

## *Cephalotrichum stemonitis* as a biofilm inhabitant in the gold mine in Poland

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*Cephalotrichum stemonitis* and its synanamorph *Echinobotryum atrum* isolated from bacterial biofilm is presented.

**Key words:** mine fungi, *Cephalotrichum*, biofilm, distribution

### INTRODUCTION

Dry-spored synnematosus anamorphs from the form-genus *Cephalotrichum* Link are the asexual states of *Microascaceae*. However so far a teleomorph state is known for no species of this genus (Abbott 2000). *Microascaceae* include five genera: *Microascus*, *Kernia*, *Petriella*, *Pseudoallescheria* and *Lophotrichus* (Abbott et al. 2002) as well as eight anamorphic genera containing about 50 species (Abbott 2000). Abbott (2000) analyzed the subunit 18S rDNA for 34 taxa of *Microascaceae*. He obtained *Microascus* clade consisting of *Microascus longirostris*, *M. nidicola*, *M. cirrhosus*, *M. trigonosporus*, *Cephalotrichum stemonitis*, *Wardomyces anomalus*, *Cephalotrichum cylindricum* and *Microascus brevicaulis*. It indicate that the genus *Microascus* can be recognized as hypothetical teleomorph state for *Cephalotrichum stemonitis*. In GenBank there are available 3 sequences of ribosomal RNA genes (5.8S rRNA, 28S rRNA and large subunit rRNA) of *Doratomyces stemonitis* (in fact *Cephalotrichum stemonitis*) with access numbers: AF400852, EF029213 and AJ608983.

Synnemata of *Cephalotrichum* producing chains of powdery conidia are relatively large with ‘bootle brush’ or “feather” appearance. Ovoid conidia are produced from annelidic conidiogenous cells covering sporogonous area. *Cephalotrichum* synnemata are adapted for dispersion not only by air currents but also by insects (Abbott 2000, 2002). Habitat of *Cephalotrichum* fungi includes soil, compost, wood, wood treated with fungicide, herbaceous stems, oat seeds, decaying plant material and dung, finger nail, sawdust and straw used for growing shiitake, airborne contaminant of wheat-straw agar plate, egg of gypsy moth (*Lymantria dispar*), roots of potato,

basidiomycete detritus, manure pile, bronchial washing, right feel, leaves of needle-leaved tree (Abbott 2000). *C. stemonitis* was noted in coyote and rat dung, indoor air of honeybee (*Apis mellifera*) overwintering facility, cone of white spruce, sandy soil, decayed wood of white spruce, soil of elm woods, agricultural soil. It is also important colonizer of pPVC buried in soil (Sabev et al. 2006). These authors mentioned it as *Doratomyces* spp. in the article, but in GenBank it is more precisely determined as *D. stemonitis*.

Malloch and Hubart (1987) described unnamed species of *Microascus* from Ramoil Cave. It means that fungi from this group can inhabit also underground environments. Fassatiova (1970) noted *C. stemonitis* on wood in uranium mine in Czech Republic.

## STUDY AREA AND METHODS

Gray biofilms of rock-inhabiting bacteria in Gertruda Adit in a closed gold mine located in Złoty Stok in Lower Silesia were chosen for microbial analysis. This mine possesses constant conditions such as low temperature ca 10<sup>o</sup> C, darkness, high humidity, high As concentration and other toxic substances (Chlebicki et al. 2005). So far 12 species of fungi were noted in this mine (Chlebicki et al. 2005; Chlebicki, Lorenc 2006). Bacteria with fungi were collected in sterile plastic tubes and refrigerated at 10<sup>o</sup> C. Fungal growth was performed on DRBC, RBC, YMA and PDA media. Inoculated media on Petri dishes were putted in incubator at 10<sup>o</sup> C. The morphological characters of the living fungi were examined in water and cotton blue in lactophenol using light microscopy (Nikon SMZ 1500, Nikon Labophot 2 and Nikon Eclipse 800). Microphotographs were taken with these microscopes equipped with a digital camera. For scanning electron microscope (SEM) studies mycelium was coated with gold, and photographed using a LEO 1430 VP Zeiss microscope with a working distance of ca 10 mm. Fungus-species nomenclature follows Abbott (2000).

## RESULTS

### *Cephalotrichum stemonitis* (Pers.) Nees

Magazin Ges. naturf. Freunde, Berlin 3: 20 (1809)

Synonymy: *Doratomyces stemonitis* (Pers.) F.J. Morton & G. Sm., Mycol. Pap. 86: 70, 1963

*Doratomyces stemonitis* var. *keratinolyticus* (Dominik & Majchr.) Dominik & Majchr. Ekol. Pol., Ser. A 18: 603, 1970 *Echinobotryum atrum* Corda, Sturm's Deutschl. Flora, III (Pilze) 3(12): 51 (1829) *Periconia stemonitis* Pers., Syn. meth. fung. (Göttingen): 687 (1801) *Stysanus stemonitis* (Pers.) Corda, Icon. fung. (Prague) 1: 22 (1837)

Description: *mycelium* creamy-white to dark brown, after 7 days on PDA 17,5-21 mm diam., on RBC 17,0 mm diam., and on DRBC 13-17 mm, all in room temperature (Fig. 1A, B, D). First conidiophores of synanamorph – *Echinobotryum atrum* – appeared throughout the mycelium after 12 days on PDA. Mycelium growing on RBC after two weeks produced rings of densely distributed synnemata of *C. stemonitis* in central part of the mycelium. Mycelium on PDA formed at the beginning *E.*

*atrum* throughout the surface of dark brown colony and later numerous synnemata of *C. stemonitis*. Mycelium on YMA form brown and irregular colony patches (Fig. 1C). *Synnemata* 190-300 µm long, sterile part 130-210 µm, fertile part 60-120 µm long, (irregular heads) similar to feather (Fig. 2B, C) associated with synanamorph *Echinobotryum atrum* (Fig. 2A), *conidiophores* synnematos, brown, conidiogenous cells ampulliform, percurrent, *conidia* smooth (Fig. 2D), ovoid with pointed apex and truncate base 7-9,1 x 4,2-6,1 µm, pale brown.

Material examined: on rock bacterial biofilm in Gertruda Adit in gold mine in Żłoty Stok, Lower Silesia, Poland, 16 October 2006, coll. A. Chlebicki. First culture obtained on DRBC medium, then transferred to other media.

## DISCUSSION

So far two species of the genus *Cephalotrichum* Link were noted in Poland. Dominik and Majchrowicz (1965) and Dominik (1970) described a new variety *Doratomyces stemonitis* var. *keratinolyticus* (Dominik & Majchr.) Dominik & Majchr. on the basis of specimen isolated from the soil. Conidia of this specimen are something smaller than typical variety, 4-6 x 2.5-4 µm and 'very slightly rough' (Dominik 1970). It resembles *Cephalotrichum microsporum* (Sacc.) P. M. Kirk. Moreover these authors did not mention the presence of *Echinobotryum* synanamorph which is diagnostic character of *Cephalotrichum stemonitis*.

The next species *Cephalotrichum putredinis* (Corda) S. P. Abbott was reported by Dominik (1970) as *Doratomyces albus* (Szilvinyi) Dominik. Unfortunately, these collections are not available for investigation.

Presence of *Cephalotrichum stemonitis* in bacterial biofilm is accidental. However, as indicate informations of Fassatiova (1970) and Malloch and Hubart (1987), such fungi were noted in subterranean environments. Mille-Lindblom (2005), Hogan and Kolter (2002) and Kirkwood (2002) noted mostly antagonistic relation between fungi and bacteria. Fungi were always negatively affected by presence of bacteria. Penetration of fungal hyphae of *Fusarium oxysporum* was not observed where microcolonies of *Pseudomonas* were present, moreover *Pseudomonas* bacteria attached and colonized fungus hyphae (Bolwerk et al. 2003). Also growth of *Cephalotrichum stemonitis* was suppressed by bacteria from the genus *Pseudomonas* isolated from biofilm.

## REFERENCES

- Abbott S. P. 2000. Holomorph studies of the Microasceae (PhD Dissertation). Edmonton, Alberta: Univ. Alberta. 196 pp.
- Abbott S. P. 2002. Insects and other arthropods as agents of vector-dispersal in fungi. [www.precisionenv.com/PDFS/AbbottInsectdispersal.pdf](http://www.precisionenv.com/PDFS/AbbottInsectdispersal.pdf)
- Abbott S. P., Lumley T. C., Sigler L. 2002. Use of holomorph characters to delimit *Microascus nidicola* and *M. soppii* sp. nov., with notes on the genus *Pithoascus*. *Mycologia* 94 (2): 362-369.
- Bolwerk A., Lagopodi A. L., Wijffes A. H. M., Lamers G. E. M., Chin-A-Woeng T. F. C., Lugtenberg B. J. J., Blomberg G. V. 2003. Interaction in the tomato rhizosphere of two *Pseudomonas* biocontrol strains with the phytopathogenic fungus *Fusarium oxysporum* f. sp. *radicis-lycopersici*. *Mol. Plant-Microbe Interact.* 16: 983-993.
- Chlebicki A., Godzik B., Lorenc M. W., Skłodowska A. 2005. Fungi and arsen-tolerant bacteria in the hypogean environment of an ancient gold mine in Lower Silesia SW Poland. *Polish Bot. Stud.* 19: 81-95.

- Chlebicki A., Lorenc M. W. 2006. The troglophile fungus, *Physisporinus vitreus* on a mine wall at Złoty Stok, Poland. Polish. Bot. J. 22: 149–154.
- Dominik T. 1970. Observations of new or noteworthy fungi from region of Szczecin. Zeszyty Naukowe Wyższej Szkoły Rolniczej w Szczecinie 32: 71–108.
- Dominik T., Majchrowicz I. 1965. Second contribution to the knowledge of keratolytic and keratophilic soil fungi in the region of Szczecin. Ekologia Polska 13: 415–447.
- Fassatiowa O. 1970. Micromycetes inhabiting the mines of Příbram (Czechoslovakia). Česka Mykol. 24: 162–165.
- Hogan D. A., Kolter R. 2002. *Pseudomonas-Candida* interactions: and ecological role of virulence factors. Science 296: 2229–2232.
- Kirkwood M. L. 2002. Bacteria-fungi interactions: pathogenesis meets ecology. Trends in Microbiology 10: 397–398.
- Malloch D., Hubart J. M. 1987. An undescribed species of *Microascus* from the Cave of Ramoiul. Canad. J. Bot. 65: 1281–1283.
- Mille-Lindblom C. 2005. Interaction between bacteria and fungi on aquatic detritus-causes and consequences. Acta Universitatis Upsaliensis. Digital comprehensive summaries of Uppsala dissertations from the Faculty of Science and Technology 46. 42 pp., Uppsala.
- Sabev H. A., Handley P. S., Robson G. D. 2006. Fungal colonization of soil-buried plasticized polyvinyl chloride (pPVC) and the impact of incorporated biocides. Microbiology 152: 1731–1739.

### *Cephalotrichum stemionitis* zasiedlający bakteryjny biofilm w kopalni złota w Polsce

#### Streszczenie

Praca zawiera opis grzyba *Cephalotrichum stemionitis* i jego synanamorfy *Echinobotryum atrum* wyizolowanych z bakteryjnego biofilmu z Sztolni Gertrudy w Kopalni Złota w Złotym Stoku. Pierwsza informacja o tym gatunku podana z Polski przez T. Dominika i I. Majchrowicz jest niezbyt dokładna. Diagnostyczną cechą gatunku *C. stemionitis* jest obecność synanamorfy *E. atrum* o czym wymienieni autorzy nie wspominają. Podano również podstawowe dane o siedliskach i ekologii tego gatunku. Izolacja tego rzadkiego gatunku grzyba z bakteryjnego biofilmu nie była dotychczas notowana.

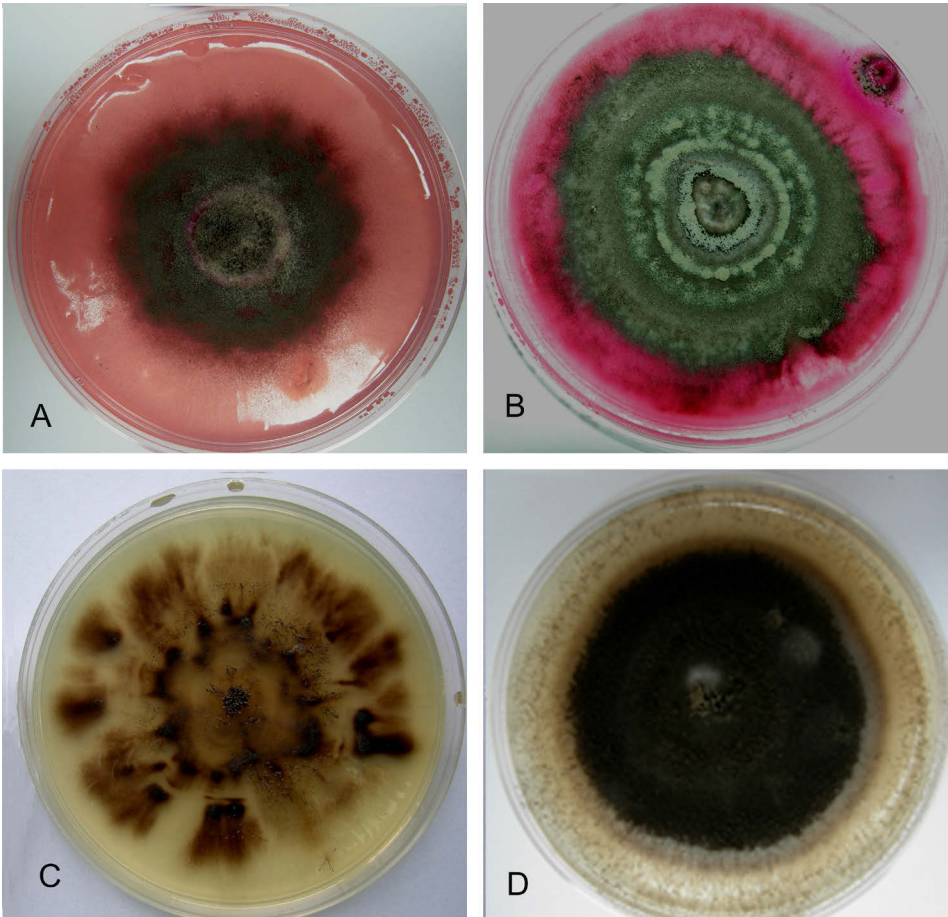


Fig. 1. Colonies of *Cephalotrichum stemonitis* on different media: A – on RBC; B – on DRBC; C – on YMA; D – on PDA.

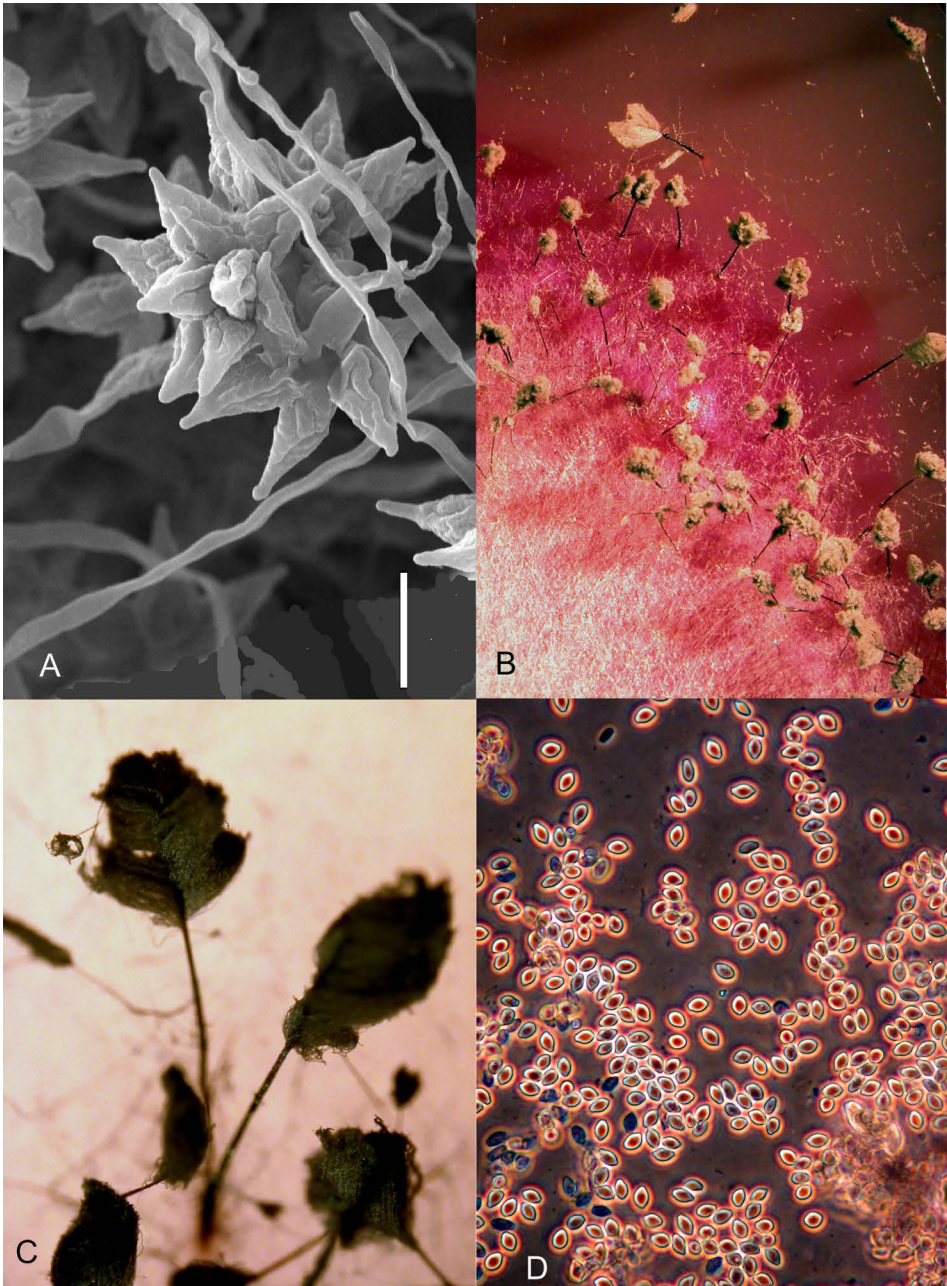


Fig. 2. Morphology of *Cephalotrichum stemonitis* and its synanamorph – *Echinobotryum atrum*: A – conidia of *E. atrum*; scale bar = 10  $\mu\text{m}$ ; B – synnemata of *C. stemonitis* on DRBC medium; C – synnemata of *C. stemonitis* similar to feather; D – conidia of *C. stemonitis*.