ACTA MYCOLOGICA Vol. 34 (1): 115-123 1999

Growth responses of maritime sand dune plant species to arbuscular mycorrhizal fungi

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Tadych M., Błaszkowski J. Growth responses of maritime sand dune plant species to arbuscular mycorrhizal fungi. Acta Mycol. 34 (1): 115-123, 1999

In a por experiment conducted in a greenhouse, the response of 6 plant spacies dominating in the succession of vegetation of a definition bellow of the Loba Bar to inoculation with arbuscular mycorrhizal fung (AMF) was investigated. The inoculum was a mixture of soil, roots and spores of 5 species of AMF with the dominant spocies Gloma engregareame. Except for Compression consecutes and Fernance raders subsept, memoria, both the growth and the dry matter of above-ground consecutes and Fernance raders subsept, memoria, both the growth and the dry matter of above-ground arrichater and 1, a fulficus inoculated with AMF were higher than those growing in soils tacking infection propagates of these fungi, Inoculation with AMF decreased the dry matter of root: shoot ratios in 5 plants spoise. This property was not determined in Fenance raders subsept, armania due to the death of all control plants. The level of mycorrhizal infection was low and did not correlate with the growth reaction of Jenance spot. AMF found in this study suggests that the opinion of non-mycotrophy or low dependence of plants of Americance on formation of morocontrates.

Key words: arbuscular mycorrhiza, dune plants, Glomales, mycorrhizal dependence.

INTRODUCTION

About 80% of plants of the Earth associate with arbuscular mycorrhizal fungi (AMF) of the order Glomales (Zygomycota; Gianinazzinazzinad Gianinazzi-Pearson 1986). Sites especially favouring AMF are dune soils, mainly due to their low content of phosphorous (Błaszkowski 1995): Koske 1987).

Plants associated with AMF are known to have enhanced inorganic nutrition (C o o p e r 1984), greater rates of photosynthesis (A l l e n et al. 1981) and improved resistance to drought (N e l s o n 1987) and pathogens (S c h ô n b c c k 1978). Inoculation of plants with AMF increased their growth and vigour (P f l e g er and L in d er m an 1994; S m i t h and R c a d 1997). However, most of the investigations dealt with cultivated plants, whose species and even cultivars of the same species differed in their response to mycorrhizal infections (A z c o n and O c a m p o 1981; S a i f 1987). Of dune plants, responses to inoculation with AMF have been recognized in only 9 species, including _dmmophila _amenia_A _breviligation and 7 plant species colonizing tropical dunes of the Gulf of Mexico (C orkida) and 7 plant species colonizing tropical dunes of the Gulf of Mexico (C orkida) and R os ke 1989, 1997; G em_m and K os ke 1989, 1997;

Nicolson and Johnston 1979).

The unique areas of the Leba Bar are deflation hollows (O s t r o w s k i and S y m o n i d e s 1994). The consistent occurrence of AM in plants dominating in the succession in these hollows (T a d y c h and B f a s z k o w s k i in press; B 1 a s z k o w s k i 1995) suggests that the plants are decendent on AMF. However, the decree of their decendence is unknown.

The soils of initial successional stages of vegetation of the Lebs Bar delation hollows either have no or contain very low numbers of infection propagules of AMF (T a d y c h and B l a s z k o w s k i in press). Therefore, recognition of the degree of dependence of plants initiating and dominating in the succession of the deflation hollows may help in their stabilization through either planting non-mycorrhizal plants, plants of a low mycorrhizal dependence or introduction of AMF.

The aim of this paper was to determine the influence of a mixture of AMF on growth of 6 plant species dominating in the succession of vegetation of the 12 deflation hollow of the Łeba Bar.

MATERIALS AND METHODS

The experiment was conducted in a greenhouse of the Agricultural Academy in Szezecin in 1996. Pots of a diameter of 7 cm and a height of 8 cm were filled with a mixture of soil and roots coming from a many-species pot culture of AMF whose host plant was Plantago lancoolata. The culture was established based on a mixture of soil samples collected from the 12 deltation hollow considered. The spores abundance in 10 g dry soil of the culture was 739. The spores represented 5 species of AMF, i.e., Acadiospore Aoskel Blaszk, an undescribed Glomus 107 and Szuetlespore persica (Koske of Walker) Walker et Sanders. The fungus dominating in the inocultum was G. aggregatum. Control pots were filled with the same mixture of soil and roots earlier autoclaved 3 times for 2 h at 24-h intervals. To equalize non-mycorrhizal microflora, 50 ml of filtered washing from a living culture was added to each control pot.

About 20—50 seeds of Agrostis stolonifera, Ammophila arenaria, Corymentor canescens, Festuca rubra subsp. arenaria, Juncus articulatus and J. balticus were seeded at a depth of 0.3—0.5 cm. After 2 weeks, the plant density in each pot was thinned to 5, remaining seedlings of equal height. Plants were watered 3 times a week, using 10 ml of tap water per pot. Plants were harvested 25 weeks after sowing.

The growth rate of plants was measured at 2-week intervals beginning 11 weeks after sowing. After harvest, shoots and roots were dried at 105°C for 16 h and then weighted. Levels of mycorrhizal infections were determined based on 1-1.5 cm root fragments stained in 0.05% trypan blue (Phillips and H a y m a n 1970). The percent length of roots with arbuscules, vesicles and intramatrical hyphae was calculated on the basis of 50 root fragments examined under a light microscope (Mc G o n i g l e et al. 1990).

RESULTS

Except for Corynephorus canescens and Festuca rubra subsp. arenaria, incuclation with AMF of the other four plant species increased both their growth rate in the study period and height determined in the last term of measurement [Fig. 1]. The highest differences in the growth between mycorrhizal and non-mycorrhizal plants appeared in Agrostis stolonifera, Juncus articulatus and 1, balticus. The lower average beight of plants of Corynephorus conescens found beginning from the 7th measurement resulted from the death of some plants. The height and rate of growth of Festuca rubras subsp. arenarias were not determined due to the death of control plants before the first term of measurement.

Dry weight of shoots. The highest increases in dry weight obotos (Fig. 2) were found in mycorrhizal Juneus articulatus (by 5533.33%), Aerostis stolanifera (1610.64%) and Juneus balticus (1040.00%).

Dry weight of roots. Compared with control plants (Fig. 3), inoculation with AMF decreased the dry weight of roots of Corynephorus canescens (by 82.25%), Ammophila arenaria (49.51%) and Agrostis stolonifers (22.51%), but increased it in Juneus articulatus (35.65%) and J. balticus (144.69%). The dry weights of shoots and roots of Festuca rubra subsp. arenaria growing in pots without AMF were not determined due to the death of plants.

Dry matter of root: shoot ratio. Inoculation with AMF most decreased the dry matter of root: shoot ratio (Fig. 4) in Agrastis stolonifera (1:2208) and Inneus articulatus (1:8.05). Lower decreases of this parameter occurred in J. balticus (1:4.67), Corynephorus canescens (1:3.68) and Innunohila arearai (1:226).

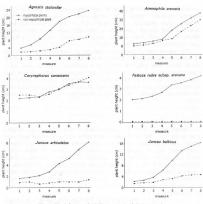


Fig. 1. The influence of AMF on growth of 6 plant species

Arbuscular mycorrhizae

A r b u s c u l e s. The highest values of the percent of root length with arbuscules (Fig. 5) were found in *Anunophila arenaria* and *Juncus articulatus*, and lowest in *Corvnephorus canescens* and *Juncus balticus*.

and lowest in Corynephorus canescens and Juncus outness.

V e s i e l e s. The highest values of root colonization by vesicles (Fig. 5) were found in Agrostis stolonifera, Ammophila arenaria and Juncus articulatus.

Only control plants of Juncus articulatus harboured a very low level of colonization by vesicles.

H y p h a e. Plants having the highest root infections with intramatrical hyphae (Fig. 5) were Juncus articulatus and Anmophila arenaria. Very low infections with intramatrical hyphae were found in Juncus articulatus coming from control plots.

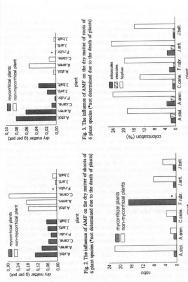


Fig. 4. The influence of AMF on the dry matter shoot:root ratio of 6 plant species (*not determined due to the death of plants)

Percent

Fig.

DISCUSSION

The increased growth and content of dry matter of above-ground plant parts found in this study in most of the dune plant species inoculated with a mixture of AMF corresponds with many literature reports of the positive influence of these fungi on plant condition (S m i t h and R e a d 1997). However, of the plant species investigated here, an increased growth and survival following inoculation with AMF has earlier been revealed only in Ammophila arenaria (Nicolson and Johnston 1979). A similar effect has been observed in the closely related species Ammonbila breviligulata (Gemma and Koske 1989, 1997).

The high positive growth response of Juncus articulatus and J. balticus to inoculation with AMF found in this study was surprised, because members of Juncaceae are considered to form rarely AM (Harley and Harley 1987a, 1987b, 1990). Members of Juncaceae essentially inhabit wet sites. Such wet habitats are generally known to inhibit the formation of AM (S m i t h and Read 1997), although J. conglomeratus growing in periodically submerged dune soils has been found to harbour both extent AM and numerous spores of AMF (Błaszkowski 1993a, 1994; Błaszk o w s k i unpubl.). K o s k e et al. (1985) showed that the infection of Equisetum species, plants commonly considered to be non-mycorrhizal, by AMF depended on associated plants and/or soil moisture conditions. Hence, the lack of colonization with AMF in species of Juncaceae mentioned in the literature apparently expressed the influence of high moisture on the mycorrhizal development rather than the mycotrophic potential of Juncaceae.

The early death of control plants of Festuca rubra subsp. arenaria suggests it to be an obligate mycotroph. No literature data exist of the mycorrhizal status of this plant species

The decreased root: shoot dry weight ratios found in most mycorrhizal plants investigated here corresponds with results of many earlier studies (e.g., Corkidi and Rincón 1997: Saif 1987) Lower root: shoot ratios of mycorrhizal plants have been attributed to the ability of AMF to substitute for the relatively greater amounts of root matter required by non-mycorrhizal plants for phosphorous uptake, allowing mycorrhizal plants to apportion a greater proportion of assimilates to shoot production (Bethlenfalvay, Ulich and Brown 1985).

The level of mycorrhizal infections in AMF-inoculated plants was low. For example, the percent of root length with arbuscules ranged from 0.33% to 3.33% and generally was not correlated with the plant growth responses found. Arbuscules are the main sites of exchange of nutrients between an AM fungus and a host plant (B o n f a n t e-F a s o l o 1984) and, thereby, their presence could be considered the most important measure of the amount of exchange and functionality of an AM symbiosis (Claperton and Reid 1992). The lack of correlation between the level of mycorrhizal infection and the degree of enhancement of plant growth parameters has many times been found in other investigations on mycorrhizal dependence of different cultivated and uncultivated plants (Schubert and Hayman 1986; Hayman and Tavares 1985). Gemma and Koske (1989) found improved growth of Ammophila breviligulata in response to inoculation in the field when colonization extent was as low as 1.5%. Boyetchko and Tewari (1990), Graham et al. (1982) and Hetrick et al. (1985) concluded that plant growth improvement is dependent on a minimum level of root colonization and degree of growth improvement is more correlated with the extent of extramatrical hyphae. Unfortunately, the amount of extramatrical hyphae associated with roots of the plant species investigated here was not determined. According to Saif (1987) and Maniunath and Habte (1991), other important factors affecting mycorrhizal dependence are root fibrosity and root geometry.

This study was supported in part by the Committee for Scientific Research, grant no. 6.P04C.015.13.

REFERENCES

- Allen M. F., Smith W. K., Moore Jr. T. S., Christensen M. 1981.

 Comparative water relations and photosynthesis of mycorrhizal and non-mycorrhizal
- Bouteloua gracilis (H. B. K.) Lag ex Steud. New Phytol. 88: 683-693.

 A z c o n R., O c a m p o J. A. 1981. Factors effecting the vesicular-arbuscular infection and
- Azcón R, Ocampo J. A. 1981. Factors effecting the vesicular-arbuscular infection and mycorrhizal dependency of thirteen wheat cultivars. New Phytol. 87: 677-685.
- Bethlen falvay G. J., Ulich J. M., Brown M. S. 1985. Plant response to mycorrhizal fungi: host, endophyte and soil effects. Soil Sci. Soc. Am. J. 49: 1164–1168. Błaszkow ski J. 1993a. Comparative studies of the occurrence of arbuscular fungi and
- mycorrhizae (Giomales) in cultivated and uncultivated soils of Poland. Acta Mycol. 28: 93-140.

 82 a z k o w k i J 1993b. The occurrence of arbuscular funci and mycorrhizae (Giomales) in
- B ł a s z k o w s k i J. 1993b. The occurrence of arbuscular fungi and mycorrhizae (Glomales) in plant communities of maritime dunes and shores of Poland. Bull. Pol. Ac. Sci. Biol. 41: 377-392.
- Błaszko wski J. 1994. Arbuscular fungi and mycorrhizae (Glomales) of the Hel Peninsula,
- Poland. Mycorrhiza 5: 71-88.

 Błaszkowski J. 1995. Acaulospora koskei, a new species in Glomales from Poland. Mycol.
- Res. 99: 237—240.

 Bonfante-Fasolo P. 1984. Anatomy and morphology of VA mycorrhizae. In: C. L.
 Powell, D. J. Bagyarai (eds.) VA Mycorrhiza. CRS Press, Inc., Boca Raton,
- Powell, D. J. Bagyaraj (eds.) VA Mycorrhiza. CRS Press, Inc., Boca Raton, Florida: 5-33. Boyetchko S. M., Tewari J. P. 1990. Root colonization of different hosts by the
- Boyetch ko S. M., 1 ewari J. P. 1990. Root colonization of dillerent norsts by the vesicular-arbuscular mycorrhizal fungus Glomus dimorphicum. Plant and Soil 129: 131-136. Clapperto M. J., Reid D. M. 1992. A reationship between plant growth and increasing
 - VA mycorrhizal inoculum density. New Phytol. 120: 227-234.

- Cooper K. M. 1984. Physiology of VA mycorrhizal associations. In: C. L. Powell, D. J. Bagyaraj (eds.) VA Mycorrhiza. CRS Press, Inc., BocaRaton, Florida: 155-186. Corkidi L., Rincón E. 1997, Arbuscular mycorrhizae in a tropical sand dune ecosystem on the Gulf of Mexico. II. Effects of arbuscular mycorrhizal fungi on the growth of species
- distributed in different early successional stages. Mycorrhiza 7: 17-23. Gemma J. N., Koske R. E. 1989. Field inoculation of American beachgrass (Ammophila
- breviligulata) with V-A mycorrhizal fungi. J. Environ. Manag. 29: 173-182. G e m m a J. N., K o s k e R. E. 1997. Arbuscular mycorrhizae in sand dune plants of the North
- Atlantic coast of the U.S.: Field and greenhouse inoculation and presence of mycorrhizae in planting stock. J. Environ. Manag. 50: 251-264.
- Gianinazzi S., Gianinazzi-Pearson V. 1986. Progress and headaches in endomycorrhiza biotechnology. Symbiosis 2: 139-149.
- Graham J. L., Linderman R. G., Menge J. A. 1982. Development of external hyphae by differnt isolates of mycorhizal Glomus spp. in relation to root colonization and
- growth of troyer citrange. New Phytol, 91: 183-189. Harley J. L., Harley E. L. 1987a. A check-list of mycorrhiza in the British flora. New
- Phytol. 105: 1-102. Harley J. L., Harley E. L. 1987b. A check-list of mycorrhiza in the British flora
- addenda, errata and index. New Phytol, 107: 741-749. Harley J. L., Harley E. L. 1990. A check-list of mycorrhiza in the British flora - second
- addenda and errata. New Phytol. 115: 699-711. Hayman D. S., Tavares M. 1985. Plant growth responses to vesicular-arbuscular mycorrhiza. XV. Influence of soil pH on the symbiotic efficiency of different endophytes. New
- Phytol. 100: 367-377. Hetrick B. A. D., Bloom J., Feverheim S. M. 1985, Root colonization nattern of
- Glomus epigaeum in nine host species. Mycologia 77: 825-828. Koske R. E., Friese C. F., Olexia P. D., Hauke R. I. 1985. Vesicular-arbuscular
- mycorrhizas in Equisetum, Trans. Br. Mycol. Soc. 85: 350-353. K o s k e R. E. 1987. Distribution of VA mycorrhizal fungi along a latitudinal temperature
- gradient. Mycologia 79: 55-68. Manjunath A., Habte M. 1991. Root morphological characteristics of host species
- having distinct mycorrhizal dependency. Canad. J. Bot. 69: 671-676. McGonigle T.P., Miller M.H., Evans D.G., Fairchild C.L., Swan J.A. 1990. A new method which gives an objective measure of colonization of roots by
- vesicular-arbuscular mycorrhizal fungi. New Phytol. 115: 495-501. Nelson C. E. 1987. The water relations of vesicular-arbuscular mycorrhizal systems. In: G. R. Safir (ed.) Ecophysiology of VA Mycorrhizal Plants. CRS Press, Inc., Boca Raton.
- Florida: 71-92 Nicolson T. H., Johnston C. 1979. Mycorrhiza in the Gramineae III. Glornus fasciculatum as the endophyte of pioneer grasses in a maritime sand dune. Trans. Br. Mycol.
- Soc. 72: 261-268. Ostrowski M., Symonides E. 1994. Słowiński National Park. SCI and ART. Warsaw.
- Pfleger F. L., Linderman R. G. 1994. Mycorrhizae and plant health. APS Press. Phillips J. M., Hayman D. S. 1970. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. Trans. Br. Mycol. Soc. 55: 158-161.
- Saif S. R. 1987. Growth responses of tropical forage plant species to vesicular-arbuscular mycorrhizae. Plant and Soil 97: 25-35.

Schönbeck F. 1978. Einfluss der endotrophen Mykorrhiza auf die Krankheitsresistenz höherer Pflanzen. Z. PflKrankh. PflSchutz. 85: 191—196. Schubert A., Haym an D. S. 1986. Plant growth responses to vesicular-arbuscular

mycorrhiza. XVI. Effectiveness of differnt endophytes at differnt levels of soil phosphate. New Phytol. 103: 79-90.

- Smith S. E., Read D. J. 1997. Mycorhizal symbiosis. Academic Press. Harcourt Brace and Company, Publishers. San Diego, London, New York, Boston, Sydney, Tokyo, Toronto.
 Tad yeh M. Błaszkowski J. Arbusculaf kungi and mycorrhizae (Glomales) of the
 - Słowiński National Park, Poland. Mycotaxon (in press).

Wpływ arbuskularnych grzybów mikoryzowych na wzrost nadmorskich roślin wydmowych

Streszczenie

W doświadczeniu wazonowym przeprowadczonym w uklami zbadano rakcję 6 gatunków profini donimiących w ukceju piod effectjegos Mierze Liebekije a nieculacją patwikanymi grzybami mikoryzowymi (AGM), Inokulum była mieszaniaa głeby, korzeni i zazodników 5 gatunków AGM się gatunkiem dominiejem Glowa agregama. Z wyjakiem Competowa canesceni Fennce rabra subsp. arenaria, zatówno wrost jak i sucha masa części nadziennych AGM były wyżac nia u rożlin romajcych w głebach bez AGM. Inokulega obnicyła sozunek AGM były wyżac nia u rożlin romajcych w głebach bez AGM. Inokulega obnicyła sozunek such jawa pieczenia popodou z gatunków rolini. Wanacio i jak obreżenia prz. Arwar subsp. arenaria z powodu zamarcia rożlin kontrolnych. Voziom inkejej naktowności, rodnostwa z swienia powieczenia prowodu z swienie powieczenia powieczenia powieczenia prowodu z swienie powieczenia powiecz