268.
- Lisiewska M. 1987. Flora Polska. Grzyby (Mycota) 17: *Mycena*. PWN. Warszawa–Kraków.
- Lisiewska M., Bujakiewicz A. 1976. Grzyby wyższe na tle zespołów leśnych. In: T. Wojterski (ed.) Roślinność rezerwatu „Dębina” pod Wągrowcem w Wielkopolsce. Bad. Fizjogr. nad Polską Zach. 29, B: 119–134.
- Lisiewska M., Połczyńska M. 1998. Changes in macromycetes of the oak-hornbeam forests in the “Dębina” reserve (Northern Wielkopolska). Acta Mycol. 33 (2): 191–230.
- Łuszczynski J. 1998. Macromycetes of the *Potentillo albae-Quercetum* in the Świętokrzyskie Mts. – monitoring studies. Acta Mycol. 33 (2): 231–245.
- Maas Geesteranus R. A. 1992. Mycenes of the Northern Hemisphere. 1: 1–391, 2: 1–493. K. Ned. Akad. Wet., Ser. C, 90.
- Michael E., Hennig B., Kreisel H. 1983–1987. Handbuch für Pilzfreunde, 1–5. VEB G. Fischer Verl. Jena.
- Moser M. 1983. Die Röhrlinge und Blätterpilze (Polyporales, Boletales, Agaricales, Russulales). In: H. Gams (ed.) Kleine Kryptogamenflora IIb/2. Basidiomyceten. 2. Teil. VEB G. Fischer Verl. Jena.
- Myczkowski S. 1981. Lasy grądowe Puszczy Niepołomickiej. Stud. Ośrod. Dok. Fizjogr. PAN Oddz. Kraków 9: 117–130.
- Nespiak A. 1959. Studia nad udziałem grzybów kapeluszowych w zespołach leśnych na terenie Białowieskiego Parku Narodowego. Monogr. Bot. 8: 3–141.
- Reid D. 1974. A monograph of the British *Dacryomycetales*. Trans. Brit. Mycol. Soc. 62: 433–494.
- Roberts P. 1994. Globose and ellipsoid-spored *Tulasnella* species from Devon and Surrey, with a key to the genus in Europe. Mycol. Res. 98 (12): 1431–1452.
- Ryvarden L., Gilbertson R. L. 1993. European Polypores. Part 1. *Abortiporus-Lindtneria*. Syn. Fung. 6. Fungiflora. Oslo.
- Ryvarden L., Gilbertson R. L. 1994. European Polypores. Part 2. *Meripilus-Tyromyces*. Syn. Fung. 7. Fungiflora. Oslo.
- Skirgiello A. 1991. Flora Polska. Grzyby (Mycota) 20: *Russula*. PWN. Warszawa–Kraków.
- Skirgiello A. 1998a. Flora Polska. Grzyby (Mycota) 25: *Lactarius*. PAN IB im. W. Szafera. Kraków.
- Skirgiello A. 1998b. Macromycetes of oak-hornbeam forests in the Białowieża National Park—monitoring studies. Acta Mycol. 33 (2): 171–189.
- Smith A. H. 1947. North American species of *Mycena*. Univ. Mich. Stud. Scient. Ser. 17. Repr.: 1971. Bibliotheca Mycol. 31: i–xviii + 1–521. Verl. J. Cramer. Lehre.
- Smith A. H., Singer R. 1964. A monograph on the genus *Galerina* Earle. Hafner Publ. Comp. New York–London.
- Smoliski S. 1981. Zarys przeszłości Puszczy Niepołomickiej. Stud. Ośr. Dokum. Fizjogr. PAN Oddz. Kraków 9: 9–24.
- Stangl J. 1989. Die Gattung *Inocybe* in Bayern. Hoppea, Denkschr. Regensb. Bot. Ges. 46: 1–394.
- Vampola P. 1993. Co je ohrovec obecný? Mykol. Listy, 50: 9–12.
- Vellinga E. C. 1990. *Pluteus* Fr. In: C. Bas et al. (eds.), Flora Agaricina Neerlandica. 2. A. A. Balkema. Rotterdam/Brookfield: 31–55.

- Watling R. 1982. *Bolbitiaceae: Agrocybe, Bolbitius and Conocybe*. In: D. M. Henderson et al. (eds.), British Fungus Flora. 3: 1–139.
- Watling R., Gregory N. M. 1987. *Strophariaceae and Coprinaceae* p.p. In: D. M. Henderson et al. (eds.), British Fungus Flora. 5: 1–121.
- Watling R., Gregory N. M. 1989. *Crepidotaceae, Pleurotaceae and other pleurotoid agarics*. In: D. M. Henderson et al. (eds.), British Fungus Flora. 6: 1–157.
- Watling R., Gregory N. M., Orton P. D. 1993. *Cortinariaceae* p.p. In: D. M. Henderson et al. (eds.), British Fungus Flora. 7: 1–131.
- Watling R., Taylor M. 1987. Observations on the *Bolbitiaceae*. 27. *Bibliotheca Mycol.* 117: 1–61 + 15 figs. + 1 pl.
- Wojewoda W. 1977. Flora Polska. Grzyby (Mycota) 8: *Auriculariales, Septobasidiales, Tremellales*. PWN. Warszawa – Kraków.
- Wojewoda W. 1978. Grzyby wielkoowocnikowe rezerwatu Lipówka w Puszczy Niepołomickiej. *Studio Naturae*, Ser. A, 19: 159.
- Wojewoda W. 1979. Rozmieszczenie geograficzne grzybów tremelloidalnych w Polsce. *Acta Mycol.* 15 (1): 75–144.
- Wojewoda W., Komorowska H. 1997. Notes on *Phleogena faginea* (Fungi, Atractiellales). *Fragn. Flor. Geobot.* 42 (1): 153–160.
- Wojewoda W., Ławrynowicz M. 1992. Red list of threatened macrofungi in Poland. In: K. Zarzycki, W. Wojewoda, Z. Heinrich (eds.). List of threatened plants in Poland. 2 ed.: 27–56. Pol. Acad. of Sci. W. Szafer Inst. of Bot. Kraków.

Macromycetes lasu dębowo-lipowo-grabowego w Puszczy Niepołomickiej – badania monitoringowe

Streszczenie

W latach 1994–1996, w ramach międzynarodowego programu „Mycological monitoring in European oak forests”, którego koordynatorem w Polsce była prof. M. Ławrynowicz, przeprowadzono badania grzybów wielkoowocnikowych w Puszczy Niepołomickiej k. Krakowa. Badania wykonano w lesie należącym do zbiorowiska *Tilio-Carpinetum stachytesum* (ryc. 1, 4, 5), na czterech powierzchniach badawczych (każda o wymiarach 1000 m²), podzielonych na 10 podpowierzchni (każda o wymiarach 100 m²), na dwóch stanowiskach (w oddziałach leśnych 447 i 460).

Obserwacje (w liczbie 26) prowadzono od wiosny do jesieni, raz lub dwa razy w miesiącu. Na każdej podpowierzchni notowano: liczbę owocników poszczególnych gatunków oraz określano ich podłożę i trofizm.

Stwierdzono 274 gatunki, w tym 16 gatunków z *Ascomycota* i 158 z *Basidiomycota*. Zastosowana metoda bardzo dokładnych obserwacji na małych podpowierzchniach, pozwoliła na znalezienie wielu gatunków o drobnych owocnikach, które łatwo można przeoczyć. Stwierdzono m. in. 16 gatunków nowych dla Polski i kilka bardzo rzadkich, nie notowanych w Polsce od ok. 100 lat. W zebranym materiale 173 gatunki grzybów występowaly na drewnie, 51 na ziemi oraz 50 na ściółce (ryc. 7). Do grzybów nadziennych zaliczono 4 gatunki grzybów mikofilnych i 5 briofilnych pośrednio rosnących na drewnie (uproszczenie to ułatwiło graficzne przedstawienie uzyskanych wyników). Zebrano 234 gatunki grzybów saprobowych, 33 mikoryzowych i 7 pasożytniczych (ryc. 9). Znaleziono 15 gatunków związanych wyłącznie lub głównie z dębem. Wykazano zależność liczby gatunków od opadów atmosferycznych (ryc. 11, 12). Wpływ gospodarki leśnej zaznaczył się mniejszą liczbą grzybów w lesie zagospodarowanym (na powierzchniach III i IV), a większą w rezerwacie ścisłym Lipówka (na powierzchniach I i II).

Wymieniona liczba 33 gatunków grzybów mikoryzowych (ryc. 9, 10), jest bardzo mała, co można tłumaczyć wpływem zanieczyszczeń przemysłowych aglomeracji Krakowa, Górnosłąskiego Okręgu Przemysłowego i Tarnowa z przemysłem chemicznym, bliskim sąsiedztwem ruchliwej szosy i zbieraniem grzybów do celów konsumpcyjnych. Na tak silną antropopresję stosunkowo odporne są grzyby saprobowe.

Porównanie wyników monitoringu z różnych regionów Polski, w tym samym okresie (1994–1996) wykazało najmniejszy udział grzybów mikoryzowych w Puszczy Niepołomickiej – 12% wszystkich stwierdzonych gatunków, w Wielkopolsce – 20% (Lisiewska i Polczińska 1998), w Białowieskim Parku Narodowym – 21% (Skirgielko 1998b), a w Górach Świętokrzyskich – 31% (Luszczynski 1998).